

Bioactive peptide-enriched deer meat sauce: a novel functional food innovation

Kana Kogiso^{1,2*} and Kazumasa Furuta³

¹ Faculty of Health and Human Development, The University of Nagano, 8-49-7 Miwa, Nagano City, Nagano 380-8525, Japan

² Graduate School of Health and Nutrition Science, The University of Nagano, 8-49-7 Miwa, Nagano City, Nagano 380-8525, Japan

³ BioSpin Division, Bruker Japan K.K, 3-9 Moriya-cho, Kanagawa-ku, Yokohama City, Kanagawa 221-0022, Japan

* Corresponding author, E-mail: kogiso.kana@u-nagano.ac.jp

Abstract

The development of functional soy sauce-like condiments using wild game (deer meat) represents a new approach to enhancing the value of underutilized resources. This study develops an ultra-short-term fermented meat sauce from deer meat using high hydrostatic pressure (HHP) treatment within 26 h. The study also aimed to evaluate its functionality, particularly angiotensin-converting enzyme (ACE) inhibitory activity and bioactive peptide composition, and to assess its bioactivity in nematodes. The HHP-treated deer meat sauce was compared with conventional soy sauce to explore its potential health benefits. The results demonstrated that the HHP-treated sauce significantly increased the concentrations of bioactive imidazole peptides, especially anserine and carnosine. Additionally, a dipeptide with antihypertensive properties was newly identified in the HHP-treated deer meat sauce through LC-MS analysis. The ACE inhibitory activity of the deer meat sauce was eight times higher than that of conventional soy-sauce, indicating potential antihypertensive effects. Furthermore, nematodes (*Caenorhabditis elegans*) fed with the HHP-treated deer meat sauce exhibited notable bioactivity. These findings suggest that HHP-treated deer meat sauce can serve as a functional food ingredient for health promotion and fatigue reduction.

Citation: Kogiso K, Furuta K. 2025. Bioactive peptide-enriched deer meat sauce: a novel functional food innovation. *Food Materials Research* 5: e010 <https://doi.org/10.48130/fmr-0025-0009>

Introduction

The overpopulation of sika deer (*Cervus nippon*) in Japan has caused significant wildlife damage, highlighting the need for effective utilization of deer meat as a sustainable food resource^[1]. Despite its potential, deer meat consumption remains limited due to its distinct odor and tougher texture compared to other livestock meats^[2,3]. Addressing these challenges could enhance the value of wild game meat while meeting the growing consumer demand for healthy food options, as wild game meat is high in protein, low in calories, and widely regarded as health-promoting^[4,5].

Previous studies have shown that protease treatment combined with high hydrostatic pressure (HHP) can liquefy deer and wild boar meat, increasing the levels of bioactive compounds such as anserine and umami-enhancing amino acids like *L*-glutamic acid and *L*-aspartic acid^[6]. HHP, a nonthermal high-pressure technology, inactivates pathogens and ensures food safety^[7]. It facilitates protein denaturation and enzymatic hydrolysis, thereby releasing bioactive peptides^[8]. However, the use of HHP for meat liquefaction has not been extensively explored.

Developing an economical HHP system to liquefy wild game meat could address issues related to odor and toughness within a relatively short processing time of 26 h. Currently, only large cuts of wild game meat, such as loin and thigh, are commonly utilized^[9]. Liquefaction technology would enable the effective use of smaller and less commercially valuable parts, such as front legs, by converting them into seasonings and other products, thereby reducing waste.

This method parallels traditional Japanese seasonings, such as soy sauce, which is produced through the fermentation of protein-rich soybeans, yielding a product rich in peptides and amino acids with bioactive properties^[10–12]. Similarly, creating a soy sauce-like seasoning from liquefied venison could confer functional properties

associated with bioactive peptides. The extract is particularly rich in imidazole peptides such as anserine and carnosine, which are known for their antioxidant, anti-glycation, anti-fatigue, and angiotensin-converting enzyme (ACE) inhibitory activities^[13–15]. Enhancing the concentration of these peptides in food products may provide additional health benefits.

In a previous study, a comprehensive safety and sensory evaluation of HHP-processed deer meat sauce was conducted^[16]. Safety assessments—including viable bacterial counts, lactobacillus levels in fermented meat, and renal function tests—confirmed that the product met food safety standards. Sensory analysis revealed that the HHP-processed deer meat sauce possessed a taste profile similar to soy sauce, making it suitable for human consumption. Using a taste-recognition device, it was determined that the venison-based meat sauce was less salty and acidic than conventional soy sauce. To enhance its flavor, adjustments were made by adding salt, lactic acid, and other ingredients. Sensory evaluations of the seasoned meat sauce indicated that, while its color and aroma differed from soy sauce, its levels of umami, acidity, and saltiness were comparable. These findings highlight the potential of HHP-processed deer meat sauce as a versatile and highly palatable alternative to conventional seasonings.

Caenorhabditis elegans (*C. elegans*) is a well-established model organism for studying aging and stress responses due to its short lifespan, conserved metabolic pathways, and ease of genetic manipulation^[17–19]. With a lifespan of approximately 20 d and straightforward cultivation requirements, it serves as an ideal model for analyzing the effects of dietary interventions on health and longevity.

In this study, a soy sauce-like seasoning was developed using deer meat processed with HHP treatment. This research aimed to evaluate its potential health and longevity in *C. elegans* by

increasing the concentration of bioactive peptides and ACE inhibitory activity in the deer meat sauce. Additionally, the sensory properties of the HHP-treated deer meat sauce were explored, building on previous findings, to assess its potential as a functional food product acceptable to consumers.

Materials and methods

Materials

Commercial sika deer (*Cervus nippon*) back loin meat was sourced from a local wild game processing facility, stored at -20°C , and thawed under running water before use. Protin SD-NY10 (Amano Enzyme Co., Ltd) was prepared as a 0.75% (w/w) solution to serve as the protease for enzymatic hydrolysis. Water, equivalent to 10% of the meat weight, was added to the mixture to facilitate the hydrolysis process.

High hydrostatic pressure treatment

The thawed deer meat was minced using a commercial meat grinder and thoroughly mixed with the enzyme solution and water. The mixture was subjected to HHP treatment at 200 MPa and 50°C for 24 h using a high-pressure processing apparatus (Toyo High Pressure Co., Ltd). These conditions were selected based on previous optimization studies for peptide release^[6]. The resulting product was designated as HHP-treated deer sauce.

To sterilize the samples and inactivate residual enzyme activity, the treated mixture was heated in a water bath at 100°C for 30 min. This heating process denatures proteins and inactivates enzymes without significantly affecting imidazole peptides such as anserine and carnosine^[20]. Therefore, the concentrations of these peptides were not expected to change substantially due to the heating. The sauce was produced in less than 26 h including sterilization. It was produced using only the HHP enzymatic hydrolysis process without fermentation by microorganisms.

Analysis of imidazole peptides and umami amino acids

Amino acids associated with umami taste (L-glutamic acid and L-aspartic acid) and imidazole peptides (anserine and carnosine) were analyzed as follows: Ten grams of the liquefied sample were diluted tenfold with a 2.0% (w/v) sulfosalicylic acid solution. The mixture was homogenized at 600 rpm for 1 min, then centrifuged at 3,500 rpm for 10 min at 4°C . The supernatant was filtered through a $0.45\text{ }\mu\text{m}$ cellulose acetate filter.

Filtered samples were analyzed using a free amino acid analyzer (L-8900 Amino Acid Analyzer, Hitachi High-Technologies Corporation, Tokyo, Japan), as described by Yamazaki et al.^[21]. Ten microliters of each extract were injected into the analyzer, and the peak intensities of amino acids and peptides in the chromatograms were quantified by comparison with standard solutions.

Analysis of peptide composition

Sample preparation

Two samples were prepared for liquid chromatography-mass spectrometry (LC-MS) analysis: HHP-treated deer sauce and untreated deer meat extract. The untreated extract was prepared by suspending raw deer meat in five volumes of distilled water, followed by filtration through a $0.45\text{ }\mu\text{m}$ cellulose acetate filter.

LC-MS analysis

The filtered samples were prepared at concentrations suitable for LC-MS analysis. The HHP-treated deer sauce was diluted to a 0.6% (w/v) solution, while the untreated deer meat extract was prepared at a 3.0% (w/v) solution to account for the expected lower peptide content.

A high-performance liquid chromatography system coupled with a Xevo QT of MS (Waters Corporation) was used for the analysis. Chromatographic separation was performed using a GL Sciences Intersil ODS-3 column ($3.0\text{ mm} \times 250\text{ mm}$, inner diameter) at a column temperature of 40°C , with the sample compartment maintained at 10°C and a flow rate of 0.5 mL/min.

The mobile phase comprised solvent A (water with 0.1% [v/v] formic acid) and solvent B (acetonitrile with 0.1% [v/v] formic acid). Gradient elution was initiated with 90% solvent A and 10% solvent B, gradually increasing solvent B to 60% over 30 min. Mass spectrometry was conducted in positive ion mode, following the methods detailed by Alelyunas et al.^[22].

Forty-two antihypertensive dipeptides identified in previous studies^[23–27] were selected, and their molecular structures were used to calculate the expected mass-to-charge ratios (m/z). LC-MS data were analyzed using ITA software (version 14.04.79336) following the optimized procedures described by Kogiso et al.^[28]. Molecular ion peaks corresponding to the calculated m/z values were identified, and those matching the antihypertensive dipeptides were annotated.

Determination of ACE inhibitory activity

ACE inhibitory activity was assessed *in vitro* using the ACE Kit-WST (Dojindo Laboratories), following the manufacturer's instructions and the methods described by Lam et al.^[29,30]. The results were expressed as the sample concentration required to inhibit 50% of ACE activity (IC_{50}).

Model organism experiments

Caenorhabditis elegans culture

The wild-type strain *C. elegans* var. Bristol (N2) was used in all experiments. The Bristol N2 strain was obtained from the *Caenorhabditis* Genetics Center (CGC), which is funded by the NIH Office of Research Infrastructure Programs (P40 OD010440). Worms were maintained on nematode growth medium (NGM) agar plates seeded with *Escherichia coli* OP50 at 20°C . Reagents and media were prepared according to the standard protocols established by Brenner^[31]. The plates containing the nematodes were maintained under atmospheric conditions, with oxygen replenished at five-day intervals and a constant amount of food supplied weekly. Each plate and well was placed in an identical environment, minimizing variation between groups and striving to be as uninvolved as possible in the lifespan of the nematodes.

It has been reported that utilizing fewer than 15 nematodes per well can minimize counting errors while preventing overcrowding^[32]. To ensure optimal conditions and facilitate precise tracking, six to ten synchronized nematodes were placed in each well.

Treatment groups

Worms were divided into three groups:

Control Group 1: Fed a standard diet of *E. coli* OP50.

Control Group 2: Fed the standard diet supplemented with a hot water extract of deer meat. The extract was prepared at a concentration of 100 mg/mL, with 1 mL added to 20 mL of medium, resulting in a final concentration of 5 mg/mL. This provided anserine at $15.54\text{ }\mu\text{g/mL}$ ($0.06467\text{ }\mu\text{mol/mL}$) and carnosine at $15.14\text{ }\mu\text{g/mL}$ ($0.06692\text{ }\mu\text{mol/mL}$).

HHP Group 3: Fed the standard diet supplemented with HHP-treated deer sauce. The sauce was prepared at a concentration of 100 mg/mL, with 1 mL added to 20 mL of medium, yielding a final concentration of 5 mg/mL. This provided anserine at $45.6\text{ }\mu\text{g/mL}$ ($0.1898\text{ }\mu\text{mol/mL}$) and carnosine at $15.92\text{ }\mu\text{g/mL}$ ($0.07039\text{ }\mu\text{mol/mL}$). All extracts were sterilized by autoclaving at 121°C for 15 min before being added to the culture medium.

Lifespan measurement

To synchronize worm populations, eggs were collected and incubated in an M9 buffer for 18 h to obtain L1 larvae. Approximately six to ten synchronized worms were placed in each well of a 24-well plate containing the designated treatment. To prevent progeny from interfering with lifespan measurements, 5-fluoro-2'-deoxyuridine (FUDR) was added to the medium at a final concentration of 50 μM [33].

Worms were monitored every 2–3 d using a stereomicroscope (SZX7, Olympus Corporation). Worms that responded to stimuli, such as gentle touch or light exposure, were recorded as alive, while those showing no response were considered dead.

Assessment of anti-fatigue effects

The anti-fatigue effects were assessed by measuring the distance traveled by worms under induced stress conditions. Synchronized worms were cultured on agar plates without food or on plates supplemented with HHP-treated deer sauce, providing an anserine equivalent of 0.5 $\mu\text{mol/g}$ for 3 d.

Fatigue was induced by centrifuging the worms, washing them, and incubating them in a liquid medium without food for 24 h. Subsequently, the worms were subjected to additional stress by swimming in the M9 buffer for 2 h [34]. After stress induction, the worms were placed on agar plates. A drop of 1% diacetyl solution, a known chemoattractant, was placed 6.5 cm away from the worms. The distance moved toward the diacetyl source over 10 min was measured, as described by Ramot et al. [35].

Statistical analysis

All data are presented as the mean with standard deviation (SD). Statistical analyses were conducted using JMP software (version 14.3.0, SAS Institute Inc.). For multiple group comparisons, one-way analysis of variance (ANOVA) followed by Tukey's post hoc test was applied.

Survival analyses were performed using the Kaplan-Meier method, with differences between survival curves evaluated using the log-rank test. Statistical analyses for survival data were performed using EZR [36], a graphical user interface for R (The R Foundation for Statistical Computing). A *p*-value of less than 0.05 was considered statistically significant.

Results

Imidazole peptide concentrations

The concentrations of the imidazole peptides anserine and carnosine in both deer meat extract and HHP-treated deer meat sauce

were determined using an amino acid analyzer. In the untreated deer meat extract, the concentrations of anserine and carnosine were 777 and 757 mg/100 g, respectively. However, the HHP-treated deer meat sauce exhibited significantly higher concentrations, with 2,280 mg/100 g of anserine and 797 mg/100 g of carnosine. Following HHP treatment, the concentration of anserine increased nearly threefold, while carnosine showed a moderate increase. These results indicate that HHP treatment effectively enhances the extraction and/or formation of bioactive imidazole peptides from deer meat. The standard error between batches for these components was within 10%.

Peptide composition analysis

LC-MS analysis was performed to identify dipeptides with known antihypertensive effects in both the deer meat extract and the HHP-treated deer meat sauce. In the untreated deer meat extract, the dipeptides phenylalanyl-proline (FP) and arginyl-tryptophan (RW) were detected. However, in the HHP-treated deer meat sauce, several additional antihypertensive dipeptides were identified, including alanyl-tryptophan (AW), isoleucyl-tyrosine (IY), leucyl-tyrosine (LY), tyrosyl-leucine (YL), and valyl-tyrosine (VY) (Fig. 1). These dipeptides, which were either absent or present at lower levels in the untreated extract, suggest that HHP treatment enhances the generation of bioactive peptides with potential health benefits.

ACE inhibitory activity

The ACE inhibitory activity of the HHP-treated deer meat sauce was evaluated and compared with that of other commercially available fermented foods (Table 1). The IC_{50} value for the HHP-treated deer meat sauce was 201 $\mu\text{g/mL}$, demonstrating strong ACE inhibitory activity. In comparison, the IC_{50} values of most other foods were 4 to 45 times lower, with a few exceptions [37–41]. Therefore, the ACE inhibitory activity of the HHP-treated deer meat sauce is approximately eight times higher than that of dark soy sauce. This suggests that the HHP-treated deer meat sauce has the potential to be developed as a functional seasoning with antihypertensive properties.

C. elegans lifespan extension

The effect of deer meat extracts on nematode longevity was assessed through survival curve analysis (Fig. 2). Each treatment group included 20–53 worms. The median survival of the untreated control group was 14 d. Worms supplemented with deer meat extract exhibited a median survival of 17 d, representing a

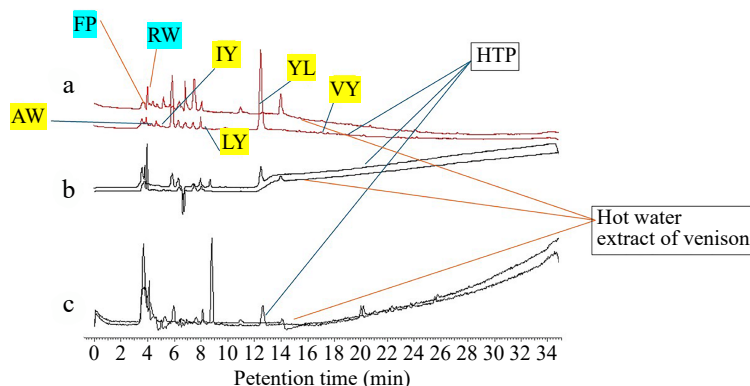


Fig. 1 Detection of dipeptides with antihypertensive activity in deer meat extracts and HHP-treated deer meat sauce. Peaks corresponding to specific dipeptides are indicated. From top to bottom: (a) UV chromatogram at 280 nm, (b) total absorption chromatogram (TAC) obtained using a diode-array detector (DAD), and (c) the ion chromatogram at $m/z = 227.1$. Lines indicate peaks assigned to antihypertensive dipeptides. HHP treatment resulted in increased levels of these dipeptides compared with untreated deer meat extract. Details of the analytical conditions are provided in the materials and methods. These data represent typical examples from at least independently conducted experiments.

Table 1. Angiotensin I-converting enzyme (ACE) inhibitory activity of various fermented and meat products.

Food name	Type	Ingredient type	IC ₅₀ value (μg/mL)	Ref.
HHP-treated deer meat sauce	HHP-treated meat	Mixed peptides (unidentified)	201	Present study
Japanese soy sauce	Fermented soybean	Mixed peptides (unidentified)	1,620	[37]
Chinese soy sauce	Fermented soybean	Mixed peptides (unidentified)	740-3,030	[38]
Natto (traditional Japanese fermented soybeans)	Fermented soybean	Polymer-blended peptide (mucus fraction)	120-950	[39]
Tofu-yō (traditional Japanese fermented tofu food)	Fermented Tofu	Crude extract (unidentified)	1,770	[40]
Livestock meat (beef, pork, chicken)	Raw meat	Mixed peptides (unidentified)	Calculated as beef (9,020); pork (4,190); chicken (3,550)	[41]

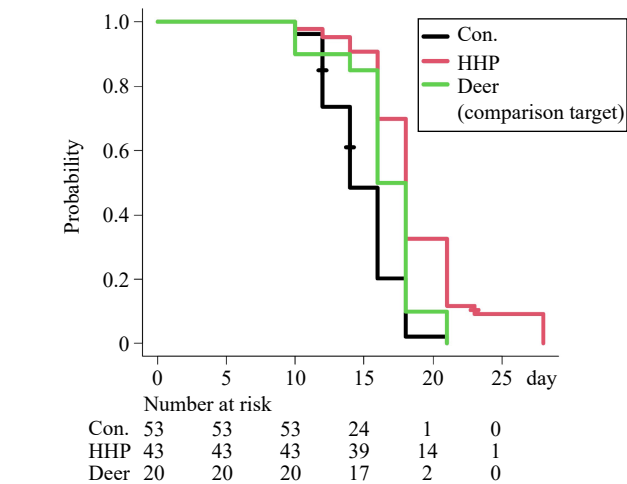


Fig. 2 Lifespan of worms supplemented with deer meat extract and HHP-treated deer meat extract. The figure shows the survival curves for *C. elegans* across three groups: worms without supplementation (control, black line), worms supplemented with deer meat extract (green line), and worms supplemented with HHP-treated deer meat sauce (red line). The Kaplan-Meier survival curves for *C. elegans* (wild-type N2) at 20 °C are shown. The experiments were repeated in three independent experiments. Statistical significance was determined by the log-rank test ($p < 0.05$).

significant extension compared to the control group ($p = 0.0057$). In comparison, worms supplemented with HHP-treated deer meat sauce showed a median survival of 18 d, significantly longer than the control group ($p < 0.0001$) and the deer meat extract group ($p = 0.0241$). These findings indicate that HHP-treated deer meat sauce is significantly more effective at enhancing nematode longevity than untreated deer meat extract.

Anti-fatigue effects in *C. elegans*

The anti-fatigue properties of the HHP-treated deer meat sauce were evaluated by measuring the migration distance of *C. elegans* under induced stress conditions. Worms supplemented with the HHP-treated deer meat sauce demonstrated a significantly greater average migration distance (40.3 ± 14.5 mm) compared to those without supplementation (21.9 ± 14.7 mm, $p = 0.0249$) (Fig. 3).

Worms in the HHP-supplemented group displayed an enhanced ability to move toward the chemoattractant (1% diacetyl), even under fatigue-inducing conditions. This improvement in physical performance highlights the potential of HHP-treated deer meat sauce to mitigate fatigue and enhance recovery.

Discussion

The development of a soy sauce-like seasoning from deer meat using HHP significantly enhanced its functional properties. The

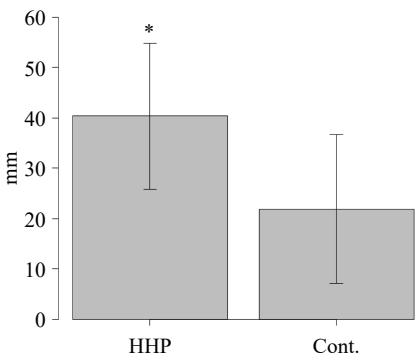


Fig. 3 Migration distances of *C. elegans* under fatigue conditions with and without supplementation of HHP-treated deer meat sauce. Data are presented as mean \pm SD. $p < 0.05$ indicates statistical significance between groups.

HHP-treated deer meat sauce demonstrated increased levels of bioactive peptides, including anserine, carnosine, and several antihypertensive dipeptides. These enhancements were associated with notable biological activities, including ACE inhibitory effects, extended lifespan, and improved anti-fatigue performance in *C. elegans*.

LC-MS analysis revealed several antihypertensive dipeptides in the HHP-treated deer meat sauce that were absent or present at low concentrations in the untreated deer meat extract. Notably, dipeptides, such as AW, IY, LY, YL, and VY were newly detected after HHP treatment. Among these, tryptophan-containing peptides such as AW, have been reported to exhibit strong ACE inhibitory activity^[42]. Similarly, tyrosine-containing dipeptides are known for their potent ACE inhibitory effects^[43,44]. This enhancement in bioactive peptide content corresponds with the significantly lower IC₅₀ values observed for HHP-treated deer meat sauce compared to commercial dark soy sauce, indicating that HHP treatment effectively boosts the concentration of peptides with potential antihypertensive properties.

The increased levels of dipeptides, as well as anserine and carnosine, in the HHP-treated deer meat sauce, likely contribute significantly to the observed biological activities in *C. elegans*. Numerous studies have demonstrated that specific peptide supplementation can extend the lifespan of *C. elegans* by influencing key metabolic and stress response pathways. For example, elevated tyrosine levels have been shown to affect developmental and longevity pathways in *C. elegans*. Ferguson et al.^[45] reported that tyrosine supplementation extended the lifespan of *C. elegans* through modulation of the insulin/IGF-1 signaling pathway. This effect was dependent on the activity of genes such as *daf-16* and *aak-2*, which encode a FOXO transcription factor and AMP-activated protein kinase, respectively—both critical regulators of stress response and metabolic homeostasis.

Similarly, tryptophan-derived peptides have exhibited neuro-protective and anti-aging effects in *C. elegans*. Manzanares et al.^[46] demonstrated that these peptides mitigate β -amyloid toxicity and inhibit amyloid aggregation—key factors implicated in age-related decline. These findings suggest that dietary peptides can positively influence protein homeostasis and neuronal health, contributing to increased lifespan and improved organismal resilience.

Carnosine and anserine, abundant in the HHP-treated deer meat sauce, are well-known for their antioxidant properties, ability to reduce reactive oxygen species (ROS), and modulation of stress response pathways^[47]. While direct studies on the effects of carnosine and anserine on lifespan extension in *C. elegans* remain limited, their mechanisms align with key longevity pathways in the organism. These dipeptides may enhance stress resistance by activating genes such as daf-16, which regulate the expression of antioxidant enzymes and protective proteins^[48].

In humans, carnosine and anserine are transported by proton-coupled oligopeptide transporters PEPT1 and PEPT2, which are important for their bioavailability and physiological effects^[49]. Similarly, *C. elegans* possesses a homologous transporter, PEPT-1, which is integral to nutrient uptake and lifespan regulation^[50]. The enhanced uptake of these dipeptides via PEPT-1 could modulate key metabolic and stress response pathways, potentially influencing longevity mechanisms and contributing to the observed lifespan extension.

Furthermore, the activation of stress response genes, such as daf-16, is a well-established mechanism for lifespan extension in *C. elegans*^[51].

Compounds that enhance DAF-16 activity lead to increased stress resistance and longevity. Dietary interventions that increase bioactive peptides, such as anserine and carnosine, may support healthspan by stimulating these conserved pathways, promoting enhanced resilience to metabolic and oxidative stress. These pathways are also consistent with the mechanism of action of captopril, one of the previously reported peptide-like components^[52], and the mechanism of neuro-metabolic regulation by dietary peptides^[53].

Although direct evidence of carnosine and anserine extending lifespan in *C. elegans* is limited, their well-established mechanisms—antioxidant activity, ROS reduction, and modulation of stress response pathways—support the hypothesis that these dipeptides contribute to the observed longevity effects. These findings align with the hypothesis, suggesting that the elevated levels of anserine and carnosine in the HHP-treated deer meat sauce enhance stress resistance and activate longevity-associated pathways in *C. elegans*.

Comparing the ACE inhibitory activity of the HHP-treated deer meat sauce with dark soy sauce further underscores its potential as a functional seasoning. The IC₅₀ value of 201 $\mu\text{g/mL}$ for the HHP-treated sauce demonstrates significantly stronger inhibitory effects than dark soy sauce, which has an IC₅₀ value of 1,620 $\mu\text{g/mL}$ ^[37]. A lower IC₅₀ value indicates greater potency in inhibiting ACE activity, a key factor in blood pressure regulation. These findings suggest that incorporating HHP-treated deer meat sauce into the diet could provide additional benefits for hypertension management, highlighting the potential of functional foods enriched in bioactive peptides as a complementary strategy for promoting cardiovascular health and managing hypertension.

The use of HHP treatment was effective in promoting peptide release without compromising food safety. HHP is recognized for its ability to inactivate pathogens and enzymes while preserving the nutritional quality and sensory attributes of foods^[7,8]. This method enables the utilization of less commercially valuable parts of wild game, such as front legs, contributing to waste reduction and sustainable food production. By converting underutilized meat parts

into value-added products, HHP treatment supports environmental sustainability and enhances resource efficiency.

These findings suggest that the HHP-treated deer meat sauce has significant potential as a novel functional food ingredient with anti-hypertensive and anti-fatigue properties.

Specifically, because of the antioxidant, pH buffering, and neuro-protective effects of carnosine and anserine, venison sauce has the potential to be used as a nutritional supplement for the elderly, as a meal for people receiving care, to aid recovery from fatigue after exercise and to support cognitive function. Furthermore, as this extract contains dipeptides with ACE inhibitory activity, it is also expected to be used as a functional seasoning or food supplement that can be used to prevent and manage lifestyle diseases such as hypertension and metabolic disorders. The observed biological activities in *C. elegans* provide a basis for further exploration of the health benefits of bioactive peptides derived from wild game meats. The use of *C. elegans* as a model organism offers advantages due to its conserved metabolic pathways and relevance to higher organisms in studies of aging and stress responses^[17,18].

Conclusions

This study successfully developed a soy sauce-like seasoning from deer meat using HHP, significantly enhancing its functional properties. The HHP-treated deer meat sauce exhibited increased levels of bioactive peptides, specifically anserine and carnosine, alongside newly detected antihypertensive dipeptides. These biochemical enhancements were associated with notable biological activities, including lifespan extension and fatigue recovery in *Caenorhabditis elegans*. These findings highlight the potential of HHP-treated deer meat sauce as a functional food ingredient that promotes health while contributing to the sustainable utilization of wild game resources.

The substantial increase in anserine and carnosine levels is particularly significant, as these imidazole dipeptides are associated with antioxidant, anti-fatigue, and antihypertensive effects^[13,14]. The identification of new antihypertensive dipeptides, such as alanyl-tryptophan and isoleucyl-tyrosine, further emphasizes the potential health benefits of the HHP-treated sauce^[54]. Enhanced ACE inhibitory activity supports the potential dietary use of this sauce in managing hypertension.

In conclusion, applying HHP treatment to deer meat offers a novel and effective approach to enhancing the functional properties of wild game meat, transforming underutilized resources into high-value functional foods. The HHP-treated deer meat sauce shows promise in promoting health, particularly in managing hypertension and reducing fatigue. Further studies, including clinical trials, sensory evaluations, and explorations of various meat cuts, are necessary to fully validate and optimize the benefits of this innovative food product.

Limitations

This study examined the biological effects of HHP-treated venison sauce on *C. elegans* at a single concentration. Because a dose-response analysis was not performed, the ability to determine the optimal range of effects is limited.

In addition, several potential mechanisms are discussed, including activation of the stress-responsive transcription factor DAF-16/FOXO, uptake via the PEPT-1 transporter, and modulation of oxidative stress (ROS), but these pathways were not directly evaluated in the current experiments. Mechanistic insights were inferred based on the known biological activities of the dipeptides (e.g., carnosine and anserine) identified in the literature and previously

reported sources. To confirm the involvement of these specific molecular pathways, future studies should include analysis of gene expression, mutant strains (e.g., daf-16, pept-1), and quantitative assays for reactive oxygen species (ROS). Recognition of these limitations will improve the scientific balance of the conclusions and highlight important directions for further investigation.

Another limitation is that the study did not evaluate the bioavailability or biological activity of these antihypertensive peptides in human cells or *in vivo*. While these results are useful as a preliminary indicator, they do not provide a complete picture of how the peptides are absorbed, metabolized, and utilized in the human body. Further cellular, animal, and clinical studies are needed to determine the bioavailability of these compounds, particularly whether they can reach their molecular targets intact after digestion.

Ethical statements

The deer meat used in this study was purchased from the Nagano City Gibier Processing Center, which legally obtains deer and holds both the 'National Gibier Certification' and the 'Shinshu Deer Meat Processing Facility Certification.' Because this study only involved the nematode *Caenorhabditis elegans*, no applicable animal ethics approval was required.

Author contributions

The authors confirm contribution to the paper as follows: study conception and design, draft manuscript preparation: Kogiso K; data collection, interpreting of the results: Furuta K. Both authors discussed the results and commented on the manuscript, and approved the final version of the manuscript.

Data availability

The data that support the findings of this study are available on request from the corresponding author.

Acknowledgments

This work was supported by Grants-in-Aid for Scientific Research (KAKENHI) (Grant No. 20K02345) and Nagano City's Funded research. Some strains were provided by the *Caenorhabditis* Genetics Center (CGC), which is funded by the NIH Office of Research Infrastructure Programs (Grant No. P40 OD010440). We would like to take this opportunity to thank you for your support. I am also grateful to Mr. Yamazaki of the Nagano Prefectural Industrial Technology Center.

Conflict of interest

The authors declare that they have no conflict of interest. The funding source had no role in study design, collection, analysis and interpretation of data, the writing of the report, and the decision to submit the article for publication.

Dates

Received 27 February 2025; Revised 14 April 2025; Accepted 30 May 2025; Published online 30 June 2025

References

1. Ministry of Agriculture, Forestry and Fisheries. 2024. Summary of the annual report on food, agriculture and rural areas in Japan 2023. www.maff.go.jp/j/wpaper/w_maff/r5/attach/pdf/index-4.pdf
2. Kogiso K, Kaneko S. 2015. The aroma characteristics and food processing of venison from Nagano. *Journal of Nagano Prefectural College* 69:13–19
3. Yamazaki S, Kogiso K, Ogasawara H, Takemura T. 2016. *Analysis enabling visualization of texture: A case study of research and support using texture testers* [in Japanese]. Preprints of the Presentation of Results, Nagano Prefectural Industrial Technology Center/Nagano Prefectural College of Technology. pp. 21–22
4. Chakanya C, Arnaud E, Muchenje V, Hoffman LC. 2020. Fermented meat sausages from game and venison: what are the opportunities and limitations? *Journal of the Science of Food and Agriculture* 100:5023–31
5. Takeda S, Kaneko S, Sogawa K, Ahmed AM, Enomoto H, et al. 2020. Isolation, evaluation, and identification of angiotensin I-converting enzyme inhibitory peptides from game meat. *Foods* 9:1168
6. Kogiso K. 2023. Assessment of functional components in sika deer and wild boar meats with improvement in processing and flavor and a novel analytical prediction method. *Applied Food Research* 3:100343
7. Lenz CA, Reineke K, Knorr D, Vogel RF. 2015. High pressure thermal inactivation of *Clostridium botulinum* type E endospores – kinetic modeling and mechanistic insights. *Frontiers in Microbiology* 6:652
8. Mora L, Toldrá F. 2023. Advanced enzymatic hydrolysis of food proteins for the production of bioactive peptides. *Current Opinion in Food Science* 49:100973
9. Kurosaki K, Wang F, Yamano H, Yoshida S, Koizumi S, et al. 2020. The current situation and problems of abattoirs for wild deer. *Nihon Chikusan Gakkaiho (Journal of the Japanese Society of Animal Science)* 91:233–39
10. Ministry of Agriculture, Forestry and Fisheries. n. d. Traditional Japanese Foods: Soy sauce, miso, and other fermented seasonings. www.maff.go.jp/e/policies/market/dento_syoku/bunrui/syouyu-miso.html
11. Kim IS, Yang WS, Kim CH. 2021. Beneficial effects of soybean-derived bioactive peptides. *International Journal of Molecular Sciences* 22:8570
12. Chatterjee C, Gleddie S, Xiao CW. 2018. Soybean bioactive peptides and their functional properties. *Nutrients* 10:1211
13. Masuoka N, Lei C, Li H, Hisatsune T. 2021. Influence of imidazole-dipeptides on cognitive status and preservation in Elders: A narrative review. *Nutrients* 13:397
14. Maemura H, Goto K, Yoshioka T, Sato M, Takahata Y, et al. 2006. Effects of carnosine and anserine supplementation on relatively high-intensity endurance performance. *International Journal of Sport and Health Science* 4:86–94
15. Tanida M, Shen J, Kubomura D, Nagai K. 2010. Effects of anserine on the renal sympathetic nerve activity and blood pressure in urethane-anesthetized rats. *Physiological Research* 59:177–85
16. Kogiso K, Sakata S, Tomizawa A, Suzuka H, Saito A, et al. 2023. Taste and safety of processed Nagano venison in very short-term meat sauce preparation. *Journal of Japan Deer Studies* 14:60–66
17. Leung MCK, Williams PL, Benedetto A, Au C, Helmcke KJ, et al. 2008. *Caenorhabditis elegans*: an emerging model in biomedical and environmental toxicology. *Toxicological Sciences* 106:5–28
18. Ha NM, Tran SH, Shim YH, Kang K. 2022. *Caenorhabditis elegans* as a powerful tool in natural product bioactivity research. *Applied Biological Chemistry* 65:18
19. Cho J, Park Y. 2024. Development of aging research in *Caenorhabditis elegans*: From molecular insights to therapeutic application for healthy aging. *Current Research in Food Science* 9:100809
20. Kouriki-Nagatomo H, Kondo T, Nagahama K, Fukui K, Kurogi K, et al. 2019. Effect of retort processing and storage on imidazole dipeptide content of chicken meat. *Nippon Shokuhin Kagaku Kogaku Kaishi (Journal of the Japanese Society for Food Science and Technology)* 66:210–14
21. Yamazaki S, Kaneko S, Takahashi Y, Yoshikawa S. 2016. Survey and analysis on components and physical properties of deer meats captured in Nagano Prefecture. *Research Reports of Nagano Prefecture General Industrial Technology Center Food Technology Department* 11:173–77
22. Alelyunas Y, Roman GT, Johnson JS, Doneanu C, Wrona M. 2015. High throughput analysis at microscale: Performance of ionKey/MS with Xevo G2-XS QToF under rapid gradient conditions. *Journal of Applied Bioanalysis* 1:128–35

23. Matsuda H, Nagaoka T, Morita H, Osajima K, Osajima Y. 1992. Angiotensin I converting enzyme inhibitory peptides generated from sardine muscle by protease for food industry. *Journal of the Japan Food Industry Society* 39:678–83 (in Japanese)
24. Matsui T, Li CH, Osajima Y. 1999. Preparation and characterization of novel bioactive peptides responsible for angiotensin I-converting enzyme inhibition from wheat germ. *Journal of Peptide Science* 5:289–97
25. Ohta T, Iwashita A, Sasaki S, Kawamura Y. 1997. Antihypertensive action of the orally administered protease hydrolysates of chum salmon head and their angiotensin I-converting enzyme inhibitory peptides. *Food Science and Technology International, Tokyo* 3:339–43
26. Matsui T, Yukiyoishi A, Doi S, Sugimoto H, Yamada H, et al. 2002. Gastrointestinal enzyme production of bioactive peptides from royal jelly protein and their antihypertensive ability in SHR. *The Journal of Nutritional Biochemistry* 13:80–86
27. Li CH, Matsui T, Matsumoto K, Yamasaki R, Kawasaki T. 2002. Latent production of angiotensin I-converting enzyme inhibitors from buckwheat protein. *Journal of Peptide Science* 8:267–74
28. Kogiso K, Furuta K, Okazaki M. 2018. Identification of the angiotensin-converting enzyme inhibitory activity peptide of the germinated brown rice sake-lees by LC-MS analysis using ACD/MS Workbook Suite. *Journal of Nagano Prefectural College* 72:15–22
29. Lam LH, Shimamura T, Sakaguchi K, Noguchi K, Ishiyama M, et al. 2007. Assay of angiotensin I-converting enzyme-inhibiting activity based on the detection of 3-hydroxybutyric acid. *Analytical Biochemistry* 364:104–11
30. Lam LH, Shimamura T, Manabe S, Ishiyama M, Ukeda H. 2008. Assay of angiotensin I-converting enzyme-inhibiting activity based on the detection of 3-hydroxybutyrate with water-soluble tetrazolium salt. *Analytical Sciences* 24:1057–60
31. Brenner S. 1974. The genetics of *Caenorhabditis elegans*. *Genetics* 77:71–94
32. Grollemund PM, Poupet C, Comte É, Bonnet M, Veisseire P, et al. 2024. A clustering-based survival comparison procedure designed to study the *Caenorhabditis elegans* model. *Scientific Reports* 14:28257
33. Mitchell DH, Stiles JW, Santelli J, Sanadi DR. 1979. Synchronous growth and aging of *Caenorhabditis elegans* in the presence of fluorodeoxyuridine. *Journal of Gerontology* 34:28–36
34. Huang C, Xiong C, Kornfeld K. 2004. Measurements of age-related changes of physiological processes that predict lifespan of *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences of the United States of America* 101:8084–89
35. Ramot D, Johnson BE, Berry TL Jr, Carnell L, Goodman MB. 2008. The Parallel Worm Tracker: a platform for measuring average speed and drug-induced paralysis in nematodes. *PLoS One* 3:e2208
36. Kanda Y. 2013. EZR: Easy R on R commander. *Bone Marrow Transplantation* 48:452–58
37. Nakahara T, Sano A, Yamaguchi H, Sugimoto K, Chikata H, et al. 2010. Antihypertensive effect of peptide-enriched soy sauce-like seasoning and identification of its angiotensin I-converting enzyme inhibitory substances. *Journal of Agricultural and Food Chemistry* 58:821–27
38. Li FJ, Yin LJ, Cheng YQ, Saito M, Yamaki K, et al. 2010. Angiotensin I-Converting Enzyme Inhibitory Activities of Extracts from Commercial Chinese Style Fermented Soybean Paste. *Food Technology* 44(2):167–72
39. Okamoto A, Hanagata H, Kawamura Y, Yanagida F. 1995. Anti-hypertensive substances in fermented soybean, natto. *Plant Foods for Human Nutrition* 47(1):39–47
40. Kuba M, Tanaka K, Tawata S, Takeda Y, Yasuda M. 2003. Angiotensin I-converting enzyme inhibitory peptides isolated from tofuyo fermented soybean food. *Bioscience, Biotechnology, and Biochemistry* 67(6):1278–83
41. Kubota D. 2012. Comparative Study on Antihypertensive Activity of Meat Hydrolysates Sourced from Bovine, Porcine and Poultry. *Bulletin of the Faculty of Agriculture, University of Miyazaki* 58:43–50
42. Lunow D, Kaiser S, Brückner S, Gotsch A, Henle T. 2013. Selective release of ACE-inhibiting tryptophan-containing dipeptides from food proteins by enzymatic hydrolysis. *European Food Research and Technology* 237:27–37
43. Song CC, Qiao BW, Zhang Q, Wang CX, Fu YH, et al. 2021. Study on the domain selective inhibition of angiotensin-converting enzyme (ACE) by food-derived tyrosine-containing dipeptides. *Journal of Food Biochemistry* 45:e13779
44. Suetsuna K, Maekawa K, Chen JR. 2004. Antihypertensive effects of *Undaria pinnatifida* (wakame) peptide on blood pressure in spontaneously hypertensive rats. *The Journal of Nutritional Biochemistry* 15:267–72
45. Ferguson AA, Roy S, Kormanik KN, Kim Y, Dumas KJ, et al. 2013. TATN-1 mutations reveal a novel role for tyrosine as a metabolic signal that influences developmental decisions and longevity in *Caenorhabditis elegans*. *PLoS Genetics* 9:e1004020
46. Manzanarez P, Martínez R, Garrigues S, Genovés S, Ramón D, et al. 2018. Tryptophan-containing dual neuroprotective peptides: prolyl endopeptidase inhibition and *Caenorhabditis elegans* protection from β -amyloid peptide toxicity. *International Journal of Molecular Sciences* 19:1491
47. Boldyrev AA, Aldini G, Derave W. 2013. Physiology and pathophysiology of carnosine. *Physiological Reviews* 93:1803–45
48. Zhou KI, Pincus Z, Slack FJ. 2011. Longevity and stress in *C. elegans*: regulation of DAF-16 by peptide binding and post-translational modifications. *Cell Metabolism* 14:161–72
49. Geissler S, Zwarg M, Knütter I, Markwardt F, Brandsch M. 2010. The bioactive dipeptide anserine is transported by human proton-coupled peptide transporters. *The FEBS Journal* 277:790–95
50. Meissner B, Boll M, Daniel H, Baumeister R. 2004. Deletion of the intestinal peptide transporter affects insulin and TOR signaling in *Caenorhabditis elegans*. *Journal of Biological Chemistry* 279:36739–45
51. Kenyon C, Chang J, Gensch E, Rudner A, Tabtiang R. 1993. A *C. elegans* mutant that lives twice as long as wild type. *Nature* 366:461–64
52. Kumar S, Dietrich N, Kornfeld K. 2016. Angiotensin converting enzyme (ACE) inhibitor extends *Caenorhabditis elegans* life span. *PLoS Genetics* 12(2):e1005866
53. Egan BM, Pohl F, Anderson X, Williams SC, Adodo IG, et al. 2024. The ACE inhibitor captopril inhibits ACN-1 to control dauer formation and aging. *Development* 151(3):dev202146
54. Khan MTH, Dedachi K, Matsui T, Kurita N, Borgatti M, et al. 2012. Dipeptide inhibitors of thermolysin and angiotensin I-converting enzyme. *Current Topics in Medicinal Chemistry* 12:1748–62



Copyright: © 2025 by the author(s). Published by Maximum Academic Press on behalf of Nanjing Agricultural University. This article is an open access article distributed under Creative Commons Attribution License (CC BY 4.0), visit <https://creativecommons.org/licenses/by/4.0/>.