

Nutritional strategies for managing joint health in companion animals

2025 Volume 2, Article number: e022

<https://doi.org/10.48130/animadv-0025-0019>

Received: 16 March 2025

Revised: 10 April 2025

Accepted: 15 April 2025

Published online: 28 August 2025

Xinzi Guo¹, Xueqi Yuan², Nisha Farooq³, Yatong Yang⁴, Zhaoxuan Zhu¹ and Lian Li^{4*}

¹ MOE Joint International Research Laboratory of Animal Health and Food Safety, College of Veterinary Medicine, Nanjing Agricultural University, No. 1 Weigang, Nanjing 210095, China

² Hangzhou Lingxian Pet Food Co., Ltd., Building 4, EFC Centre, Cangqian Street, Hangzhou 311100, China

³ Institute of Biotechnology and Genetic Engineering, The University of Agriculture, Peshawar, Pakistan

⁴ College of Animal Science, Nanjing Agricultural University, No. 1 Weigang, Nanjing 210095, China

* Corresponding author, E-mail: lilian@njau.edu.cn

Abstract

Joint health is crucial for the quality of life of companion animals, influencing their mobility, physical activity, and overall well-being. Nutritional strategies play an essential role in mitigating joint degeneration, managing inflammation, and enhancing musculoskeletal resilience across different life stages. This review explores the impact of dietary interventions, including macronutrient balance, bioactive compounds, and specialized supplementation, in maintaining joint integrity and function. Emerging evidence highlights the significance of omega-3 fatty acids, chondroprotective agents, and antioxidant-rich diets in modulating inflammatory pathways, and slowing disease progression. Additionally, tailored nutrition during growth phases is essential to prevent developmental orthopedic diseases, particularly in large-breed dogs. A lifelong, nutrition-centered approach to joint health-integrating dietary modifications with veterinary oversight offers a proactive framework for enhancing mobility, reducing disease burden, and improving the quality of life in companion animals. This review underscores the need for further research into molecular mechanisms linking diet and joint homeostasis, facilitating evidence-based dietary recommendations for veterinary practice.

Citation: Guo X, Yuan X, Farooq N, Yang Y, Zhu Z, et al. 2025. Nutritional strategies for managing joint health in companion animals. *Animal Advances* 2: e022 <https://doi.org/10.48130/animadv-0025-0019>

Introduction

Joint disorders are a significant concern in companion animal health, particularly in aging individuals and large-breed dogs, where both developmental and degenerative joint diseases (DJDs) can severely compromise mobility and overall quality of life^[1]. Among these disorders, osteoarthritis (OA) is the most prevalent, characterized by progressive cartilage degradation, subchondral bone remodeling, synovial inflammation, and oxidative stress-induced damage^[2]. At the molecular level, OA is driven by an imbalance between anabolic and catabolic processes within the extracellular matrix (ECM), primarily regulated by pro-inflammatory cytokines such as interleukin-1 β (IL-1 β), tumor necrosis factor α (TNF- α), and (interleukin-6) IL-6 through activation of the nuclear factor- κ B (NF- κ B) signaling pathway^[3]. Additionally, matrix metalloproteinases (MMPs), especially MMP-3 and MMP-13, exacerbate collagen degradation and impair joint homeostasis^[4]. Given the increasing prevalence of OA in pets and its multifactorial etiology, a comprehensive understanding of risk factors and management strategies is crucial.

Proper nutritional interventions play a vital role in mitigating these degenerative processes by modulating inflammatory pathways, oxidative stress responses, and chondrocyte apoptosis^[5]. Omega-3 fatty acids, including eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), exert anti-inflammatory effects by inhibiting the arachidonic acid cascade, reducing prostaglandin E2 (PGE2) synthesis, and suppressing the NF- κ B pathway, which in turn limits cytokine-induced cartilage degradation^[6]. Glucosamine and chondroitin sulfate, also, have chondroprotective potential and may improve clinical scores

with slower onset than non-steroidal anti-inflammatory drugs (NSAIDs) in canine OA trials^[7]. Moreover, polyphenolic compounds such as resveratrol, curcumin, and epigallocatechin-3-gallate (EGCG) have shown nuclear factor erythroid 2-related factor 2 (NRF2)-linked antioxidative and anti-catabolic effects in chondrocytes and animal OA models, while clinical evidence in companion animals remains preliminary^[8,9].

Beyond DJDs, nutritional imbalances during critical developmental stages can predispose companion animals to lifelong orthopedic disorders. Excess calcium (Ca) intake during rapid growth impairs endochondral ossification and increases risk for developmental orthopedic disorders in large-breed puppies, as shown in controlled Great Dane feeding studies; appropriate Ca-phosphorus (P) balance and energy control are therefore critical^[10]. Dysregulation of the Ihh-PTHrP axis is central to endochondral ossification; nutritional imbalances that disturb mineral homeostasis may secondarily perturb these pathways^[11]. Vitamin D deficiency primarily compromises intestinal Ca and P absorption, causing defective mineralization (rickets/osteomalacia) and skeletal deformities in growing animals; its effects are best framed in mineral homeostasis rather than reduced receptor activator of nuclear factor κ B ligand (RANKL) signaling^[12,13].

This review aims to explore the nutritional strategies for managing joint health, focusing on evidence-based interventions, the role of specific nutrients, and emerging therapies that may enhance joint health outcomes in companion animals. By elucidating these connections, we hope to create awareness and encourage proactive management approaches that ultimately lead to healthier and more active lives for these beloved pets.

Understanding joint health and nutrition

As one of the most prevalent joint disorders in companion animals, OA warrants close attention due to its complex etiology and significant impact on mobility and quality of life. This section explores the fundamental aspects of joint health in companion animals, with an emphasis on OA, joint anatomy, and the influence of nutrition on inflammatory processes.

OA is a type of DJD characterized by the progressive breakdown of cartilage, which cushions the ends of bones in joints^[14]. This deterioration leads to bones rubbing against each other, causing pain, stiffness, and reduced mobility. Current estimates indicate that OA affects approximately 80% of dogs over the age of eight, making it one of the most prevalent chronic conditions in companion animals^[15]. Furthermore, recent studies have identified radiographic evidence of OA—including osteophytes, enthesophytes, subchondral sclerosis, bone erosions, and cysts—in a notable proportion of younger dogs (aged 8 months to 4 years), indicating that OA may be underdiagnosed in early life stages^[16]. This highlights the pressing need for effective management strategies to address joint health issues early in life.

Understanding joint anatomy is essential to comprehending how nutritional choices affect joint health. The primary components of a synovial joint include articular cartilage, synovial fluid, ligaments, and the surrounding joint capsule^[17]. Articular cartilage is composed of chondrocytes and an ECM rich in collagen and proteoglycans, which provide structural integrity and resilience to the joint^[18]. The synovial fluid lubricates the joint, minimizes friction, and serves as a shock absorber. Nutrition plays a critical role in maintaining the health of these structures, with key nutrients such as omega-3 fatty acids, glucosamine, chondroitin sulfate, and antioxidants being vital for cartilage maintenance and the reduction of inflammation^[19–22].

The relationship between nutrition and inflammation is particularly relevant in the context of joint problems such as OA^[23]. Chronic inflammation is a hallmark of OA and contributes significantly to the degeneration of both cartilage and surrounding tissues^[24]. Certain dietary components, such as omega-3 fatty acids, possess anti-inflammatory properties that can modulate the inflammatory response and mitigate the symptoms associated with joint disease^[25]. Conversely, diets high in processed foods, sugar, and unhealthy fats can exacerbate inflammatory processes, leading to the progression of joint conditions^[26]. This underscores the necessity of formulating diets rich in anti-inflammatory nutrients to support joint health and improve the quality of life in affected pets.

In summary, understanding the importance of joint health in companion animals, along with the intricate relationships between OA, joint anatomy, and nutrition, is essential for effective management. A multifaceted approach that includes a tailored diet rich in key nutrients can significantly contribute to the maintenance of joint

health and overall quality of life for pets suffering from joint-related issues.

Molecular mechanisms regulating joint homeostasis

Chondrocyte function and cartilage turnover

Chondrocytes are pivotal in maintaining cartilage homeostasis by balancing the synthesis and degradation of the ECM. This regulation is primarily mediated through pathways such as SRY-box transcription factor 9 (SOX9), a transcription factor essential for chondrogenesis, which upregulates the expression of collagen type II and aggrecan, key components of cartilage that promote its integrity and functional capacity^[27,28]. Conversely, catabolic factors such as MMPs and a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS) drive cartilage degradation. Notably, MMP-13 specifically targets collagen type II for degradation, while ADAMTS-5 primarily degrades aggrecan, thereby facilitating the progression of joint degeneration^[29,30]. In addition, mechanistic target of rapamycin complex 1 (mTORC1) signaling promotes chondrocyte anabolic activity, including proliferation and matrix synthesis, while concurrently suppressing autophagy. Sustained hyperactivation of mTORC1 disrupts cellular homeostasis and contributes to OA progression. In contrast, inhibition of mTORC1 restores autophagy and alleviates cartilage degeneration, underscoring its potential as a therapeutic target for OA^[31].

Synovial inflammation and joint degeneration

The synovial membrane is integral to immune regulation within joints. In pathological conditions such as OA and rheumatoid arthritis (RA), activated fibroblast-like synoviocytes (FLS) and macrophage-like synoviocytes (MLS) release pro-inflammatory cytokines including TNF- α , IL-1 β , and IL-6. These cytokines are instrumental in promoting chronic inflammation and synovial hyperplasia, which exacerbate joint damage^[32,33]. Furthermore, dysregulated signaling pathways, notably NF- κ B and mitogen-activated protein kinase (MAPK), amplify inflammatory responses by increasing the production of additional MMPs and other inflammatory mediators, contributing to the degradation of articular cartilage^[34,35].

The anatomical structure of synovial joints is largely conserved across companion animal species, including dogs, cats, and horses. A four-stage histological model has been established to characterize the progressive degeneration of articular cartilage (Fig. 1). The process typically begins with the initial loss of proteoglycans and collagen network integrity in the superficial zone (SZ), which constitutes approximately 10%–20% of the cartilage thickness. This layer contains flattened chondrocytes and parallel collagen fibers (CF), providing shear resistance and protecting the underlying structure. As degeneration progresses, the middle zone (MZ)—accounting for 40%–60% of

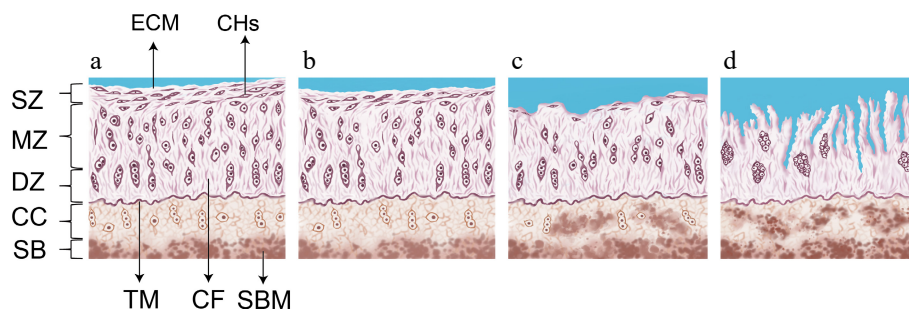


Fig. 1 Schematic diagram of the progression of articular cartilage degeneration. (a) Healthy joint cartilage. (b) Stage 0 to 1 of articular cartilage degeneration. (c) Stage 2 of articular cartilage degeneration. (d) Stage 3 of articular cartilage degeneration. ECM, extracellular matrix; CHs, chondrocytes; TM, tidemark; CF, collagen fibres; SBM, subchondral bone marrow; SZ, superficial zone; MZ, middle zone; DZ, deep zone; CC, calcified cartilage; SB, subchondral bone. Note: This figure was illustrated by Yatong Yang, one of the authors of this review. The histological evaluation criteria depicted were adapted from the concepts described in Vincent & Wann^[38], with context-specific modifications introduced in this study.

the cartilage volume—exhibits disruption of its obliquely arranged collagen network and sparsely distributed spherical chondrocytes. The damage then extends to the deep zone (DZ), which makes up approximately 30% of the volume and contains vertically oriented collagen fibers and columnar chondrocytes, offering maximal resistance to compressive loads. The tidemark (TM) separates the DZ from the calcified cartilage (CC), a mineralized region containing hypertrophic chondrocytes that anchor the cartilage to subchondral bone (SB). Histological sections (Fig. 1a–d) illustrate this progressive deterioration from superficial disruption to full-depth cartilage loss^[36,37].

Given the intricate molecular networks governing joint health, a comprehensive understanding of the mechanisms that underpin synovial and cartilage homeostasis is critical. Such insights are essential for developing targeted interventions aimed at preserving joint function and preventing degenerative conditions. As many of these molecular pathways are closely influenced by cellular metabolic states, understanding the metabolic context of joint tissues provides additional insight into the development and progression of DJDs. Recent studies have highlighted how nutrient availability and metabolite signaling affect inflammation, cartilage turnover, and chondrocyte viability^[39].

Metabolism of joints

Despite the molecular mechanism behind joint problems in animals have been thoroughly studied, metabolites—byproducts of cellular processes—have only just come to the attention of medicinal researchers. Despite the negative effects of NSAIDs, nutritional interventions that target metabolic pathways have become viable alternatives. There is increasing evidence that glucosamine, chondroitin sulfate, omega-3 fatty acids, and certain amino acids are effective in maintaining joint health^[40].

Companion animals' skeletal systems are primarily supported by their bones and joints, which are mostly composed of organic materials called collagen (which is high in glycine, proline, and 4-hydroxyproline) and glycosaminoglycans. Continuous cellular turnover and bone remodeling are necessary for the preservation of these structures, and during these processes, minerals, especially Ca and P and amino acids (AAs) play important functional and metabolic functions. Through nitric oxide (NO) pathways, L-arginine primarily affects bone resorption, whereas taurine, creatine, glycine, serine, and methionine encourage the production of new bone tissue. Additionally, the production of glucosamine, proteoglycans, and glycosaminoglycan (GAGs)—all of which are vital for preserving joint health—requires AAs. The integrity of supporting ligaments and tendons, whose function depends on the collagen content of those tissues, is essential to joint stability. Collagen formation depends on AAs, especially glycine and proline, which makes them vital for total joint integrity. Furthermore, the production of glucosamine, proteoglycans, and GAGs—all essential for preserving joint health—requires certain AAs^[41].

A comprehensive strategy is usually used to address OA and joint issues, which includes improving the quality of synovial fluid, controlling weight, and engaging in regulated activity^[42]. Pain relief and joint inflammation reduction are greatly aided by anti-inflammatory pharmaceuticals, especially NSAIDs. It is advised to use gabapentin and amantadine as supplements or substitutes, especially in situations of neuropathic pain or NSAID intolerance. Although it works well for severe pain, the bioavailability of tramadol varies from species to species^[2].

It is worth noting that prolonged use of NSAIDs can be harmful, particularly in animals that have renal dysfunction at the same time, such as those with chronic kidney disease-mineral and bone disorder

(CKD-MBD). It is necessary to take NSAIDs sparingly in these individuals to prevent further impairment of renal function^[43]. As a result, nutritional supplementation-based metabolic therapies have become viable treatment options. Through anti-inflammatory and chondroprotective processes, recent research has shown the effectiveness of targeted nutrient supplementation, including glucosamine, chondroitin sulfate, omega-3 fatty acids, and certain AAs, in promoting joint health.

Nutritional management for joint health

The bioactive compounds that are commonly included in prescription diets for supporting joint function in dogs and cats include leucine and other AAs, glucosamine and chondroitin, omega-3 fatty acids and green-lipped mussels, Type II collagen, plant polyphenols, minerals, antioxidants, and other combinations that have been identified by modern research as important nutrients that promote joint health and mobility^[44].

Leucine and other amino acids

Leucine, a branched-chain amino acid, promotes the mTOR signaling pathway to boost muscle protein synthesis and preserve muscle mass, particularly during periods of elevated physiological stress, such as joint-related disorders or age-associated muscle atrophy. Leucine supplements improve muscular strength and function, which are essential for maintaining joint stability and general musculoskeletal health when paired with a sufficient protein diet and regular exercise^[45,46].

The potential of collagen hydrolysates (CH), a blend of peptides and AAs made from collagen, as a dietary aid for dogs with OA was examined by Blees et al. Based on *in vitro* and animal research, the review examined CH absorption, bioavailability, and mechanisms of action, indicating possible chondroprotective benefits through decreased synovial inflammation, enhanced collagen synthesis, and decreased cartilage breakdown. After taking CH supplements, dogs with OA showed improvements in their lameness and pain levels in clinical studies. The authors did, however, point out several shortcomings in the existing literature, such as the use of non-validated outcome measures, the absence of consistent dosage, and the requirement for bigger, more carefully planned trials. Future studies should focus on determining CH bioavailability in dogs, identifying bioactive peptides, and using objective outcome measures to assess CH efficacy in managing canine OA^[47]. AlRaddadi et al. investigated the anti-inflammatory effects of several creatine- and amino acid-based supplements—including creatine monohydrate (CM), creatine hydrochloride (CHCl), creatinine (CRN), and Ethyl- α -(guanido-methyl) ethanoate (AlphaGEE)—in IL-1 β -stimulated primary canine chondrocytes as an *in vitro* model of OA. All compounds significantly reduced the release of PGE₂ and TNF- α in a time- and dose-dependent manner, with AlphaGEE showing the strongest effect (76% PGE₂ inhibition at 8 h). These effects were associated with decreased COX-2 expression and reduced NF- κ B phosphorylation. In contrast to the NSAID carprofen, which increased TNF- α despite suppressing PGE₂, all supplements consistently reduced TNF- α levels. The compounds also modulated oxylipin profiles—suppressing pro-inflammatory metabolites such as 12-HETE and dihomopGF₂ α , while restoring anti-inflammatory oxylipins including 18-HEPE and 20-HDoHE—indicating multi-pathway anti-inflammatory potential. Interestingly, even creatinine, typically regarded as an inactive metabolite of creatine, demonstrated notable anti-inflammatory activity^[48].

Glucosamine and chondroitin sulfate

In articular cartilage, the main substrate for the formation of GAG is the amino monosaccharide glucosamine^[49]. Because of its hydrophilic nature and resistance to compression, chondroitin sulfate, a sulfated GAG, preserves cartilage homeostasis^[50]. Both substances are structurally

incorporated into the ECM of cartilage, where they support the biomechanical characteristics of joints^[51]. Whereas chondroitin sulfate is taken from sources of animal cartilage, commercial glucosamine is made from bacterial fermentation or chitin found in crab shells^[52]. In a randomized, controlled study, Gupta et al. showed that dogs given glucosamine hydrochloride (2,000 mg/d) and chondroitin sulfate (1,600 mg/d) showed a significant reduction in pain by day 90. On day 150, the effects peaked, with observational assessments showing a 51% reduction in overall pain, a 48% reduction in limb manipulation pain, and a 43% reduction in post-exercise pain. Nevertheless, over the 150-d treatment period, objective ground force plate tests revealed no appreciable variations in peak vertical force and impulse area^[53].

Fatty acid balance and green-lipped mussels

Marine organisms represent the main source of the essential omega-3 polyunsaturated fatty acids, EPA, and DHA^[54]. The conversion of these fatty acids into specific pro-resolving mediators (SPMs), such as protectins and resolvins, regulates inflammatory responses^[55]. Dietary supplements are required to maintain optimal levels for maintaining joint health through anti-inflammatory mechanisms because endogenous production is restricted in mammals^[56].

Age-dependent cellular and functional changes were found when Lorke et al. examined the effects of dietary supplementation enhanced with antioxidants, mitochondrial cofactors, and omega-3 fatty acids on aging biomarkers in 74 shepherd dogs. Perhaps as a result of increased telomerase activity and musculoskeletal tissue support, the enriched food markedly increased the minimum telomere length and shoulder joint mobility in older dogs. According to these results, certain dietary treatments can slow down age-related physiological deterioration, with the benefits being most noticeable in elderly animals^[57]. The effects of a diet with a modified omega-6 : omega-3 ratio on dogs with OA were assessed in another investigation. At 6, 12, and 24 weeks, dogs fed a diet that reduced the omega-6 : omega-3 ratio by 34 times exhibited elevated blood omega-3 fatty acids and enhanced mobility in a randomized, double-blind, controlled study. Therefore, OA symptoms may be reduced by altering the omega-6 : omega-3 ratio in dog diets^[58]. Building on this, current research supports targeting an n-6 : n-3 fatty acid ratio below 10:1 for dogs and cats to optimize anti-inflammatory benefits and overall health^[59].

A variety of vital vitamins and minerals, omega-3 polyunsaturated fatty acids, including EPA, and aminoglycans like chondroitin sulfate are among the bioactive substances found in abundance in green-lipped mussels (*Perna canaliculus*)^[60]. Eason et al. comprehensively reviewed veterinary trials investigating the therapeutic potential of Greenshell™ mussels in managing joint-related disorders across companion animals. The analysis included research on horses, dogs, and cats and found consistent evidence that nutritional supplements can reduce the symptoms of DJD and OA. A complex matrix of bioactive substances, such as carotenoids, omega-3 polyunsaturated fatty acids, and newly discovered pro-resolving lipid mediators, were thought to be responsible for the therapeutic effectiveness. Research has shown that supplementation dosages ranging from 4 to 49 mg/kg/d can increase mobility and reduce joint discomfort^[61].

Type II collagen

The main structural protein in articular cartilage, type II collagen, creates a fibrillar network that is necessary for the mechanical characteristics of the tissue^[18]. It comes in both native and denatured (hydrolyzed) forms and is mostly derived from animal cartilage^[62]. A new immunomodulatory nutraceutical that targets companion animal joint health, undenatured type II collagen (UC-II) has a distinct molecular approach from conventional supplements. To successfully mitigate immune-mediated cartilage deterioration, UC-II induces oral tolerance, which in turn activates T-regulatory cells in Peyer's patches. These cells then move to articular tissues and emit anti-inflammatory cytokines. Extensive

research conducted on a variety of companion animal models (canine, horse, and feline) has methodically proven the therapeutic effectiveness of UC-II, showing notable improvements in locomotor function, mobility enhancement, and joint pain reduction with no side effects. Significantly, UC-II performed better than traditional glucosamine and chondroitin supplements, with recommended daily doses falling between 10 and 160 mg. Its safety profile has been thoroughly confirmed by rigorous toxicological tests, establishing UC-II as a potential, immunologically advanced treatment for osteoarticular diseases in companion animals^[63].

Plant polyphenols

Joint dysfunction and persistent pain are the results of OA, a multifactorial DJD marked by subchondral bone sclerosis, synovial inflammation, and gradual cartilage degradation. Chronic inflammation and oxidative stress are key components of its pathophysiology, and they reinforce one another. Anabolic activities are suppressed, chondrocyte death is induced, and ECM breakdown occurs when reactive oxygen species (ROS) overproduction upsets redox equilibrium. Pro-inflammatory cytokines including IL-1 β , TNF- α , and IL-6 also increase cartilage degradation by triggering catabolic enzymes and sustaining oxidative stress via signaling pathways like MAPK and NF- κ B. Tea polyphenols, curcumin, and resveratrol are polyphenolic compounds whose anti-inflammatory and antioxidant properties have been confirmed in both *in vitro* and *in vivo* models of OA^[9].

Tea polyphenols

Tea polyphenols, a family of bioactive compounds found in green tea, are mostly catechins such as EGCG and are well known for their potent anti-inflammatory and antioxidant properties^[64]. According to Singh et al., OA chondrocytes' generation of NO was markedly suppressed by EGCG when IL-1 β was present. The inhibition of inducible NO synthase (iNOS) mRNA and protein expression were linked to this impact in a dose-dependent manner. By blocking the breakdown of its inhibitor, I κ B α , EGCG mechanistically decreased NF- κ B activation by interfering with NF- κ B nuclear translocation and transcriptional activity. These findings suggest that EGCG effectively mitigates IL-1 β -driven inflammatory responses in OA chondrocytes through the inhibition of the NF- κ B/iNOS pathway^[65]. Akhtar & Haqqi demonstrated that EGCG significantly suppressed IL-1 β -induced inflammatory responses in human OA chondrocytes. EGCG reduced the production of key pro-inflammatory cytokines, including IL-6, interleukin-8 (IL-8), and TNF- α , in a dose-dependent manner. The study further revealed that these inhibitory effects were mediated through the suppression of NF- κ B and c-Jun N-terminal kinase (JNK) activation. Additionally, EGCG modulated the expression of chemokines and matrix-degrading enzymes, identifying new potential therapeutic targets^[66]. Ahmed et al. demonstrated that EGCG effectively inhibited IL-1 β -induced GAG release and the expression of MMPs (MMP-1 and MMP-13) in human chondrocytes. EGCG exhibited a differential, dose-dependent inhibitory effect on MMP expression and activity, as well as on transcription factors NF- κ B and AP-1^[67].

Curcumin

Turmeric (*Curcuma longa*) contains a bioactive component called curcumin, which is well known for its strong anti-inflammatory and antioxidant qualities^[68]. According to Zeng et al.'s comprehensive review and meta-analysis, curcumin and extract from *Curcuma longa* considerably reduced OA symptoms, including joint stiffness and function as well as pain (VAS and WOMAC-pain ratings). Curcumin had significant therapeutic benefits with a safety profile similar to that of a placebo. It showed fewer side effects and was just as effective as NSAIDs. Additionally, the combination of curcumin with NSAIDs further enhanced symptom relief and functional improvement without increasing adverse effects, reinforcing its value as a safe and effective supplement for OA, particularly with long-term use of 12 weeks or more^[69]. Chandran & Goel

conducted a randomized pilot study to evaluate the efficacy and safety of curcumin in patients with active RA. The study demonstrated that curcumin (500 mg) significantly improved Disease Activity Score (DAS28) and American College of Rheumatology (ACR) criteria scores (ACR20, ACR50, and ACR70) compared to diclofenac sodium (50 mg) or their combination. Curcumin alone achieved the highest percentage of improvement, particularly in pain reduction and inflammatory markers like CRP, without adverse effects. These findings suggest curcumin as a safe and effective therapeutic option for RA, meriting further investigation in larger trials^[70]. Chen et al. demonstrated that curcumin effectively attenuates MSU crystal-induced inflammation by inhibiting the degradation of I κ B α and suppressing NF- κ B activation. Curcumin treatment reduced mitochondrial damage, decreased ROS production, and inhibited nucleotide-binding oligomerization domain-like receptor family pyrin domain containing 3 (NLRP3) inflammasome activation in macrophages. In mouse models, curcumin alleviated joint swelling, neutrophil infiltration, and inflammatory cytokine levels (IL-1 β , IL-6, TNF- α , COX-2, PGE2)^[71]. In an explant model of cartilage inflammation, Henrotin et al. found that curcumin effectively mitigates IL-1 β -induced ECM degradation and GAG release. Treatment with 100 μ mol/L curcumin reduced GAG release triggered by IL-1 β by 20% at 10 ng/mL and 27% at 25 ng/mL, bringing levels back to those of unstimulated controls, which suggests curcumin's capability to counteract cytokine-driven cartilage degradation, positioning it as a promising candidate for therapeutic use in managing OA^[72].

Resveratrol

Resveratrol, a naturally occurring polyphenolic compound predominantly found in grape skin, red wine, and berries, has emerged as a pivotal molecule with multifaceted biological activities^[73]. Collins et al. demonstrated that low oxygen tension and acidic microenvironments induce significant cellular dysfunction in equine articular chondrocytes, characterized by reduced viability, increased glycosaminoglycan release, mitochondrial membrane depolarization, and altered ROS generation. Notably, targeted antioxidant interventions using resveratrol and N-acetylcysteine effectively mitigated these deleterious processes by restoring mitochondrial membrane potential, attenuating ROS accumulation, and normalizing antioxidant enzyme expression. Mechanistically, N-acetylcysteine exhibited superior efficacy in glutathione redox balance restoration, while resveratrol primarily modulated mitochondrial function and ROS scavenging^[74]. Ememe et al. investigated the effects of a joint supplement containing resveratrol and hyaluronic acid (RH supplement) on biochemical parameters in aged lame horses, which revealed that the RH supplement significantly reduced creatine kinase (CK) levels in the third week and decreased blood glucose levels in the second and third weeks compared to the control group. Other biochemical parameters, including cholesterol, urea, and electrolytes, remained within normal ranges without significant differences between groups, so these findings suggest that the RH supplement may alleviate oxidative stress-related muscle damage and hyperglycemia in aging horses, offering potential benefits for joint health and overall well-being^[75].

Cannabidiol

Cannabidiol (CBD) is a non-psychoactive phytocannabinoid derived from Cannabis sativa^[76]. It acts through multiple pathways, including modulation of cannabinoid receptors and inflammatory mediators^[77]. CBD's therapeutic potential is attributed to its anti-inflammatory and antioxidant properties, mediated through activation of cannabinoid type-2 (CB2) receptors and regulation of inflammatory cytokines^[78]. In a study, the researchers assessed pet owners' perceptions and experiences with CBD use in companion animals. A survey of 1,238 participants revealed that CBD, primarily in the form of treats/chews and oils, was commonly administered to dogs and cats for conditions like anxiety,

stress, and joint pain. While dosage and frequency of use were inconsistent, many owners reported improvements in their pets' condition with mild or no side effects. However, uncertainty about CBD's efficacy and safety prevented wider adoption^[79].

Minerals

Minerals such as P and Ca are important for preserving joint health. Proteoglycan function and chondrocyte survival are unaffected by short-term Ca depletion, but low Ca concentrations severely hinder GAG synthesis; total Ca deficit reduces synthesis to 52% of normal levels. The prolonged deficit may lower proteoglycan content, which might affect cartilage's biomechanical characteristics^[80].

Other antioxidants

Sick animals, such as those suffering from joint disorders or stress, produce more free radicals, which can contribute to oxidative damage and hasten the deterioration of joint tissue. In comparison to a placebo group, antioxidants such as vitamin C, vitamin E, and taurine are essential for scavenging free radicals, lowering oxidative stress, and safeguarding joint structures by halting cellular damage and promoting tissue healing processes^[81].

Other combinations

To maximize the advantages of these dietary supplements for companion animals' joint health, several researchers have looked at different combinations of these nutrients. However, a thorough assessment of the formulations' safety is crucial when creating mixtures with several ingredients. Glucosamine reached a maximum plasma concentration (C_{max}) of 9.69 μ g/mL at 2 h post-administration and was detectable for up to 8 h, according to a pharmacokinetic study on Phycos®, a multi-component joint supplement sold for both dogs and horses. In contrast, polyphenols were not detectable in plasma. Glucosamine and specific polyphenols were quantified using validated LC-MS/MS techniques; the results indicate that the Phycos® formulation may improve glucosamine absorption in comparison to data found in the literature^[82].

Comblain et al. reported that a diet supplemented with curcuminoids extract, hydrolyzed collagen, and green tea extract (CCOT) reduced pain in dogs with OA, particularly manipulation-induced pain and pain severity scores. Although no significant improvements were observed in objective measures like ground reaction forces or OA biomarkers, the CCOT group showed a significant improvement in the ability to rise from lying down, which indicate potential benefits of CCOT in alleviating chronic pain and improving specific daily activities in osteoarthritic dogs^[83]. Chen et al. demonstrated that the combination of curcumin (CUR) and catalase (CAT) effectively mitigates oxidative stress and protects chondrocytes from damage in both *in vivo* and *in vitro* models of knee osteoarthritis (KOA). *In vivo*, CUR and CAT synergistically reduced cartilage degradation and alleviated KOA progression, as evidenced by improvements in histopathological analysis and reduced oxidative damage. *In vitro* studies using primary chondrocytes isolated from SD rats demonstrated that the combination scavenged ROS, reduced lipid peroxidation, restored mitochondrial membrane potential and inhibited IL-1 β -induced chondrocyte apoptosis. Mechanistically, CUR and CAT jointly activated the NRF2/heme oxygenase-1 (HO-1) signaling pathway and enhanced the level of antioxidant enzymes, while reducing the levels of matrix-degrading enzymes (MMP3, MMP13) and promoting anti-apoptotic protein expression (BCL2)^[84]. Martello et al. demonstrated in a randomized, placebo-controlled, double-blind clinical trial ($n = 40$ dogs) that a novel multicomponent dietary supplement significantly improved clinical signs of canine OA after 60 d, as assessed by veterinary-assigned lameness scores and owner-reported HCPI, without adverse effects^[85].

Prevention and early intervention strategies

Lifestyle and weight management for joint health

Being overweight significantly exacerbates OA because it restricts physical activity and increases mechanical stress on the joints^[86,87]. Gilbert et al. found that dogs weighing above 35 kg and boxers had significantly higher OA scores in the stifle joint at the time of the diagnosis of cranial cruciate ligament (CCL) impairment^[88].

Apart from the joint stress caused by obesity, excessive physical activity has also been found to be a significant risk factor for the development of OA, especially in companion animals like dogs. A series of inflammatory processes are triggered in injured joints when they are exposed to severe mechanical loading and recurrent stress. Several evidence-based lifestyle changes are advised to preserve ideal joint health and delay the onset of OA: (1) the use of structured, moderate exercise regimens that prevent excessive mechanical stress; (2) the inclusion of sufficient rest intervals in between physical activities; (3) the maintenance of healthy body weight to lessen the mechanical stress on weight-bearing joints; and (4) prompt action when joint stress symptoms are noticed, such as adjusting the duration and intensity of activity^[89].

In clinical settings, body condition scoring (BCS) systems provide a standardized and objective method to assess and monitor the weight status of companion animals. Widely used 5-point or 9-point BCS scales allow veterinarians and pet owners to evaluate adiposity and guide nutritional interventions. Regular BCS assessments are essential for detecting early overweight trends and implementing timely lifestyle modifications to reduce mechanical stress on weight-bearing joints and prevent OA progression.

Nutritional interventions at different stages of the life cycle

In large-breed, fast-growing dogs, where excess nutrients (Ca and energy) and rapid growth increase the risk of developing skeletal problems, proper nutrition is especially important for orthopedic health. Furthermore, fatty acids can alter inflammatory and immunological responses, which can have a direct impact on the course of OA^[90].

It is equally important not to overlook the role of joint-supporting nutritional supplements during energy intake management. These supplements, which often include chondroprotective agents such as glucosamine and chondroitin sulfate, and omega-3 fatty acids, play a critical role in mitigating cartilage degradation, reducing inflammation, and promoting joint health. Integrating these supplements into dietary strategies ensures a comprehensive approach to joint health, particularly in elderly animals with heightened nutritional demands and age-related metabolic changes. Older dogs exhibited more pronounced degenerative changes in cruciate ligaments and increased severity of synovitis in the stifle joint compared to younger individuals^[91]. Given that many animals requiring joint-support diets are elderly, their age and associated metabolic changes must also be carefully considered when developing early intervention and age-specific management strategies to mitigate the progression of joint degeneration and inflammation. Integrating energy regulation with targeted supplementation establishes cornerstone strategies for optimizing therapeutic outcomes and enhancing the quality of life. In addition, effectively managing Lyme disease-associated joint disorders necessitates comprehensive preventive strategies, ranging from routine vaccination programs and regular tick checks to applying insect repellents before woodland exposure, especially for animals in endemic regions^[92].

Conclusions and perspectives

Maintaining joint health is fundamental to enhancing mobility, comfort, and overall quality of life in companion animals. Nutritional

strategies—encompassing essential fatty acids, chondroprotective agents, amino acids, antioxidants, and targeted supplements—play a central role in mitigating inflammation, supporting cartilage integrity, and slowing the progression of OA. Weight control and structured physical activity further complement these interventions.

Despite encouraging evidence, critical knowledge gaps remain regarding the precise mechanisms, optimal formulations, and long-term effects of nutritional therapies. Future research should prioritize well-controlled clinical trials integrating molecular, biochemical, and biomechanical assessments to validate efficacy. Moreover, the development of personalized nutrition, informed by genetic, metabolic, and lifestyle factors, represents a promising direction for tailoring joint health interventions. Collaborative efforts among veterinarians, researchers, and the pet food industry will be key to translating these insights into effective, evidence-based dietary solutions that improve the longevity and quality of life of companion animals.

Author contributions

The authors confirm their contributions to the paper as follows: study conception and design: Li L; manuscript preparation: Yuan X; analysis and interpretation of results: Guo X; figure design: Yang Y; draft manuscript preparation: Guo X; manuscript revision: Farooq N, Zhu Z, Li L. All authors reviewed the results and approved the final version of the manuscript.

Data availability

The data will be made available on reasonable request from the corresponding author.

Acknowledgments

The authors acknowledge Dr. Ilyas Ali for his collaboration support.

Conflict of interest

The authors declare that they have no conflict of interest.

References

1. Gildea E, Scales-Theobald E, Thompson J, Cook A, Forde K, et al. 2024. Development and validation of a quality of life and treatment satisfaction measure in canine osteoarthritis. *Frontiers in Veterinary Science* 11:1377019
2. Rychel JK. 2010. Diagnosis and treatment of osteoarthritis. *Topics in Companion Animal Medicine* 25:20–25
3. Yao Q, Wu X, Tao C, Gong W, Chen M, et al. 2023. Osteoarthritis: pathogenic signaling pathways and therapeutic targets. *Signal Transduction and Targeted Therapy* 8:56
4. Mukherjee A, Das B. 2024. The role of inflammatory mediators and matrix metalloproteinases (MMPs) in the progression of osteoarthritis. *Biomaterials and Biosystems* 13:100090
5. Budberg SC, Bartges JW. 2006. Nutrition and osteoarthritis in dogs: does it help? *The Veterinary Clinics of North America Small Animal Practice* 36:1307–23
6. Carlisle C, Metzger BT, Tintle NL, Polley K, Jackson KH, et al. 2024. The effects of omega-3 supplementation on the omega-3 index and quality of life and pain scores in dogs. *Animals* 14:3108
7. McCarthy G, O'Donovan J, Jones B, McAllister H, Seed M, et al. 2007. Randomised double-blind, positive-controlled trial to assess the efficacy of glucosamine/chondroitin sulfate for the treatment of dogs with osteoarthritis. *The Veterinary Journal* 174:54–61
8. Zhu W, Tang H, Cao L, Zhang J, Li J, et al. 2022. Epigallocatechin-3-O-gallate ameliorates oxidative stress-induced chondrocyte dysfunction and exerts chondroprotective effects via the Keap1/Nrf2/ARE signaling pathway. *Chemical Biology & Drug Design* 100:108–20

9. Sirše M. 2022. Effect of dietary polyphenols on osteoarthritis—molecular mechanisms. *Life* 12:436
10. Dämmrich K. 1991. Relationship between nutrition and bone growth in large and giant dogs. *The Journal of Nutrition* 121:S114–S121
11. Tiet T, Alman B. 2003. Developmental pathways in musculoskeletal neoplasia: involvement of the indian hedgehog-parathyroid hormone-related protein pathway. *Pediatric Research* 53:539–43
12. Parker VJ, Rudinsky AJ, Chew DJ. 2017. Vitamin D metabolism in canine and feline medicine. *Journal of the American Veterinary Medical Association* 250(11):1259–69
13. Zafalon RVA, Risolia LW, Pedrinelli V, Vendramini THA, Rodrigues RBA, et al. 2020. Vitamin D metabolism in dogs and cats and its relation to diseases not associated with bone metabolism. *Journal of Animal Physiology and Animal Nutrition* 104:322–42
14. Di Nicola V. 2020. Degenerative osteoarthritis is a reversible chronic disease. *Regenerative Therapy* 15:149–60
15. Roitner M, Klever J, Reese S, Meyer-Lindenberg A. 2024. Prevalence of osteoarthritis in the shoulder, elbow, hip and stifle joints of dogs older than 8 years. *The Veterinary Journal* 305:106132
16. Enomoto M, de Castro N, Hash J, Thomson A, Nakanishi-Hester A, et al. 2024. Prevalence of radiographic appendicular osteoarthritis and associated clinical signs in young dogs. *Scientific Reports* 14:2827
17. Johnston SA. 1997. Osteoarthritis: Joint anatomy, physiology, and pathobiology. *The Veterinary Clinics of North America Small Animal Practice* 27:699–723
18. Sophia Fox AJ, Bedi A, Rodeo SA. 2009. The basic science of articular cartilage: structure, composition, and function. *Sports Health* 1:461–68
19. Jerosch J. 2011. Effects of glucosamine and chondroitin sulfate on cartilage metabolism in OA: outlook on other nutrient partners especially Omega-3 fatty acids. *International Journal of Rheumatology* 2011:969012
20. Messina OD, Vidal Wilman M, Vidal Neira LF. 2019. Nutrition, osteoarthritis and cartilage metabolism. *Aging Clinical and Experimental Research* 31:807–13
21. Clark KL. 2007. Nutritional considerations in joint health. *Clinics in Sports Medicine* 26:101–18
22. Goggs R, Vaughan-Thomas A, Clegg PD, Carter SD, Innes JF, et al. 2005. Nutraceutical therapies for degenerative joint diseases: a critical review. *Critical Reviews in Food Science and Nutrition* 45:145–64
23. Thomas S, Browne H, Mobasheri A, Rayman MP. 2018. What is the evidence for a role for diet and nutrition in osteoarthritis? *Rheumatology* 57:iv61–iv74
24. Heinegård D, Saxne T. 2011. The role of the cartilage matrix in osteoarthritis. *Nature Reviews Rheumatology* 7:50–56
25. Zivkovic AM, Telis N, German JB, Hammock BD. 2011. Dietary omega-3 fatty acids aid in the modulation of inflammation and metabolic health. *Calif Agric* 65:106–11
26. Zeng J, Franklin DK, Das A, Hirani V. 2023. The effects of dietary patterns and food groups on symptomatic osteoarthritis: A systematic review. *Nutrition & Dietetics* 80(1):21–43
27. Akiyama H, Chaboissier MC, Martin JF, Schedl A, De Crombrughe B. 2002. The transcription factor Sox9 has essential roles in successive steps of the chondrocyte differentiation pathway and is required for expression of Sox5 and Sox6. *Genes & development* 16:2813–28
28. Liu CF, Samsa WE, Zhou G, Lefebvre V. 2017. Transcriptional control of chondrocyte specification and differentiation. *Seminars in Cell & Developmental Biology* 62:34–49
29. Glasson SS, Askew R, Sheppard B, Carito B, Blanchet T, et al. 2005. Deletion of active ADAMTS5 prevents cartilage degradation in a murine model of osteoarthritis. *Nature* 434:644–48
30. Tetlow LC, Adlam DJ, Woolley DE. 2001. Matrix metalloproteinase and proinflammatory cytokine production by chondrocytes of human osteoarthritic cartilage: associations with degenerative changes. *Arthritis & Rheumatism* 44:585–94
31. Chen J, Long F. 2018. mTOR signaling in skeletal development and disease. *Bone Research* 6:1
32. del Río E. 2025. Rethinking Osteoarthritis Management: Synergistic Effects of Chronoexercise, Circadian Rhythm, and Chondroprotective Agents. *Biomedicines* 13:598
33. McInnes IB, Schett G. 2011. The pathogenesis of rheumatoid arthritis. *New England Journal of Medicine* 365:2205–19
34. Thysen S, Luyten FP, Lories RJU. 2015. Targets, models and challenges in osteoarthritis research. *Disease Models & Mechanisms* 8:17–30
35. Xu C, Fang MY, Wang K, Liu J, Tai GP, et al. 2020. Discovery and development of inflammatory inhibitors from 2-phenylchromonone (flavone) scaffolds. *Current Topics in Medicinal Chemistry* 20:2578–98
36. Goldring MB, Marcu KB. 2009. Cartilage homeostasis in health and rheumatic diseases. *Arthritis Research & Therapy* 11:224
37. Caola G. 2003. The Anatomy and Physiology of Diarthroses. *Veterinary Research Communications* 27:69–73
38. Vincent T, Wann A. 2019. Mechanoadaptation: articular cartilage through thick and thin. *The Journal of Physiology* 597(5):11271–81
39. Mobasheri A, Henrotin Y, Biesalski HK, Shakibaei M. 2012. Scientific Evidence and Rationale for the Development of Curcumin and Resveratrol as Nutraceuticals for Joint Health. *International Journal of Molecular Sciences* 13:4202–32
40. Birlik A. 2023. Nutritional Supplements for Musculoskeletal Health. In *Functional exercise anatomy and physiology for physiotherapists*, ed. Kaya Utlu D. Cham: Springer. pp. 547–59 doi: 10.1007/978-3-031-27184-7_28
41. Lopez AN, Bazer FW, Wu G. 2024. Functions and metabolism of amino acids in bones and joints of cats and dogs. In *Nutrition and Metabolism of Dogs and Cats*, ed. Wu G. Cham, Switzerland: Springer. pp. 155–75 doi: 10.1007/978-3-031-54192-6_7
42. Andraskar K. 2024. Recent advances in the understanding and management of osteoarthritis: a comprehensive review. *Journal of Internal Medicine and Pharmacology* 1:53–62
43. LaForge JM, Urso K, Day JM, Bourgeois CW, Ross MM, et al. 2023. Non-steroidal anti-inflammatory drugs: clinical implications, renal impairment risks, and AKI. *Advances in Therapy* 40:2082–96
44. Fujiki M, Shineha J, Yamanokuchi K, Misumi K, Sakamoto H. 2007. Effects of treatment with polysulfated glycosaminoglycan on serum cartilage oligomeric matrix protein and C-reactive protein concentrations, serum matrix metalloproteinase-2 and -9 activities, and lameness in dogs with osteoarthritis. *American Journal of Veterinary Research* 68:827–33
45. Devries MC, McGlory C, Bolster DR, Kamil A, Rahn M, et al. 2018. Leucine, not total protein, content of a supplement is the primary determinant of muscle protein anabolic responses in healthy older women. *The Journal of Nutrition* 148:1088–95
46. Norton LE, Layman DK. 2006. Leucine regulates translation initiation of protein synthesis in skeletal muscle after exercise. *The Journal of Nutrition* 136:533s–537s
47. Bles NR, Teunissen M, Dobenecker B, Prawitt J, Tryfonidou MA, et al. 2024. Collagen hydrolysates as nutritional support in canine osteoarthritis: a narrative review. *Journal of Animal Physiology and Animal Nutrition* 109:590–600
48. AlRaddadi EA, Winter T, Aukema HM, Miller DW. 2019. Effects of various dietary supplements on inflammatory processes in primary canine chondrocytes as a model of osteoarthritis. *Canadian Journal of Veterinary Research* 83:206–17
49. Henrotin Y, Chevalier X, Herrero-Beaumont G, McAlindon T, Mobasheri A, et al. 2013. Physiological effects of oral glucosamine on joint health: current status and consensus on future research priorities. *BMC Research Notes* 6:115
50. Djerbal L, Lortat-Jacob H, Kwok J. 2017. Chondroitin sulfates and their binding molecules in the central nervous system. *Glycoconjugate Journal* 34:363–76
51. Martel-Pelletier J, Barr AJ, Cicuttini FM, Conaghan PG, Cooper C, et al. 2016. Osteoarthritis. *Nature Reviews Disease Primers* 2:16072
52. Gregory PJ, Fellner C. 2014. Dietary supplements as disease-modifying treatments in osteoarthritis: a critical appraisal. *P & T* 39:436–52
53. Gupta RC, Canerdy TD, Lindley J, Konemann M, Minniear J, et al. 2012. Comparative therapeutic efficacy and safety of type-II collagen (UC-II), glucosamine and chondroitin in arthritic dogs: pain evaluation by ground force plate. *Journal of Animal Physiology and Animal Nutrition* 96:770–77
54. Calder PC. 2017. Omega-3 fatty acids and inflammatory processes: from molecules to man. *Biochemical Society Transactions* 45:1105–15
55. Basil MC, Levy BD. 2016. Specialized pro-resolving mediators: endogenous regulators of infection and inflammation. *Nature Reviews Immunology* 16:51–67

56. Barden AE, Moghaddami M, Mas E, Phillips M, Cleland LG, et al. 2016. Specialised pro-resolving mediators of inflammation in inflammatory arthritis. *Prostaglandins, Leukotrienes and Essential Fatty Acids* 107:24–29
57. Lorke M, Willen M, Lucas K, Schille JT, Lüder Ripoli F, et al. 2020. Effect of antioxidants, mitochondrial cofactors and omega-3 fatty acids on telomere length and kinematic joint mobility in young and old shepherd dogs - a randomized, blinded and placebo-controlled study. *Research in Veterinary Science* 129:137–53
58. Roush JK, Dodd CE, Fritsch DA, Allen TA, Jewell DE, et al. 2010. Multi-center veterinary practice assessment of the effects of omega-3 fatty acids on osteoarthritis in dogs. *Journal of the American Veterinary Medical Association* 236:59–66
59. Burrin S, Richards T, Krebs G, Trevizan L, Rankovic A, et al. 2024. The balance of n-6 and n-3 fatty acids in canine, feline, and equine nutrition: exploring sources and the significance of alpha-linolenic acid. *Journal of Animal Science* 102:skae143
60. Cobb CS, Ernst E. 2006. Systematic review of a marine nutraceutical supplement in clinical trials for arthritis: the effectiveness of the New Zealand green-lipped mussel *Perna canaliculus*. *Clinical Rheumatology* 25:275–84
61. Eason CT, Adams SL, Puddick J, Romanazzi D, Miller MR, et al. 2018. Greenshell™ mussels: a review of veterinary trials and future research directions. *Veterinary Sciences* 5:36
62. Bello AE, Oesser S. 2006. Collagen hydrolysate for the treatment of osteoarthritis and other joint disorders: a review of the literature. *Current Medical Research and Opinion* 22:2221–32
63. Gencoglu H, Orhan C, Sahin E, Sahin K. 2020. Undenatured Type II Collagen (UC-II) in joint health and disease: a review on the current knowledge of companion animals. *Animals* 10:697
64. Musial C, Kuban-Jankowska A, Gorska-Ponikowska M. 2020. Beneficial Properties of Green Tea Catechins. *International Journal of Molecular Sciences* 21:1744
65. Singh R, Ahmed S, Islam N, Goldberg VM, Haqqi TM. 2002. Epigallocatechin-3-gallate inhibits interleukin-1 β -induced expression of nitric oxide synthase and production of nitric oxide in human chondrocytes: suppression of nuclear factor κ B activation by degradation of the inhibitor of nuclear factor kappaB. *Arthritis and Rheumatism* 46:2079–86
66. Akhtar N, Haqqi TM. 2011. Epigallocatechin-3-gallate suppresses the global interleukin-1 β -induced inflammatory response in human chondrocytes. *Arthritis Research & Therapy* 13:R93
67. Ahmed S, Wang N, Lalonde M, Goldberg VM, Haqqi TM. 2004. Green tea polyphenol epigallocatechin-3-gallate (EGCG) differentially inhibits interleukin-1 β -induced expression of matrix metalloproteinase-1 and -13 in human chondrocytes. *The Journal of Pharmacology and Experimental Therapeutics* 308:767–73
68. Kotha RR, Luthria DL. 2019. Curcumin: biological, pharmaceutical, nutraceutical, and analytical aspects. *Molecules* 24:2930
69. Zeng L, Yu G, Hao W, Yang K, Chen H. 2021. The efficacy and safety of Curcuma longa extract and curcumin supplements on osteoarthritis: a systematic review and meta-analysis. *Bioscience Reports* 41:BSR20210817
70. Chandran B, Goel A. 2012. A randomized, pilot study to assess the efficacy and safety of curcumin in patients with active rheumatoid arthritis. *Phytotherapy Research* 26:1719–25
71. Chen B, Li H, Ou G, Ren L, Yang X, et al. 2019. Curcumin attenuates MSU crystal-induced inflammation by inhibiting the degradation of I κ B α and blocking mitochondrial damage. *Arthritis Research & Therapy* 21:193
72. Henrotin Y, Clutterbuck AL, Allaway D, Lodwig EM, Harris P, et al. 2010. Biological actions of curcumin on articular chondrocytes. *Osteoarthritis and Cartilage* 18:141–49
73. Tian B, Liu J. 2020. Resveratrol: a review of plant sources, synthesis, stability, modification and food application. *Journal of the Science of Food and Agriculture* 100:1392–404
74. Collins JA, Moots RJ, Clegg PD, Milner PI. 2015. Resveratrol and N-acetylcysteine influence redox balance in equine articular chondrocytes under acidic and very low oxygen conditions. *Free Radical Biology and Medicine* 86:57–64
75. Ememe MU, Abdullahi US, Sackey AKB, Ayo JO, Mshelia WP, Edeh RE. 2016. Effects of a joint supplement whose main components are resveratrol and hyaluronic acid on some biochemical parameters in aged lame horses. *Journal of Equine Science* 27:19–22
76. Pisanti S, Malfitano AM, Ciaglia E, Lamberti A, Ranieri R, et al. 2017. Cannabidiol: state of the art and new challenges for therapeutic applications. *Pharmacology & Therapeutics* 175:133–50
77. Burstein S. 2015. Cannabidiol (CBD) and its analogs: a review of their effects on inflammation. *Bioorganic & Medicinal Chemistry* 23:1377–85
78. Mechoulam R, Peters M, Murillo-Rodríguez E, Hanuš L. 2007. Cannabidiol--recent advances. *Chemistry & Biodiversity* 4: 1678–92
79. Corsato Alvarenga I, MacQuiddy B, Duerr F, Elam LH, McGrath S. 2023. Assessment of cannabidiol use in pets according to a national survey in the USA. *Journal of Small Animal Practice* 64:513–21
80. Palmoski MJ, Brandt KD. 1979. Effect of calcipenia on proteoglycan metabolism and aggregation in normal articular cartilage in vitro. *The Biochemical Journal* 182:399–406
81. Rhouma M, El-Warrak A, Troncy E, Beaudry F, Chorfi Y. 2013. Anti-inflammatory response of dietary vitamin E and its effects on pain and joint structures during early stages of surgically induced osteoarthritis in dogs. *Canadian journal of veterinary research [Revue canadienne de recherche vétérinaire]* 77:191–98
82. Martinez SE, Lillico R, Lakowski TM, Martinez SA, Davies NM. 2017. Pharmacokinetic analysis of an oral multicomponent joint dietary supplement (Phycos(®)) in dogs. *Pharmaceutics* 9:30
83. Comblain F, Barthélémy N, Lefebvre M, Schwartz C, Lesponne I, et al. 2017. A randomized, double-blind, prospective, placebo-controlled study of the efficacy of a diet supplemented with curcuminoids extract, hydrolyzed collagen and green tea extract in owner's dogs with osteoarthritis. *BMC Veterinary Research* 13:395
84. Chen B, He Q, Chen C, Lin Y, Xiao J, et al. 2023. Combination of curcumin and catalase protects against chondrocyte injury and knee osteoarthritis progression by suppressing oxidative stress. *Biomedicine & Pharmacotherapy* 168:115751
85. Martello E, Bigliati M, Adami R, Biasibetti E, Bisanzio D, et al. 2022. Efficacy of a dietary supplement in dogs with osteoarthritis: A randomized placebo-controlled, double-blind clinical trial. *PLoS One* 17:e0263971
86. Logerstedt DS, Ebert JR, MacLeod TD, Heiderscheit BC, Gabbett TJ, et al. 2022. Effects of and Response to Mechanical Loading on the Knee. *Sports Medicine* 52:201–35
87. Felson DT. 1996. Does excess weight cause osteoarthritis and, if so, why? *Annals of the Rheumatic Diseases* 55:668–70
88. Gilbert S, Langenbach A, Marcellin-Little DJ, Pease AP, Ru H. 2019. Stifle joint osteoarthritis at the time of diagnosis of cranial cruciate ligament injury is higher in Boxers and in dogs weighing more than 35 kilograms. *Veterinary Radiology & Ultrasound* 60:280–88
89. Bland SD. 2000. Canine osteoarthritis and treatments: a review. *Veterinary Science Development* 5(2):5931
90. Richardson DC, Schoenherr WD, Zicker SC. 1997. Nutritional Management of Osteoarthritis. *Veterinary Clinics of North America: Small Animal Practice* 27:883–911
91. Döring AK, Junginger J, Hewicker-Trautwein M. 2018. Cruciate ligament degeneration and stifle joint synovitis in 56 dogs with intact cranial cruciate ligaments: correlation of histological findings and numbers and phenotypes of inflammatory cells with age, body weight and breed. *Veterinary Immunology and Immunopathology* 196:5–13
92. Littman MP, Goldstein RE, Labato MA, Lappin MR, Moore GE. 2006. ACVIM small animal consensus statement on Lyme disease in dogs: diagnosis, treatment, and prevention. *Journal of Veterinary Internal Medicine* 20:422–34



Copyright: © 2025 by the author(s). Published by Maximum Academic Press on behalf of Nanjing Agricultural University. This article is an open access article distributed under Creative Commons Attribution License (CC BY 4.0), visit <https://creativecommons.org/licenses/by/4.0/>.