

Identification of the key odorants causing soapy and metallic off-flavors in alkali-refined rapeseed oil via molecular sensory science

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Abstract

Alkali refining may sometimes induce soapy and metallic off-flavors in cold-pressed rapeseed oil, yet the molecular mechanisms remain unclear. A comparative flavor sensomics study was performed on crude cold-pressed, thermally treated control, water-degummed, and alkali-refined rapeseed oils. Quantitative descriptive analysis (QDA) showed that alkali refining increased soapy and metallic sensory attributes. Aroma extract dilution analysis (AEDA) detected 46 odor-active compounds across processing stages. The metallic off-flavors were primarily associated with elevated levels of *trans*-4,5-epoxy-(E)-2-decenal, (Z)-1,5-octadien-3-one, and 1-octen-3-one. The soapy attribute was linked to the accumulation of medium-chain saturated aldehydes, particularly nonanal and decanal. Omission experiments confirmed that these aldehydes collectively contributed to the soapy off-flavors, despite their low individual odor activity values (OAVs). Collectively, aldehydes and ketones were identified as the primary sources of off-flavor, marking the alkali neutralization step as the critical control point for flavor quality. These findings can offer theoretical guidance for process optimization of cold-pressed rapeseed oil refining.

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Introduction

Rapeseed oil currently occupies a pivotal position in the global edible oil market, widely valued by consumers and manufacturers alike for its distinctive sensory profile and favorable nutritional properties^[1,2]. However, crude rapeseed oil is a chemically complex matrix containing free fatty acids, phospholipids, pigments, and trace metal ions, which could compromise its sensory quality and oxidative stability^[3]. Consequently, the crude oil must undergo a standardized, multi-stage refining process to ensure consumption safety and meet requisite industrial versatility standards. This refining sequence is essential for optimizing key physicochemical attributes, most notably by elevating the smoke point and ensuring the superior oxidative stability of the final product^[4]. Currently, refined rapeseed oil has become indispensable across a wide spectrum of applications, ranging from domestic cooking to large-scale industrial food manufacturing^[5].

Traditionally, the chemical refining sequence of rapeseed oil includes hydration degumming, alkali refining, adsorption decolorization, and high-temperature deodorization^[6]. Among these unit operations, alkali refining represents a critical intervention point. In this step, free fatty acids are neutralized by aqueous alkaline solutions to form soapstock, a process central to reducing acidity and improving stability^[7]. While the primary function of neutralization is the efficient removal of free fatty acids and residual phospholipids, exposure to high pH and elevated temperatures may trigger complex, non-enzymatic secondary reactions^[8]. Although standard refining protocols generally yield high-quality neutral oils, sensory deviations manifested as soapy or metallic off-flavors have been frequently observed in substandard batches. Distinct from the well-characterized oxidative rancidity associated with long-term storage, these process-induced off-flavors occur immediately following neutralization^[9]. This immediate degradation significantly diminishes consumer acceptance and necessitates aggressive

downstream deodorization; conversely, such severe processing can strip the oil of its natural tocopherols and desirable aroma compounds^[10]. Although the flavor profile of cold-pressed rapeseed oil has been comprehensively characterized, the molecular mechanisms driving the formation of soapy and metallic off-flavors during alkali refining remain poorly understood.

For the accurate elucidation of off-flavor compounds in complex vegetable oil matrices, modern flavor chemistry has established a systematic research paradigm centered on molecular sensory science^[11,12]. In this approach, gas chromatography-olfactometry (GC-O) is first employed to localize key regions within the aroma-active fraction, thereby focusing on odorants that are directly relevant to human perception and avoiding misinterpretation based solely on chemical abundance^[13]. Subsequently, AEDA is applied to rank odor potency, followed by quantitative determination of selected compounds and calculation of OAV, which translate analytical concentrations into indices of sensory relevance^[14]. Finally, aroma recombination and omission experiments are conducted to verify the causal contribution of candidate key odorants, ensuring that the proposed molecular set is capable of reproducing the characteristic odor of the original sample^[15]. Using this analytical strategy, we have previously investigated the volatile off-flavor formation in small mill sesame oil and peanut oil, successfully identifying the key off-flavors responsible for their characteristic sensory defects^[16,17].

Thus, this study aims to elucidate the impact of the alkali refining process on the flavor quality of rapeseed oil and its molecular mechanisms. Specific objectives include: (1) using QDA to characterize the evolution of sensory attributes in rapeseed oil from crude to alkali-refined stages; (2) applying GC-O combined with AEDA to screen high-potency odor compounds, and identifying key odorants responsible for soapy and metallic flavors through gas chromatography-mass spectrometer (GC-MS) analysis and OAV calculations;

(3) verifying the contributions of key odorants to overall off-flavor characteristics via flavor recombination and omission tests. This research seeks to provide a scientific explanation for alkali-induced off-flavors in rapeseed oil and offer theoretical guidance for targeted regulation and quality optimization in the refining process.

Materials and methods

Samples

Rapeseeds (*Brassica napus* L., cv. Zhongyouza 501) were obtained from Hubei Province, China. Crude oil was extracted using a low-temperature screw press (CA59G, Komet, Germany) with the oil outlet temperature controlled below 50 °C. The thermally treated control oil was prepared by heating the crude oil in a reactor at 65 °C for 40 min under constant stirring (60 rpm) without water addition. For the preparation of the water-degummed oil, the crude oil was similarly heated to 65 °C and mixed with distilled water (2.5% w/w). The mixture was stirred rapidly (400 rpm) for 2 min to disperse water, followed by a slow agitation period (30 min, 60 rpm) to induce phospholipid hydration and flocculation. The aqueous gum phase was separated by centrifugation (5,000 × *g*, 20 min, 20 °C). Then, the alkali-refined oil was produced by further processing the water-degummed oil. The oil was heated to 85 °C for deacidification, treated with a sodium hydroxide solution (18° Bé) added dropwise over 10 min under stirring (350 rpm), and maintained for 25 min. After soapstock removal via centrifugation (4,500 × *g*, 15 min), the oil phase was washed twice with hot deionized water (90 °C, 10% w/w) and vacuum-dried at 50 °C for 30 min. All samples were flushed with high-purity nitrogen and stored in amber glass bottles at −20 °C prior to analysis.

Chemicals

All chemicals were of analytical grade, and solvents were of HPLC grade. Authentic flavor standards (purity ≥ 90%) used for identification and quantification, as well as the internal standard 2-methyl-3-heptanone, were obtained from Sigma-Aldrich (St. Louis, MO, USA) and are listed in [Supplementary Table S1](#). Dichloromethane and anhydrous sodium sulfate were supplied by Merck (Darmstadt, Germany). Sodium hydroxide and citric acid used for the refining process were of food grade. Both the recovery rate study and the construction of the aroma recombination model used deodorized sunflower seed oil as a matrix.

Sensory evaluation

The sensory profile of the rapeseed oil samples was quantitatively characterized by a panel of 12 trained assessors (six males, six females; aged 20–35 years) recruited from China Agricultural University. Sensory assessors were selected and calibrated according to ISO 8586 protocols to ensure the accurate and consistent application of specific sensory descriptors and intensity scales. The study protocol was approved by the university's human research ethics committee (CAUHR-20240703). Based on preliminary trials and the standard lexicon for rapeseed oil, seven distinct sensory attributes were defined: pungent, cabbage-like, green, fatty, soapy, metallic, and rancid. Authentic chemical reference standards, presented in deodorized sunflower oil, served to anchor the panel's intensity ratings and minimize inter-assessor variation. The definitions and reference concentrations were established as follows ([Supplementary Table S2](#)). Sensory sessions were conducted in a

climate-controlled sensory laboratory at 22 °C. Ten mL oil samples were presented monadically in coded, capped amber glass vials. Panelists were instructed to assess the intensity of each attribute on a 10-point linear scale (0 = not detected; 10 = very strong). Each sample was evaluated in triplicate by every assessor across three independent sessions.

Isolation, identification, and quantification of volatile flavor compounds

An accurately weighed sample of rapeseed oil (50.00 ± 0.01 g) was dispersed in 150 mL of HPLC-grade dichloromethane within a sealed amber flask. Then 200 µL of an internal standard solution containing 2-methyl-3-heptanone (0.816 mg/mL in dichloromethane) was added. The mixture was agitated on a thermostatic shaker at 180 rpm for 8 h at 4 °C. The solvent-assisted flavor evaporation (SAFE) unit was operated under a high vacuum of 10^{−5} mbar, generated by a molecular turbine vacuum pump. The flask containing the dichloromethane extract was immersed in a circulating water bath maintained at 40 °C. The distilled volatiles were efficiently trapped in a cryogenic collection flask cooled with liquid nitrogen. Upon completion of the distillation, the recovered distillate was dried over anhydrous sodium sulfate. The volume of the dried extract was then reduced to approximately 1 mL using a Vigreux column at 40 °C. A final concentration step to a precise volume of 100.0 µL was performed under a gentle stream of high-purity nitrogen. The resulting concentrated extract was immediately used for all subsequent analyses to minimize the loss of volatile components.

Instrumental analysis was conducted on an Agilent 7890B gas chromatograph interfaced with an Agilent 5977B mass selective detector and a Gerstel ODP 3 olfactory detection port. A 1.0 µL aliquot of the final extract was introduced into the GC system via a splitless injection, with the inlet temperature held at 250 °C. Chromatographic separation was performed on an HP-INNOWax fused-silica capillary column (30 m × 0.25 mm × 0.25 µm). The carrier gas was high-purity helium, maintained at a constant flow rate of 1.0 mL/min. The oven temperature program consisted of an initial hold at 40 °C for 5 min, a 4 °C/min ramp to 160 °C held for 5 min, followed by a 5 °C/min increase to 240 °C held for 5 min. The MSD was configured to operate in electron ionization mode at 70 eV, with the ion source and quadrupole temperatures set at 230 °C and 150 °C, respectively. A solvent delay of 5.0 min was employed. Mass spectra were acquired over an *m/z* range of 30–500. For olfactometry, the column effluent was split 1:1 (v/v) between the MSD and the ODP. The ODP transfer line was heated to 250 °C, and humidified air was supplied at 50 mL/min.

Flavor compound identification and aroma extract dilution analysis

The identification of volatile compounds was established using mass spectral data, retention index, and odor characteristics. Initially, the acquired mass spectra were screened against the NIST 2020 library, requiring a similarity match factor exceeding 80%. To substantiate these spectral matches, the retention indices were experimentally determined relative to a homologous series of *n*-alkanes on the HP-INNOWax column. These values were aligned with the retention indices of authentic standards analyzed under identical chromatographic conditions. Finally, conclusive identification was secured through specific olfactory confirmation at the olfactory detection port, ensuring that the sensory descriptors perceived by the trained assessors were consistent with the aromas of the corresponding reference standards.

The sensory potency of each odorant was assessed by AEDA. This required the preparation of a serial 1:2 (v/v) dilution series of the original extract using dichloromethane. Each dilution was then analyzed by the GC-O panel, which consisted of three trained assessors (two female, one male; aged 20–30 years). The flavor dilution (FD) factor was established as the maximum dilution at which a characteristic aroma was still discernible and correctly identified by a minimum of two of the three assessors.

Quantitative analysis and odor activity value calculation

The quantitative analysis of targeted volatile flavor compounds was conducted utilizing an external calibration method that incorporated an internal standard, 2-methyl-3-heptanone. A series of mixed calibration standard solutions was formulated. The preparation involved dissolving precisely weighed amounts of authentic analyte standards and a consistent mass of the internal standard in dichloromethane to achieve seven distinct concentration levels. Each calibration standard was analyzed in triplicate via GC-MS with a 1.0 μL injection volume. Calibration curves were subsequently established by plotting the peak area ratio of the analyte to the IS against the mass ratio of the analyte standard to the internal standard. The resulting data points were fitted using linear regression analysis to derive the calibration equation, represented as:

$$Y = \alpha X + \beta$$

In this equation, the variable Y denoted the analyte-to-IS peak area ratio, while X corresponded to the mass ratio of the analyte standard to the IS within the calibration solutions.

To evaluate the method's efficacy, a recovery study was performed. Blank deodorized oil samples were fortified with the target analytes at three separate concentrations, alongside a constant quantity of the internal standard, before being subjected to the entire SAFE extraction and GC-MS analysis procedure. The recovery rate was determined using the expression:

$$R = (A_1 - A_0)/A \times 100\%$$

Here, A_1 was the concentration of the analyte determined in the spiked blank oil sample after undergoing the complete SAFE analytical procedure, A_0 represented the concentration of the analyte determined in the unspiked blank oil sample after undergoing the complete SAFE analytical procedure, and A was the theoretical concentration of the analyte that was added into the blank oil sample. Furthermore, the limits of detection and quantification for the method were established by analyzing the chromatograms of standard solutions, defining them at signal-to-noise ratios of 3:1 and 10:1, respectively.

To estimate the sensory contribution of each odor-active compound to the overall aroma, OAV were calculated. The OAV for each compound was determined by dividing its quantified concentration by its odor threshold. The odor thresholds were selected from peer-reviewed literature, prioritizing values that were determined in an oil matrix^[18,19].

Aroma recombination and omission experiments

To validate the accuracy of the identification and quantification results, an aroma recombination model was prepared to simulate the flavor profile of the alkali-refined rapeseed oil^[20]. A deodorized oil matrix was used as the base. Stock solutions of the identified odorants were prepared in odorless medium-chain triglyceride oil and added to the blank matrix at the exact concentrations determined in the alkali-refined samples. The mixture was stirred for

30 min and equilibrated at room temperature for 1 h to ensure homogeneity. The sensory profile of the recombination model was then evaluated by the trained panel using the same protocol and attributes described in the sensory evaluation section. The similarity between the recombination model and the original oil sample was assessed by comparing the mean intensity scores of the key attributes.

To elucidate the contribution of specific flavor classes or individual odorants to the soapy and metallic off-flavors, omission experiments were conducted. A series of omission models was prepared by excluding a specific group of compounds, such as nitriles, aldehydes, or ketones, from the complete recombination model described above. The models were presented to the panelists in a randomized order. Assessors were instructed to rate the intensities of the soapy and metallic attributes for each omission model using the 10-point scale. The significance of the sensory differences between the omission models and the full recombination model was determined by statistical analysis.

Statistical analysis

All experiments were conducted in triplicate, and data are presented as mean \pm standard deviation. Statistical analysis was performed using SPSS software (v. 22.0, IBM, Armonk, NY, USA). Significant differences between means were determined by one-way analysis of variance (ANOVA) followed by Tukey's post-hoc test.

Results

Evolution of the sensory profile from crude to alkali-refined oil using quantitative descriptive analysis

To systematically evaluate the impact of processing stages on the aroma profile of rapeseed oil, a comparative QDA was conducted on the crude cold-pressed oil, the thermally treated control oil, the water-degummed oil, and the alkali-refined oil. As shown in Fig. 1, the refining process significantly changed the sensory profile of the rapeseed oil ($p < 0.05$).

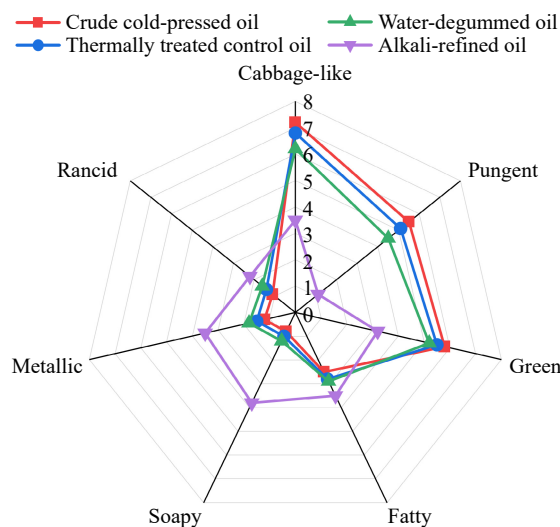


Fig. 1 Sensory attribute profiles of rapeseed oils at different processing stages.

The crude cold-pressed oil exhibited the highest intensities for cabbage-like (7.2 ± 0.6) and green (5.8 ± 0.5) odors, alongside a notable pungency (5.5 ± 0.7). These attributes are typical of high-quality cold-pressed oils, deriving largely from the enzymatic hydrolysis of glucosinolates and the lipoxygenase pathway activity inherent to the raw seeds^[21,22].

In contrast, the alkali-refined oil exhibited a change in its sensory profile. The characteristic pungent and green attributes decreased to levels of 1.1 and 3.2, respectively, indicating a loss of the original characteristics. Concurrently, the alkali treatment was associated with the formation of undesirable off-flavors. The soapy and metallic attributes reached intensities of 3.8 ± 0.7 and 3.5 ± 0.6 in the alkali-refined oil. These values exceeded those observed in all other samples ($p < 0.05$), including the thermally treated control oil. The thermally treated control oil and water-degummed oil exhibited sensory profiles that were transitional between those of the crude and refined oils. Thermal stress alone in the thermally treated control oil was associated with a slight increase in metallic nuances (1.5) relative to the crude cold-pressed oil (1.2), and a decrease in pungency (5.1). However, these values remained lower than those observed in the alkali-refined oil, reinforcing the hypothesis that the high pH environment is a catalyst for specific off-flavor formation. Interestingly, the cabbage-like attribute, while highest in the crude oil, was reduced in the alkali-refined oil (3.5). These sensory data indicated that while thermal treatment initiates general flavor degradation, the alkali environment represents the primary condition associated with the dominant soapy and metallic defects. These sensory results provided the necessary foundation for the subsequent flavor identification.

Identification of odor-active compounds in rapeseed oil across processing stages via aroma extract dilution analysis

To elucidate the chemical basis of the sensory changes described above, AEDA was applied to the volatile flavor compounds isolated from the crude cold-pressed oil, thermally treated control oil, water-degummed oil, and alkali-refined oil. This stepwise screening resulted in the detection of 46 odor-active compounds in the FD factor range of 1 to 512. Comparative evaluation of FD factors across the four processing stages (Table 1) revealed a shift that aligned with the sensory observed by QDA.

The characteristic pungency of the raw material, which received the highest intensity score in the crude cold-pressed oil, was clearly attributable to a specific group of nitriles and isothiocyanates. In the crude cold-pressed oil, allyl isothiocyanate (pungent and sulfury odor) exhibited a high FD factor of 128. This compound is a well-known product of sinigrin and is a key marker for the biting taste of *Brassica* vegetables^[19,23]. However, this flavor compound was progressively depleted during the processing simulation (FD = 128 vs 64 vs 32 vs < 1). A similar downward trend was observed for other nitrile compounds, such as 3-butenenitrile (pungent, mustard-like, FD = 64 vs 64 vs 32 vs 8) and 4-pentenitrile, which dropped from the raw material to the alkali-refined oil. This drastic reduction in the FD factors of nitrogen- and sulfur-containing compounds aligned with the statistical decrease of the pungent sensory attribute in the alkali-refined oil, confirming that the refining process effectively strips the oil of its natural pungency.

Furthermore, the fresh green attribute of the crude cold-pressed oil was dominated by (Z)-3-hexenal. This lipoxygenase-derived aldehyde showed the highest activity in the raw oil (FD = 128) but was labile during refining (fresh green, FD = 128 vs 64 vs 32 vs < 1). Its

degradation, likely via isomerization to (E)-2-hexenal or oxidation to acid, correlated with the loss of the freshly crushed seed character. In contrast, hexenal, representing a more generic fatty, grassy, and green note, showed an increase in the processed samples (FD = 32 vs 32 vs 64 vs 64)^[24]. This increase likely reflected mild oxidative changes occurring during the heating and mixing steps.

The emergence of the unpleasant soapy attribute in the alkali-refined oil was associated with the accumulation of medium-chain saturated aldehydes. Specifically, nonanal showed a notable increase in the alkali-treated sample. While present in the crude oil, its potency increased substantially in the final stage (citrus-like, soapy, FD = 16 vs 32 vs 32 vs 128). This was accompanied by increases in decanal, which is described as having soapy and coriander-like notes (FD = 4 vs 8 vs 16 vs 64). The elevated FD factors of these specific aldehydes in the final stage contrast with the loss of shorter-chain green aldehydes.

The metallic attribute, which was perceived in the alkali-refined oil, can be primarily traced back to specific lipid peroxidation products. The most potent contributor was identified as *trans*-4,5-epoxy-(E)-2-decenal. This compound had a negligible FD factor in the crude cold-pressed oil, but increased to the highest FD factor after alkali-refined (intense metallic, FD = < 1 vs 4 vs 8 vs 256). The detection of this epoxy-aldehyde has an extremely low odor threshold, meaning even trace amounts can dominate the aroma profile^[9]. Its contribution was likely synergistically enhanced by vinyl ketones, such as 1-octen-3-one (mushroom-like, metallic, FD = 8 vs 16 vs 32 vs 128), which also reached its maximum intensity in the final oil. Finally, the cabbage-like odor was consistent with the presence of volatile sulfur-containing compounds, particularly dimethyl trisulfide, which showed high FD factors in the crude oil (FD = 256) but decreased significantly in the alkali-refined oil (FD = 16), mirroring the sensory reduction of the cabbage-like odor^[25].

Quantitative analysis and odor activity value calculation of key aroma compounds in rapeseed oil across processing stages

To identify the key aroma compounds responsible for the soapy and metallic off-flavors, the concentrations of the 46 identified volatiles were quantified using external standard calibration curves constructed using peak area ratios relative to the internal standard (Supplementary Table S3). The concentrations (Fig. 2; Supplementary Table S4) were then divided by their respective odor thresholds to calculate OAV (Table 2).

The sensory profile of the crude cold-pressed oil, characterized by pungent and cabbage-like attributes, was rationalized by high concentrations of sulfur-containing compounds. Allyl isothiocyanate (*no.* 38) was quantified at 139.2 $\mu\text{g}/\text{kg}$ in the crude cold-pressed oil. A depletion was observed during the refining simulation, decreasing to 15.4 $\mu\text{g}/\text{kg}$ in the alkali-refined oil. This loss aligned with the decline in the perception of pungent attributes^[26]. The OAV calculation reflected this trend, showing a decrease from 5 in the crude oil to 1 in the alkali-refined oil. This demonstrated that alkali refining could remove some volatile hydrolysis products of glucosinolates, likely due to their volatility and susceptibility to nucleophilic attack by hydroxide ions. Similarly, (Z)-3-hexenal (*no.* 14), responsible for the fresh green note, decreased from 162.3 $\mu\text{g}/\text{kg}$ in the crude cold-pressed oil to 45.6 $\mu\text{g}/\text{kg}$ in the alkali-refined oil. Consequently, its odor activity value decreased across the processing stages (OAV = 54 vs 47 vs 37 vs 15), confirming its heat-lability and the corresponding loss of green odor. The cabbage-like odor driver, dimethyl trisulfide, also decreased from

Table 1. Flavor dilution factors of volatile flavor compounds in rapeseed oils subjected to different treatments via aroma extract dilution analysis.

No.	Odorant	CAS number	Odor quality	RI ^a	FD factors ^b			
					CCO ^c	TTC ^d	WDO ^e	ARO ^f
<i>Nitriles</i>								
1	3-Butenenitrile	109-75-1	Pungent, mustard-like	1176	64	64	32	8
2	4-Pentenenitrile	592-51-8	Pungent, metallic	1280	128	128	128	64
3	4-(Methylsulfanyl)butyl nitrile	59121-24-3	Broth-like, garlic-like, sulfuric	1721	64	64	32	16
4	5-(Methylsulfanyl)pentanenitrile	59121-25-4	Broccoli-like, cabbage-like	1845	16	16	8	4
5	Phenylacetone nitrile	140-29-4	Mushroom-like	1922	8	8	8	2
6	3-Phenylpropanenitrile	645-59-0	Sour, fatty	2051	4	4	4	4
<i>Aldehydes</i>								
7	Hexanal	66-25-1	Fatty, grassy, green	1078	32	32	64	64
8	Heptanal	111-71-7	Green, rancid	1192	8	8	16	16
9	Octanal	124-13-0	Citrus-like, fatty, soapy	1280	8	16	16	32
10	Nonanal	124-19-6	Citrus-like, soapy, waxy	1381	16	32	32	128
11	Decanal	112-31-2	Citrus-like, soapy, coriander-like	1492	4	8	16	64
12	Undecanal	112-44-7	Citrus-like, metallic	1592	< 1	2	4	16
13	Dodecanal	112-54-9	Floral, soapy	1690	< 1	< 1	< 1	4
14	(Z)-3-Hexenal	69112-21-6	Fresh green	1129	128	64	32	< 1
15	(E)-2-Heptenal	505-57-7	Fatty, almond	1220	8	16	16	32
16	(E)-2-Octenal	2548-87-0	Fatty, nutty, green	1420	8	16	16	32
17	(E)-2-Nonenal	18829-56-6	Cucumber-like, fatty	1527	4	8	16	64
18	(E,Z)-2,6-Nonadienal	557-48-2	Cucumber-like, fatty	1577	16	16	8	4
19	(E,E)-2,4-Nonadienal	5910-87-2	Deep-fried, fatty, green	1693	8	16	32	64
20	(E)-2-Decenal	3913-81-3	Tallowy, metallic	1629	4	8	16	32
21	(E,E)-2,4-Decadienal	25152-84-5	Deep-fried, fatty, flowery	1801	8	16	32	64
22	<i>Trans</i> -4,5-Epoxy-(E)-2-decenal	134454-31-2	Metallic	2003	< 1	4	8	256
23	3-Methylbutanal	590-86-3	Malty, dark chocolate	927	16	16	16	32
24	2-Methylbutanal	96-17-3	Malty, almond	931	16	16	16	32
25	Phenylacetaldehyde	122-78-1	Flowery, honey-like	1650	32	32	16	8
26	Benzaldehyde	100-52-7	Almond-like, burnt sugar	1512	4	4	4	8
<i>Ketones</i>								
27	1-Penten-3-one	1629-58-9	Pungent, fishy	1017	8	8	16	64
28	6-Methyl-5-hepten-2-one	110-93-0	Green, metallic	1340	4	4	2	2
29	1-Octen-3-one	4312-99-6	Mushroom-like, metallic	1294	8	16	32	128
30	(Z)-1,5-Octadien-3-one	65767-22-8	Geranium-like, metallic	1364	< 1	2	4	16
<i>Alcohols & Phenols</i>								
31	1-Octen-3-ol	3391-86-4	Mushroom-like	1435	16	16	8	2
32	(Z)-3-Hexen-1-ol	928-96-1	Cut grass, cucumber-like	1381	32	16	8	< 1
33	2-Methoxyphenol	90-05-1	Smoky, phenolic, sweet	1860	64	32	16	< 1
34	4-Methylphenol	106-44-5	Fecal, stable-like	2081	32	16	8	< 1
35	4-Vinylguaiaicol	7786-61-0	Spicy, clove-like, woody	2194	64	32	16	< 1
36	2-Phenylethanol	60-12-8	Flowery, honey-like	1909	8	8	4	< 1
<i>Pyrazines</i>								
37	2-Isopropyl-3-methoxypyrazine	25773-40-4	Earthy, potato-like	1426	4	4	4	2
<i>Volatile sulfur-containing compounds</i>								
38	Allyl isothiocyanate	57-06-7	Green, sulfury, pungent	1360	128	64	32	< 1
39	Dimethyl disulfide	624-92-0	Moldy, onion-like	1066	8	16	32	64
40	4-Isothiocyanato-1-butene	3386-97-8	Pickled, pungent, spicy	1453	32	16	8	< 1
41	Dimethyl trisulfide	3658-80-8	Cabbage-like, sulfury	1365	256	128	64	16
42	3-methylthiopropanal	3268-49-3	Cooked potato-like	1448	32	32	32	8
<i>Others</i>								
43	2-Pentylfuran	3777-69-3	Vegetable-like	1232	8	16	32	128
44	Acetic acid	64-19-7	Pungent, sour, vinegar-like	1444	64	64	32	< 1
45	Butanoic acid	107-92-6	Sweaty, cheese-like	1620	32	32	16	< 1
46	Octanoic acid	124-07-2	Soapy, spicy	2054	16	16	16	< 1

^a RI meant retention index calculated in this experiment. ^b FD meant flavor dilution factor. ^c CCO meant crude cold-pressed oil. ^d TTC meant thermally treated control oil. ^e WDO meant water-degummed oil. ^f ARO meant alkali-refined oil.

2.1 µg/kg (OAV = 70) in the crude oil to 1.2 µg/kg (OAV = 40) in the alkali-refined oil, consistent with the sensory reduction.

The metallic sensory defect may be elucidated through the formation of specific lipid peroxidation products. The quantitative data identified *trans*-4,5-epoxy-(E)-2-decenal (*no.* 22) as a potent odorant associated with this defect^[27]. In the crude cold-pressed oil, this compound was present at trace levels (0.50 µg/kg), resulting in an

OAV less than 1. However, the alkali refining process was associated with a significant accumulation, elevating its concentration to 8.50 µg/kg in the alkali-refined oil. Due to its extremely low odor threshold, this increase resulted in a rise in its odor activity value to 7. This metallic profile was further intensified by a group of vinyl ketones. (Z)-1,5-Octadien-3-one (*no.* 30), characterized by its geranium-like and metallic odor^[28], showed a substantial increase from

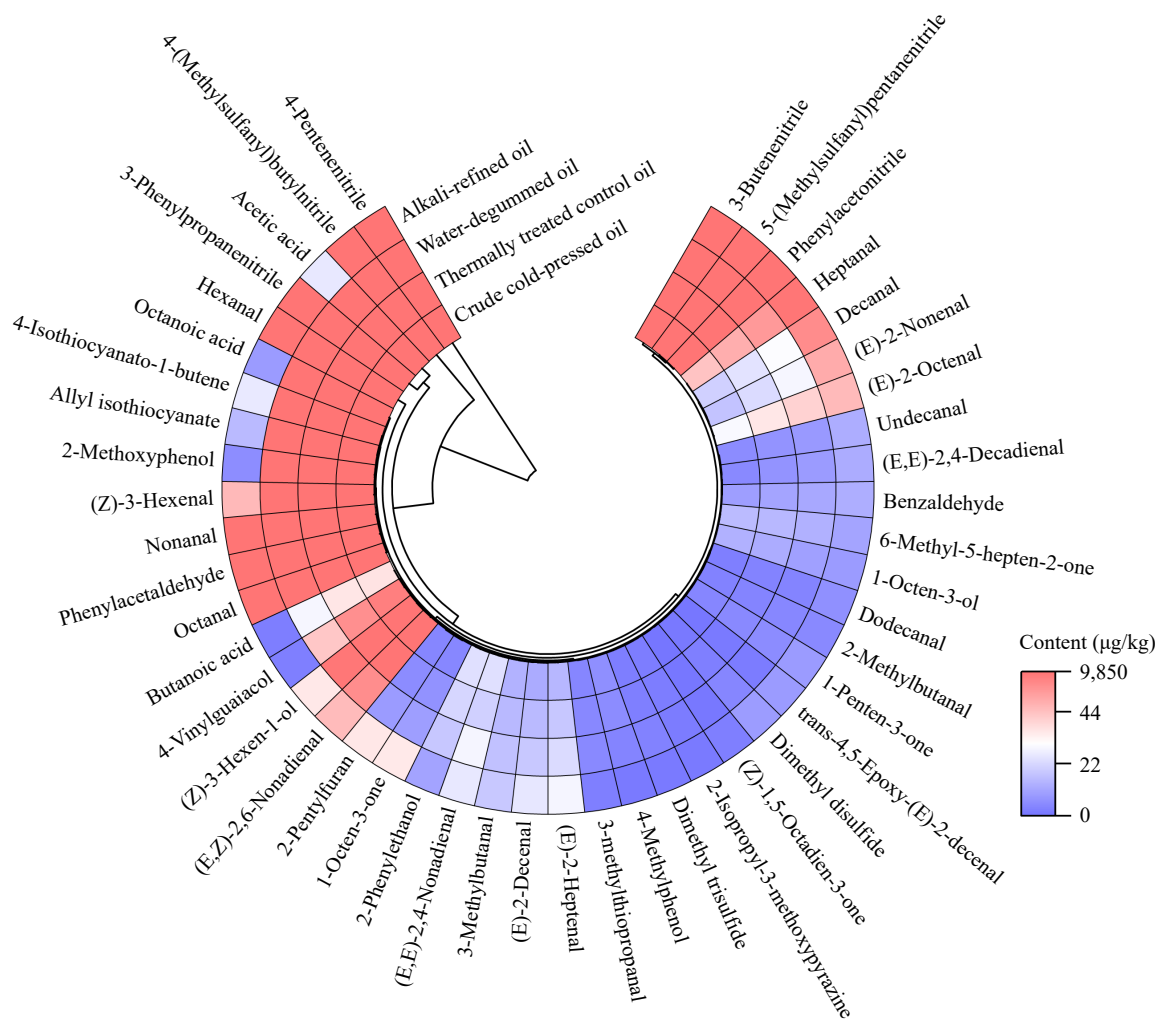


Fig. 2 Concentrations of volatile flavor compounds in crude cold-pressed, thermally treated, water-degummed, and alkali-refined rapeseed oils.

0.20 µg/kg in the crude oil to 2.50 µg/kg in the alkali-refined oil, raising its OAV from 2 to 19. 1-Octen-3-one (*no.* 29) also increased from 4.0 µg/kg to 35.2 µg/kg. Although its calculated OAV remained moderate (OAV = 0 to 4) based on the threshold used, its high FD factor (128) and significant concentration surge suggested it played a critical synergistic role. The concomitant rise of these potent compounds provides a molecular explanation for the metallic off-flavor perceived by the sensory panel.

Meanwhile, the quantitative analysis revealed that the soapy attribute was likely to increase in medium-chain saturated aldehydes. The concentration of nonanal (*no.* 10) exhibited a significant increase, reaching 256.8 µg/kg in the alkali-refined oil compared to 160.5 µg/kg in the crude cold-pressed oil. Similarly, decanal (*no.* 11) increased from 20.1 µg/kg in the crude oil to 55.4 µg/kg in the refined sample. While the calculated OAVs for decanal remained below 1, decanal reached an FD factor of 64, and nonanal reached 128 in the alkali-refined oil. This discrepancy between OAV and FD data suggested that the soapy perception was likely driven by the additive or synergistic effect of these homologous aldehydes, such as octanal, nonanal, decanal, and undecanal, rather than a single compound exceeding its threshold.

Importantly, volatile acids such as acetic acid and butanoic acid were removed during alkali refining. For example, acetic acid dropped from 1542.4 µg/kg in the crude oil to 25.5 µg/kg in the alkali-refined oil, reducing its OAV from 2 to less than 1. This

confirmed the efficacy of the deacidification process in removing free fatty acids and acidic volatiles. Furthermore, the earthy-odor 2-isopropyl-3-methoxypyrazine (*no.* 37) maintained a relatively constant trace concentration throughout the process (1.92 to 1.20 µg/kg), maintaining high OAVs (192 vs 120). Since pyrazines are stable, they act as a consistent background aroma and do not drive the differential off-flavors generated by the refining steps.

In summary, the quantitative analysis showed that the metallic off-flavors were attributed to lipid peroxidation products such as *trans*-4,5-epoxy-(E)-2-decenal and (Z)-1,5-octadien-3-one, which were formed during alkali treatment and exhibited significant OAV increases. Meanwhile, the soapy off-flavors were associated with the accumulation of medium-chain saturated aldehydes. Although their individual OAVs were low, their high FD factors and collective concentration implicate them as the contributors of the soapy off-flavor defect. These findings establish the targets for subsequent validation through aroma recombination experiments.

Aroma reconstitution and omission experiments

To validate the accuracy of the above results, an aroma recombination model was prepared using the authentic odorants at their concentrations determined in the alkali-refined oil^[29,30]. As presented in Fig. 3, the sensory profile of the recombination model was in good agreement with the original alkali-refined oil. The trained

Table 2. Odor activity values of key odorants in crude cold-pressed, thermally treated, water-degummed, and alkali-refined rapeseed oils.

Odorant	OT ($\mu\text{g}/\text{kg}$) ^a	OAV ^b				
		Crude cold-pressed oil	Thermally treated control oil	Water-degummed oil	Alkali-refined oil	
<i>Nitriles</i>	3-Butenenitrile	700	1	1	1	< 1
	4-Pentenenitrile	300	33	32	30	20
	4-(Methylsulfanyl) butylnitrile	2,000	2	2	2	1
	5-(Methylsulfanyl) pentanenitrile	50	6	6	5	2
	Phenylacetoneitrile	60	6	6	5	2
	3-Phenylpropanenitrile	500	2	2	2	2
<i>Aldehydes</i>	Hexanal	300	2	2	2	2
	Heptanal	250	< 1	< 1	< 1	< 1
	Octanal	56	2	2	2	4
	Nonanal	150	1	1	1	2
	Decanal	650	< 1	< 1	< 1	< 1
	Undecanal	5,000	< 1	< 1	< 1	< 1
	Dodecanal	3,000	< 1	< 1	< 1	< 1
	(Z)-3-Hexenal	3	54	47	37	15
	(E)-2-Heptenal	850	< 1	< 1	< 1	< 1
	(E)-2-Octenal	4	7	9	10	11
	(E)-2-Nonenal	150	< 1	< 1	< 1	< 1
	(E,Z)-2,6-Nonadienal	3.8	18	16	15	12
	(E,E)-2,4-Nonadienal	1.5	16	13	19	17
	(E)-2-Decenal	10	1	1	2	2
	(E,E)-2,4-Decadienal	180	1	1	2	3
	<i>Trans</i> -4,5-Epoxy-(E)-2-decenal	1.3	< 1	1	1	7
	3-Methylbutanal	5.4	3	3	3	3
	2-Methylbutanal	2.2	1	2	2	2
	Phenylacetaldehyde	83	2	2	2	2
	Benzaldehyde	60	< 1	< 1	< 1	< 1
<i>Ketones</i>	1-Penten-3-one	0.73	1	5	6	7
	6-Methyl-5-hepten-2-one	1,000	< 1	< 1	< 1	< 1
	1-Octen-3-one	10	< 1	1	1	4
	(Z)-1,5-Octadien-3-one	0.13	2	4	6	19
<i>Alcohols and Phenols</i>	1-Octen-3-ol	1	13	12	10	9
	(Z)-3-Hexen-1-ol	1,100	< 1	< 1	< 1	< 1
	2-Methoxyphenol	16	8	8	5	< 1
	4-Methylphenol	25	< 1	< 1	< 1	< 1
	4-Vinylguaiaicol	50	1	1	1	< 1
	2-Phenylethanol	211	< 1	< 1	< 1	< 1
<i>Pyrazines</i>	2-Isopropyl-3-methoxypyrazine	0.01	192	184	173	120
<i>Volatile sulfur-containing compounds</i>	Allyl isothiocyanate	30	5	4	3	1
	Dimethyl disulfide	12	< 1	< 1	< 1	1
	4-Isothiocyanato-1-butene	70	2	2	1	< 1
	Dimethyl trisulfide	0.03	70	83	93	40
<i>Others</i>	3-methylthiopropional	0.2	19	21	23	13
	2-Pentylfuran	100	5	4	4	1
	Acetic acid	750	2	2	1	< 1
	Butanoic acid	7.9	5	5	4	< 1
	Octanoic acid	3,000	< 1	< 1	< 1	< 1

^a Thresholds were derived from the book 'Compilations of odour threshold values in air, water and other media (Edition 2011)'^[39]. ^b OAVs were calculated by dividing the concentrations by the respective odor thresholds.

panel rated the soapy intensity of the model at 3.7 and the metallic intensity at 3.6, values which were statistically indistinguishable from the scores of 3.8 and 3.5 observed in the original alkali-refined oil ($p > 0.05$). The absence of significant differences across all attributes, including the background fatty and green odors, confirmed that the identified and quantified compounds represent the aroma profile of the alkali-refined oil.

To further clarify the contribution of specific flavor classes to the key defects, omission experiments were conducted (Table 3). A series of models was prepared by omitting one specific chemical class at a time from the full recombination model. Excluding the aldehyde fraction caused a highly significant decrease ($p < 0.001$) in

both soapy and metallic intensities. These results confirmed the dominance of aldehydes in the off-flavor profile, identifying nonanal and decanal as the drivers of the soapy off-flavor and *trans*-4,5-epoxy-(E)-2-decenal as the source of the metallic off-flavor.

Furthermore, the omission of ketones significantly lowered the metallic intensity ($p < 0.01$) but had no observed effect on the soapy note. This specific reduction aligned with the quantitative analysis, which identified vinyl ketones such as (Z)-1,5-octadien-3-one and 1-octen-3-one as potent metallic odorants that do not possess soapy characteristics. This result experimentally verified that the metallic off-flavor is a composite perception driven by them. In contrast, removing volatile sulfur compounds, nitriles, or alcohols resulted in

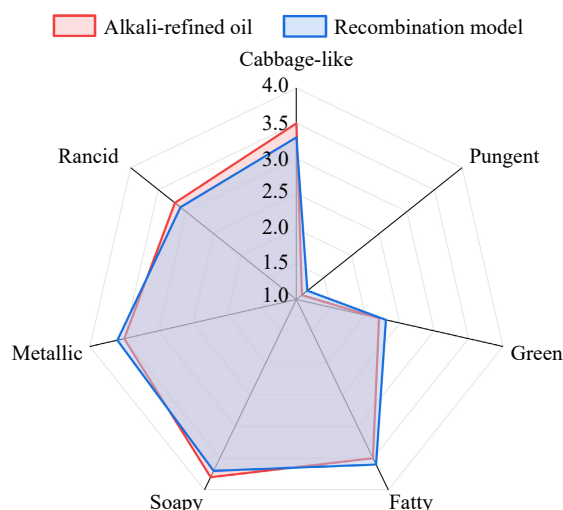


Fig. 3 Comparison of sensory profiles between alkali-refined rapeseed oil and its aroma recombination model.

Table 3. Significance of odorant groups on soapy and metallic off-flavors determined by aroma omission experiments.

No.	Omitted group	Soapy	Metallic
1	Aldehydes	****a	****a
2	Ketones	ns ^c	**b
3	Volatile sulfur-containing compounds	ns	ns
4	Nitriles	ns	ns
5	Pyrazines	ns	ns
6	Alcohols	ns	ns
7	Phenols	ns	ns
8	Acids	ns	ns
9	Isothiocyanates	ns	ns

^a $p < 0.001$. ^b $p < 0.01$. ^c Not significant.

no significant changes to the defect intensities. Although compounds such as dimethyl trisulfide were identified in the oil with high OAVs, their omission suggested they are responsible for the general background aroma derived from the raw seeds rather than the process-induced soapy and metallic off-flavors.

Discussion

Possible formation mechanisms of off-flavor formation

Based on established theories of vegetable oil flavor chemistry and the analytical data from this study, the metallic off-flavor characterized by *trans*-4,5-epoxy-(E)-2-decenal and vinyl ketones was primarily associated with the decomposition of linoleic and linolenic acid hydroperoxides^[31,32]. Specifically, the alkaline environment promotes the rearrangement of the linoleic acid 13-hydroperoxide isomer into the epoxy-aldehyde^[33]. Furthermore, the significant increase in 1-octen-3-one suggests the concurrent breakdown of the 9-hydroperoxide derived from linolenic acid, which is known to be unstable under basic conditions^[34,35].

Simultaneously, the soapy defect observed in the alkali-refined oil is likely derived from the autoxidation of oleic acid. Unlike polyunsaturated precursors that degrade readily under thermal stress, oleic acid hydroperoxides exhibit relatively higher thermal stability, which may explain their lower abundance in the thermally treated

control. However, the high pH environment significantly reduces the activation energy for their decomposition. This facilitates the breakdown of the 8- and 10-hydroperoxides, leading to the specific scission of the carbon chain that releases medium-chain saturated aldehydes, including nonanal and decanal^[36–38]. Regarding the soapy perception, it is worth noting that while the individual OAVs of decanal and nonanal were relatively low in the alkali-refined oil, their FD factors were high. This discrepancy suggests that the soapy off-flavor is not driven by a single compound exceeding its threshold, but rather by the additive or synergistic effects of octanal, nonanal, decanal, and undecanal.

Implications for industrial processing and study limitations

Several limitations of this work should be acknowledged. First, the refining process was simulated on a laboratory scale. While care was taken to replicate industrial parameters, the mass transfer rates and surface-to-volume ratios in a laboratory flask differ from those in industrial continuous refining towers, which could influence the exact yield of volatile oxidation products.

Second, this study focused on the identification of volatile odorants, whereas critical non-volatile factors were not quantified. Specifically, the reaction kinetics were likely influenced by the presence of trace metal ions and specific hydroperoxide isomers, neither of which was monitored in this study. Although the alkaline environment is the primary driver for off-flavor formation, transition metals such as iron and copper could act as co-catalysts, synergistically enhancing the decomposition of hydroperoxides even at low concentrations. Future studies should, therefore, incorporate the quantification of trace metals and hydroperoxide precursors and employ chelators to verify their specific contributions to the reaction pathways.

Third, the omission tests confirmed the role of aldehydes and ketones but did not investigate potential perceptual interactions between the off-flavors and the remaining trace amounts of native pungent compounds, which might act as masking agents in the raw material.

Despite these limitations, the findings offer actionable insights for the edible oil industry. The study identifies the alkali neutralization step as a critical control point for flavor quality. Reducing the residence time or lowering the temperature during the alkali contact phase could mitigate the formation of potent off-flavors. Monitoring the peroxide value alone may be insufficient for quality control, as the breakdown of peroxides into secondary odorants represents the actual source of the defect. Targeted monitoring of the identified key markers, specifically the epoxy-aldehyde and nonanal, would provide a more sensitive and relevant metric for assessing the sensory impact of the rapeseed oil refining process.

Conclusions

By applying a sensomics strategy that integrates quantitative descriptive analysis and aroma extract dilution analysis, this study identified the key odorants associated with soapy and metallic off-flavors in alkali-refined rapeseed oil. On the basis of the calculation of odor activity values and omission experiments, the metallic off-flavor was attributed to the formation of *trans*-4,5-epoxy-(E)-2-decenal, (Z)-1,5-octadien-3-one, and 1-octen-3-one. The soapy off-flavors were associated with the accumulation of medium-chain saturated aldehydes, particularly nonanal and decanal. Aroma recombination and omission experiments validated that aldehydes and ketones

constitute the primary flavor classes responsible for soapy and metallic off-flavors. The alkali neutralization step was identified as the critical control point for flavor quality. This work provides a theoretical basis for targeted process optimization and improvement of flavor and quality control in rapeseed oil refining.

Ethical statements

Ethical clearance for the involvement of human subjects in this study was obtained from the Research Ethics Committee of China Agricultural University, under reference number CAUHR-20240703.

Author contributions

The authors confirm contributions to the paper as follows: study conception and design: Wu J, Zhou Y, Shi H, Liu C, Xu X; data collection: Yu P, Lao F; analysis and interpretation of results: Yu P, Lao F, Wu J; draft manuscript preparation: Wang J. All authors reviewed the results and approved the final version of the manuscript.

Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

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Conflict of interest

The authors declare that they have no conflict of interest.

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References

- [1] Xu S, Huang D, Liu C, Gao Y, Li Q, et al. 2025. Development of nutrition-flavor dual process of rapeseed oil based on resource utilization of rapeseed cake. *Food Chemistry* 492:145576
- [2] Li H, Zhu H, Yao Q, Gao D, Wang L, et al. 2025. Composition, physico-chemical property and stability of canola seed oil: impact of different pretreatment. *Food Bioscience* 64:105933
- [3] Lv Y, Xiong Y, Luo S, Ye Z, Liu Y. 2025. Particulate matter and associated PAHs within the fumes emitted from heat-cooking rapeseed oils: focus on the refining level and trace components of rapeseed oil. *Journal of Agricultural and Food Chemistry* 73:14961–14972
- [4] Chu CC, Neo YP, Teng SK, Chew SC. 2025. Nutritional insights into rapeseed oil: biochemical composition, refining process and health implications. *Food Science and Biotechnology* 00:1–15
- [5] Chew SC. 2020. Cold-pressed rapeseed (*Brassica napus*) oil: chemistry and functionality. *Food Research International* 131:108997
- [6] Ye Z, Liu Y. 2023. Polyphenolic compounds from rapeseeds (*Brassica napus* L.): the major types, biofunctional roles, bioavailability, and the influences of rapeseed oil processing technologies on the content. *Food Research International* 163:112282
- [7] Wang W, Yang B, Huang F, Zheng C, Li W, et al. 2022. Synchronous pressing and refining after solid-phase preadsorption technology as a new method for rapeseed oil preparation. *LWT* 168:113939
- [8] Čmolík J, Schwarz W, Svoboda Z, Pokorný J, Réblová Z, et al. 2000. Effects of plant-scale alkali refining and physical refining on the quality of rapeseed oil. *European Journal of Lipid Science and Technology* 102:15–22
- [9] Matheis K, Granvogl M. 2016. Characterization of key odorants causing a fusty/musty off-flavor in native cold-pressed rapeseed oil by means of the sensomics approach. *Journal of Agricultural and Food Chemistry* 64:8168–8178
- [10] Zhang Y, Wu Y, Chen S, Yang B, Zhang H, et al. 2021. Flavor of rapeseed oil: an overview of odorants, analytical techniques, and impact of treatment. *Comprehensive Reviews in Food Science and Food Safety* 20:3983–4018
- [11] Li H, Zhang N, Qiang J, Cao Y, Qu G, et al. 2025. Impact of short-term storage of olive fruits on the quality of virgin olive oil. *Food Chemistry* 478:143640
- [12] Han L, Li G, Wang X, Yu B, Zhang T, et al. 2024. Characterization of volatile compounds from healthy and citrus black spot-infected Valencia orange juice and essential oil by using gas chromatography–mass spectrometry. *Food Chemistry: X* 22:101374
- [13] He Y, Liu Y, Sun J, Qiao M, Wang L, et al. 2025. Characterization of the key aroma-active compounds causing the Douchi aroma in high-temperature Daqu via integrated molecular sensory science, metabolomics, and molecular docking. *Food Chemistry* 463:141256
- [14] An K, Liu H, Fu M, Qian MC, Yu Y, et al. 2019. Identification of the cooked off-flavor in heat-sterilized lychee (*Litchi chinensis* Sonn.) juice by means of molecular sensory science. *Food Chemistry* 301:125282
- [15] Li H, Qin D, Wu Z, Sun B, Sun X, et al. 2019. Characterization of key aroma compounds in Chinese Guojing sesame-flavor Baijiu by means of molecular sensory science. *Food Chemistry* 284:100–107
- [16] Yu P, Wang J, Lao F, Shi H, Xu X, et al. 2025. Investigation on sweaty off-flavors in small mill sesame oil and its formation mechanism via molecular sensory science, preparative gas chromatography, and microbiomics. *Food Chemistry* 463:141224
- [17] Yu P, Luan Q, Wang J, Lao F, Shi H, et al. 2025. Investigation of the fruity and fermented off-flavors in peanut oil via molecular sensory science, S-curve method, and molecular docking. *Journal of Integrative Agriculture* 00:In Press, Journal Pre-proof
- [18] Sun G, Sun S, Zhang H, Zhang Y, Sun C, et al. 2025. Effect of roasting treatment on flavor and quality of fragrant corn oil. *Food Chemistry* 492:145426
- [19] Zhang Y, Stöppelmann F, Zhu L, Liang J, Rigling M, et al. 2023. A comparative study on flavor trapping techniques from the viewpoint of odorants of hot-pressed rapeseed oil. *Food Chemistry* 426:136617
- [20] Elbarbary A, Cui L, Wei L, Bakry IA, Huan H, et al. 2025. Characterization of natural flavor compounds of five commercial anhydrous milk fats via GC-TOF-MS, aroma recombination, and omission models. *Journal of Food Composition and Analysis* 140:107231
- [21] Zhou Q, Zheng C, Wei F, Yang Y. 2024. Flavor precursors identification and thermal degradation mechanisms of glucoerucin in fragrant rapeseed oil. *Food Chemistry* 435:137484
- [22] Ren X, Wang L, Xu B, Wei B, Liu Y, et al. 2019. Influence of microwave pretreatment on the flavor attributes and oxidative stability of cold-pressed rapeseed oil. *Drying Technology* 37:397–408
- [23] Liang Q, Xiong W, Zhou Q, Cui C, Xu X, et al. 2023. Glucosinolates or erucic acid, which one contributes more to volatile flavor of fragrant rapeseed oil? *Food Chemistry* 412:135594

- [24] Wagner C, Bonte A, Brühl L, Niehaus K, Bednarz H, et al. 2018. Microorganisms growing on rapeseed during storage affect the profile of volatile compounds of virgin rapeseed oil. *Journal of the Science of Food and Agriculture* 98:2147–2155
- [25] Liu J, Gu P, Chu B, Hu B, Zhang Y, et al. 2025. Decoding pickle aroma in rapeseed oil by sensomics, gas chromatography-olfactometry, quantitation, addition studies and molecular docking. *Food Chemistry* 489:145008
- [26] Zhou Q, Tang H, Jia X, Zheng C, Huang F, et al. 2018. Distribution of glucosinolate and pungent odors in rapeseed oils from raw and microwaved seeds. *International Journal of Food Properties* 21:2296–2308
- [27] Neugebauer A, Schieberle P, Granvogl M. 2021. Characterization of the key odorants causing the musty and fusty/muddy sediment off-flavors in olive oils. *Journal of Agricultural and Food Chemistry* 69:14878–14892
- [28] Schmidberger PC, Schieberle P. 2020. Changes in the key aroma compounds of raw shiitake mushrooms (*Lentinula edodes*) induced by pan-frying as well as by rehydration of dry mushrooms. *Journal of Agricultural and Food Chemistry* 68:4493–4506
- [29] Xu L, Chang J, Mei X, Zhang Y, Wu G, et al. 2022. Comparative analysis of aroma compounds in French fries and palm oil at three crucial stages by GC/MS-olfactometry, odor activity values, and aroma recombination. *Journal of the Science of Food and Agriculture* 102:2792–2804
- [30] Schulze LJ, Schäfer U, Beier R, Hartmann B, Wüst M, et al. 2024. Molecular-sensory decoding of the *Citrus latifolia* aroma. *Journal of Agricultural and Food Chemistry* 72:14874–14886
- [31] Oakley LH, Casadio F, Shull KR, Broadbelt LJ. 2018. Examination of mechanisms for formation of volatile aldehydes from oxidation of oil-based systems. *Industrial & Engineering Chemistry Research* 57:139–149
- [32] Zhang Q, Ke J, Long P, Wen M, Han Z, et al. 2024. Formation of furan from linoleic acid thermal oxidation: (E,E)-2,4-decadienal as a critical intermediate product. *Journal of Agricultural and Food Chemistry* 72:4384–4392
- [33] Wang B, Wang S, Wang Y, Zhang S, Lin X, et al. 2023. Deep exploration of lipid oxidation into flavor compounds: a density functional theory study on (E)-2-decenal thermal oxidative reaction. *Food Chemistry* 428:136725
- [34] Matsui K, Takemoto H, Koeduka T, Ohnishi T. 2018. 1-Octen-3-ol is formed from its glycoside during processing of soybean [*Glycine max* (L.) Merr.] seeds. *Journal of Agricultural and Food Chemistry* 66:7409–7416
- [35] Assaf S, Hadar Y, Dosoretz CG. 1997. 1-Octen-3-ol and 13-hydroperoxylinoleate are products of distinct pathways in the oxidative breakdown of linoleic acid by *Pleurotus pulmonarius*. *Enzyme and Microbial Technology* 21:484–490
- [36] Xiao L, Wang S, Wang Y, Wang B, Ji C, et al. 2023. Density functional theory studies on the oleic acid thermal oxidation into volatile compounds. *Food Chemistry: X* 19:100737
- [37] Shi Y, Li J, Zhou L, Zhang J, Feng X, et al. 2025. Exploring the contribution of phosphatidylcholine and triglyceride on the formation of beef aroma-active compounds with thermal oxidation system. *Current Research in Food Science* 10:100973
- [38] Liu X, Wang W, Li Z, Xu L, Lan D, et al. 2024. Lipidomics analysis unveils the dynamic alterations of lipid degradation in rice bran during storage. *Food Research International* 184:114243
- [39] van Gemert L. 2011. *Compilations of odour threshold values in air, water, and other media*. Edition 2011. Zeist: Oliemans Punter & Partners



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