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Characterization of the anthocyanin-related *flavanone 3-hydroxylase* (*F3H*) genes and functional analysis of *VcF3H2* in blueberry

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Abstract

Flavanone 3-hydroxylase (F3H) is a key enzyme involved in plant flavonoid/anthocyanin biosynthesis pathways. In this study, we identified eight candidate anthocyanin-related *F3H* genes from blueberry (*Vaccinium corymbosum*), and characterized their sequences and expression patterns. These anthocyanin-related VcF3Hs are conserved in sequences and their encoding genes express highly in blueberry fruits at late ripening stages. By using apple and blueberry fruits transient overexpression analysis, the function of *VcF3H2* was studied. Its overexpression significantly improved the anthocyanin accumulation and upregulated the expression levels of anthocyanin biosynthesis-related structural genes in fruit peels of both apple and blueberry. Numerous binding sites for MYB and bHLH transcription factors (TFs) were identified in promoters of *VcF3Hs*, especially *VcF3H1~4*. Consistently, our yeast one-hybrid and dual-luciferase assay results showed that both the anthocyanin-related VcMYB-1 and VcAN1 could bind to and activate the promoter of *VcF3H2*, indicating that its roles in anthocyanins biosynthesis were regulated by anthocyanin-related TFs. Our study characterized the anthocyanin-related *F3Hs* in blueberry and demonstrated the role of the 'VcMYB-1/VcAN1-VcF3H2' module in regulating blueberry anthocyanin biosynthesis.

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Introduction

Flavanone 3-hydroxylase (F3H), catalyzing the conversion of naringenin into dihydroflavonols, is a key enzyme involved in plant flavonoid/anthocyanin biosynthesis pathways[1,2]. The expression levels of genes encoding F3Hs have been identified to be associated with flavonoid/anthocyanin accumulation in many plants. For example, in anthocyanin-rich 'Zhongshen 1' mulberry fruits, the expression level of *F3H* is positively correlated with the anthocyanin content[3]; the expression of *CitF3H* is positively correlated with the anthocyanin accumulation in juice sacs of blood orange 'Moro' during fruit ripening[4]; the *anthocyanin reduce* (*are*) tomato mutant, caused by the mutation in the *F3H* gene, has much higher accumulation of naringenin but significantly lower levels of downstream products including anthocyanins[5]; transgenic rice overexpressing *OsF3H* showed upregulated expression of downstream flavonoid biosynthesis related structural genes (*OsDFR* and *OsFLS*), and promoted accumulation of kaempferol, quercetin, delphinidin, and cyanidin[6]. The naringenin content in transgenic Arabidopsis seedlings overexpressing the *Pohlia nutans F3H* showed significantly lower accumulation of naringenin but much higher levels of downstream flavonoids, and enhanced salt resistance and antioxidant ability[7].

Evidence revealed that the expression of *F3H* genes could be greatly influenced by many environmental factors[8]. Blue light can promote the anthocyanin biosynthesis by upregulating the expression of *CitF3H*[4]. In grapes, glucose, fructose, and sucrose treatments can all induce *F3H* transcription and promote the biosynthesis and accumulation of anthocyanins in a hexokinase-dependent pathway[9]. Similarly, in Arabidopsis, sucrose treatment can activate the expression of *AtF3H*, thereby promoting the anthocyanin accumulation in seedlings[10,11]. Moreover, it was found that additional JA and ABA treatments can further enhance the sucrose-induced *AtF3H* expression and anthocyanin accumulation[11]. The expression of *Dendrobium officinale F3H* was reported to be salt- and cold-inducible, and its upregulation resulted in increased anthocyanin and flavonol accumulation and conferred plants with a tolerance to these stresses[12]. Tobacco plants overexpressing an *F3H* from *Lycium chinense* showed upregulated expression of *DFR* and *ANS* genes, improved flavon-3-ol accumulation, and enhanced antioxidant ability[13].

Recent research demonstrated the transcriptional regulation of transcription factors (TFs) on *F3H* genes. The mulberry ERF5 can bind to the 'GCCGAC' sequences on the promoters of anthocyanin-related *MYBA* and *F3H* genes, activating their expression and promoting anthocyanin biosynthesis in fruits[14]. The rice OsC1^{PLSr} functions in regulating anthocyanins biosynthesis and accumulation by activating the expression of anthocyanin biosynthesis structural genes, including *F3H*[15]. *Erigeron breviscapus* EbMYBP1 can directly bind to the promoter of *EbF3H*, activate the gene's transcription, and promote flavonoid biosynthesis and accumulation in leaf[16].

Blueberry, a perennial woody plant, is known for its anthocyanin-rich fruits. Its fruits have very high health-care values, with anthocyanins as the main contributors[17,18]. Currently, there are some reports on the functions of blueberry *F3H* genes in anthocyanin biosynthesis. For example, Zhang et al.[19] cloned a *F3H* gene (named *VcF3H*) from blueberry leaf, and verified its anthocyanin accumulation promoting effect through heterologous expression in Arabidopsis. Lin et al.[20] identified several anthocyanin biosynthesis structural genes, including a *F3H*, from blueberry transcriptome data, and found that the expression levels of these genes are positively correlated with anthocyanin content. Xu et al.[21] studied the influences of dielectric barrier discharge cold plasma (CP) on anthocyanin accumulation, and found that CP treatment could upregulate the expression of *F3H* and some other structural genes in post-harvested blueberry fruits. To date, however, there is no report focusing on the systematic identification, characterization, and functional analysis of

anthocyanin-related *F3H* genes in blueberry. In this study, we first identified and characterized all the anthocyanin-related *F3Hs* in the blueberry genome, and investigated their expression patterns under different abiotic stress treatments in fruits at five different ripening stages using transcriptome data and quantitative real-time PCR (qRT-PCR) analysis. Furthermore, the function of *VcF3H2*, an anthocyanin-related *VcF3H* showing high expression in fruits and the most significant differential expression changes during fruit ripening, was studied using transient overexpression assays in apple and blueberry fruits. It is worth noting that, many binding sites for TFs, such as MYB and bHLH, were found in the promoters of *VcF3H2* and some other anthocyanin-related *VcF3Hs*. To reveal the possible transcriptional regulation of TFs on *VcF3Hs*, the binding and activation abilities of two anthocyanin biosynthesis regulatory TFs, VcMYB-1[22] and VcAN1[23,24], on the *VcF3H2* promoter were verified using dual-luciferase assays (LUC) and yeast one-hybrid (Y1H). Our study will be helpful for understanding the roles of *F3H* in blueberry anthocyanin biosynthesis and can provide a basis for clarifying the transcriptional regulation of TFs in anthocyanin biosynthesis.

Materials and methods

Plant materials

The blueberry plants used in this study were healthy and unique two-year-old 'Legacy' and 'FL03' seedlings cultivated in our lab. The green, pink, red, purple, and blue blueberry fruits used for gene cloning and expression analysis were harvested from two-year-old 'FL03' plants. Bagged 'Gala' apples used for transient overexpression analysis were harvested from the apple cultivation base at the Institute of Horticulture, College of Horticulture, Shanxi Agricultural University (Shanxi, China). For tobacco leaf transient overexpression, 6-week-old *Nicotiana benthamiana* plants were used.

Identification and characterization of anthocyanin-related *F3H* genes in blueberry

The genome data files of southern highbush blueberry (*Vaccinium corymbosum*) were downloaded from GDV (GENOMEDATABASE FOR VACCINIUM, www.vaccinium.org). The protein sequence of anthocyanin-related AtF3H (AT3G51240.1)[25] was downloaded from TAIR (www.arabidopsis.org), and sequences of other anthocyanin-related F3Hs, including VvF3H (NM001281105.1)[9], OsF3H (Os04g0662600)[26], GmF3H (ACM62745.1)[27], LcF3H (AID50182.1)[28], PeF3H (AIS35916.1)[29], TfF3H (AGL98422.1)[30], were downloaded from NCBI (www.ncbi.nlm.nih.gov). By using AtF3H, VvF3H and OsF3H as query sequences, local blastp was used to identify anthocyanin-related F3Hs in blueberry (E-value ≤ 1×10^{-5}, identity ≥ 60%). Then, CDD (www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi) was used to verify the presence of the PLN02515 superfamily domain in identified VcF3Hs.

MEGAX was used for the alignment and phylogenetic analysis of VcF3Hs and flavonoid/anthocyanin-related F3Hs reported from other plants. A phylogenetic tree for these F3Hs was constructed using Neighbor-Joining method (Bootstrap = 1,000 and other parameters were set as default values), ITOL (https://itol.embl.de/) was used for figure drawing. For the prediction of conserved motifs in anthocyanin-related VcF3Hs, MEME (http://meme-suite.org/tools/meme) was used. TBtools software was used to visualize the motif distribution, and to predict the conserved domains in VcF3Hs and gene structures of their corresponding genes[31]. The Advanced Circos module of TBtools was used for the synteny analysis of anthocyanin-related *VcF3Hs*, and the Simple Ka/Ks Calculator (NG) program was used to calculate the duplication time among duplicated *VcF3H* gene pairs[32].

The Protein Parameter Calc (ProtParam-based) module of TBtools and Plant-mPLoc (www.csbio.sjtu.edu.cn/bioinf/plant-multi)[33] was used to predict the physiochemical proprieties and subcellular localization of VcF3Hs, respectively.

Gene expression analysis

In our previous studies, we sequenced the transcriptomes of ABA (treated with 100 mM ABA), MeJA (100 mM MeJA), salt (200 mM NaCl and Na_2SO_4), alkali (200 mM $NaHCO_3$ and Na_2CO_3), 4 °C low temperature (LT), and 40 °C high temperature (HT) treated blueberry leaves and 'FL03' fruits at five ripening stages. In this study, the FPKM (Fragments Per Kilobases per Million reads) values of anthocyanin-related *VcF3Hs* were extracted from these transcriptome data for heatmap drawing using TBtools. To further verify their expression patterns in these samples, quantitative real-time PCR (qRT-PCR) analysis of three selected *VcF3Hs* (*VcF3H2*, *VcF3H6* and *VcF3H8*) was also performed with three replications. Primers used for the qRT-PCR verification were designed using primer3 (http://primer3.ut.ee/) and listed in Supplementary Table S1. Amplifications were performed on a Bio-Rad CFX96™ real-time fluorescent quantitative PCR machine with a reaction procedure as follows: pre-denaturation at 95 °C for 30 s; denaturation at 95 °C for 15 s, annealing at 60 °C for 30 s, and extension at 72 °C for 30 s, 45 cycles. By using the *GAPDH* (AY123769)[34] as an internal reference gene, the relative expression of selected *VcF3Hs* in different blueberry samples was calculated using the $2^{-\Delta\Delta CT}$ method.

Prediction of VcF3Hs interacting proteins

Based on the Arabidopsis protein database, STRING v12.0 (https://string-db.org/) was used to predict proteins interacting with VcF3Hs (minimum required interaction score > 0.700)[35]. Then, the Cytoscape software was used to draw the protein-protein interacting network figure of VcF3Hs[36].

Promoter analysis

The 2,000 bp sequences upstream the start codon (ATG) of anthocyanin-related *VcF3Hs* were extracted from blueberry genome data using TBtools and considered as promoter sequences. By using PlantCARE (http://bioinformatics.psb.ugent.be/webtools/plantcare/html/), and PlantRegMap (https://plantregmap.gao-lab.org/binding_site_prediction.php), the *cis*-acting elements and transcription factor binding sites in promoters of anthocyanin-related *VcF3Hs* were predicted (*p*-value ≤ 1×10^{-5}), respectively.

Gene cloning and overexpression vector construction

The *VcF3H2* coding sequence (CDS) was amplified through reverse transcription PCR by using 'FL03' blueberry fruit cDNA as template. After sequencing verification, the gene was subcloned and introduced into the pBI121 vector using the Ready-to-use Seamless Cloning Kit produced by Songon Biotech (Shanghai, China). Then, the obtained p35S::VcF3H2 recombinant vectors were transformed into Agrobacterium GV3101. Primers used for gene cloning and vector construction are listed in Supplementary Table S1.

Transient expression analysis of *VcF3H2* in apple and blueberry fruits

According to the method described by Lv et al.[37], Agrobacteria GV3101 carrying p35S::VcF3H2 and empty pBI121 vectors (EV) were cultured to OD_{600} ≈ 1.5 in 50 mL of Luria -Bertani (LB) medium. Then,the Agrobacteria solution was centrifuged at 5,000 rpm for 4 min. After discarding the supernatant, the Agrobacteria pellet was re-suspended using MES solution, adjusted to OD_{600} = 0.8, shake-cultured at 200 rpm at 28 °C for 30 min, and kept still at 28 °C in the dark for 1 h to obtain the Agrobacteria infiltration solution used for apple and blueberry fruit injection.

For apple fruit transient overexpression analysis, approximately 30 μL Agrobacteria infiltration solution was injected into each site around the equatorial regions of apple fruits, and each fruit was injected at two opposite sites. Infiltrated fruits were kept in the dark at 24 °C for 2 d, followed by light conditions with a light density of 1,500 lx and photoperiod of 16 h light/ 8h dark. For blueberry fruit transient overexpression, green fruits on two-year-old 'Legacy' blueberry plants were used, and each fruit was injected with 10~15 μL Agrobacteria infiltration solution at one site. After agrobacteria infiltration, blueberry plants were then kept in the dark at 24 °C for 2 d, and removed to normal light condition. Five days later, CR8 colorimeter (3nh, Guangzhou, China) was used to measure the color parameters of apple and blueberry fruits. Then, the anthocyanins contents in apple and blueberry peels around the injected regions were determined according to the method of Zhang et al.[38].

Apple and blueberry fruit peel total RNA was isolated using Trizol (Invitrogen, Waltham, CA, USA), and reversed transcribed into cDNA using PrimeScript RT Master Mix (Perfect Real Time) kit (Takara, Dalian, China). These cDNA samples were used as templates for qRT-PCR analysis of anthocyanin biosynthesis structural genes (*MdCHI*, *MdCHS*, *MdF3H*, *MdDFR*, *MdUFGT*, and *MdANS* in apple[39], and *VcCHS*, *VcDFR*, *VcF3H*, *VcUFGT*, and *VcANS* in blueberry[40], Supplementary Table S1) in fruit peels.

Yeast one-hybrid (Y1H)

By using 'FL03' gDNA as a template, a 1,960 bp long *VcF3H2* promoter sequence was amplified. Then, the promoter sequences were introduced into the pAbAi vector to obtain the recombinant vector *pVcF3H2*$_{1960}$, and transformed into the yeast strain Y1HGold. Then, pGADT7-VcAN1 and pGADT7-VcMYB-1 were individually transformed into the yeast strain carrying *pVcF3H2*$_{1960}$ to verify the binding activity of these two anthocyanin-related TFs on the VcF3H promoter. The pGADT7-VcAN1 and pGADT7-VcMYB-1 vectors were provided by our laboratory[23,24].

Dual-luciferase assay (LUC)

The 1,960 bp *VcF3H2* promoter sequences were introduced into pNC-Green-LUC vector to obtain the *LUC-pVcF3H2*$_{1960}$ recombinant vector. The stop codon removed *VcMYB-1/VcAN1* CDS sequences were introduced into the pNC-GreenII-SK vector to obtain the SK-MYB-1 and SK-AN1 recombinant vectors. Vectors were transformed into Agrobacteria GV3101 (pSoup-p19), and cultured till OD_{600} reached approximately 1. Agrobacteria carrying SK-MYB-1/SK-AN1 were equal volume mixed with the *LUC-pVcF3H2*$_{1960}$ Agrobacteria suspension, kept still at room temperature for 30 min, and then infiltrated into the abaxial side of *N. benthamiana* leaves. Simultaneously, pNC-Green-LUC + pNC-Green-SK, SK-VcMYB-1/VcAN1 + pNC-Green-LUC, and pNC-Green-SK + LUC-*pVcF3H2*$_{1960}$ Agrobacteria suspension mixture was also infiltrated into the same tobacco leaf at different parts. For each experiment, at least 20 tobacco leaves were used as replications. After infiltration, tobacco plants were cultured in dark at 25 °C for 2 d, and an additional 3 d under normal light conditions[41]. Then, infiltrated tobacco leaves were harvested, smeared with 1.5 mM luciferin substrate solution, kept in the dark for 5 min, and observed under a Tanon 5200 system (Tanon Science & Technology Co., Ltd., Shanghai, China).

Results

Characterization, phylogenetic, and synteny analysis results of anthocyanin-related VcF3Hs

In total, we identified eight candidate anthocyanin-related VcF3Hs from blueberry, which were named VcF3H1~8 according to their amino acid (AA) number and scaffold localization information of their corresponding encoding genes. Of them, VcF3H1~4 all have 359 AA, VcF3H5 and VcF3H6 have 366 AA, and VcF3H7 and VcF3H8 have 373 AA (Fig. 1a, Supplementary Table S2). Protein similarity analysis revealed that these eight VcF3Hs shared more than 90%

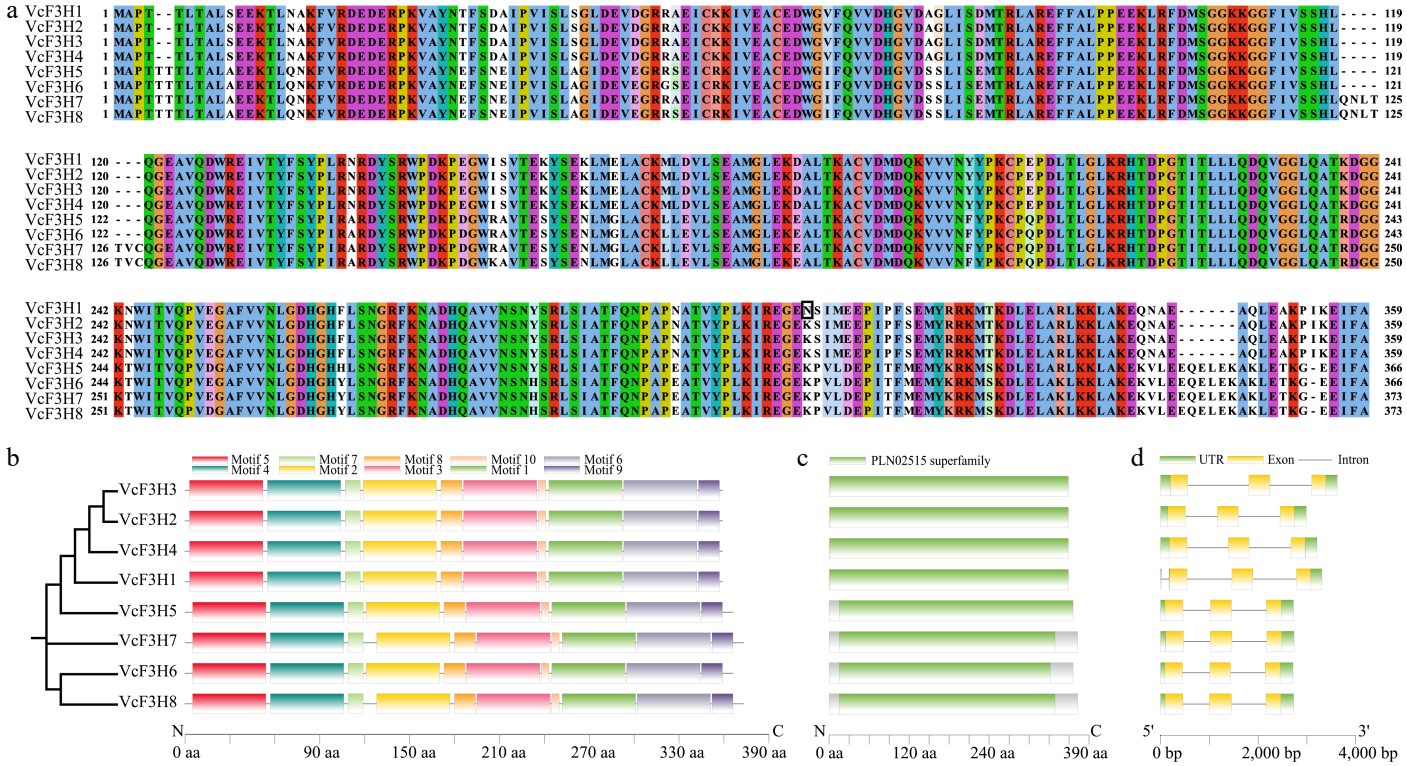

Fig. 1 (a) Sequence alignment, (b) conserved motifs, (c) domain, and (d) gene structure analysis results of the eight anthocyanin-related VcF3Hs identified in blueberry.

similarities. The protein sequences of VcF3H2, VcF3H3, and VcF3H4 are the same, and there is only one amino acid difference between VcF3H2/3/4 and VcF3H1. The protein similarities among VcF3H5~8 are all higher than 96%. In consistence with their high sequence similarities, the motifs identified in anthocyanin-related VcF3Hs are the same in both numbers and orders (Fig. 1b). Conserved domain analysis results showed that they all contained a PLN02515 superfamily domain (Fig. 1c). Gene structure analysis revealed that all the eight *VcF3Hs* have two introns in their corresponding gDNA, and one additional intron was found in the 5'UTR of *VcF3H1* (Fig. 1d). All the VcF3Hs were predicted to be localized in cytoplasm (Supplementary Table S2), which was consistent with the biosynthesis site of anthocyanins.

Phylogenetic analysis of flavonoid/anthocyanin-related F3Hs from blueberry, Arabidopsis, grape, rice, and some other plants revealed that VcF3H1~4 are close to each other, and VcF3H5/6/7/8 shared close relationships (Fig. 2a). The eight anthocyanin-related VcF3Hs shared greater more than 81% similarities with other reported flavonoid/anthocyanin-related F3Hs (Fig. 2b).

Synteny analysis showed that there were 12 pairs of duplicated genes among the eight anthocyanin-related *VcF3Hs*, including six pairs among *VcF3H1~4* and six pairs among *VcF3H5~8* (Fig. 2c, Supplementary Table S3). The Ka/Ks values of all replicated gene pairs ranged from 0 to 0.1475, indicating that the anthocyanin-related *VcF3Hs* experienced strong purification selection pressure during evolution. By calculating the time of their duplication events, it was found that these gene replication events occurred 0 million years ago (Mya) ~0.28 Mya.

Based on the Arabidopsis protein database, the interacting proteins of anthocyanin-related VcF3Hs were predicted using STRING (Fig. 2d). They were all identified as homologous proteins of AtF3H, and their interacting proteins included many key enzymes involved in the flavonoids/anthocyanin biosynthesis pathway, including C4H, CHI, CHS, F3'H, FLS, DFR, UFGT, and so on. It is suggested that these VcF3Hs are involved in blueberry anthocyanin metabolism.

Promoter analysis results

The distribution of *cis*-acting elements of the *VcF3Hs* promoters was analyzed. Besides the common CAAT-box and TATA-box

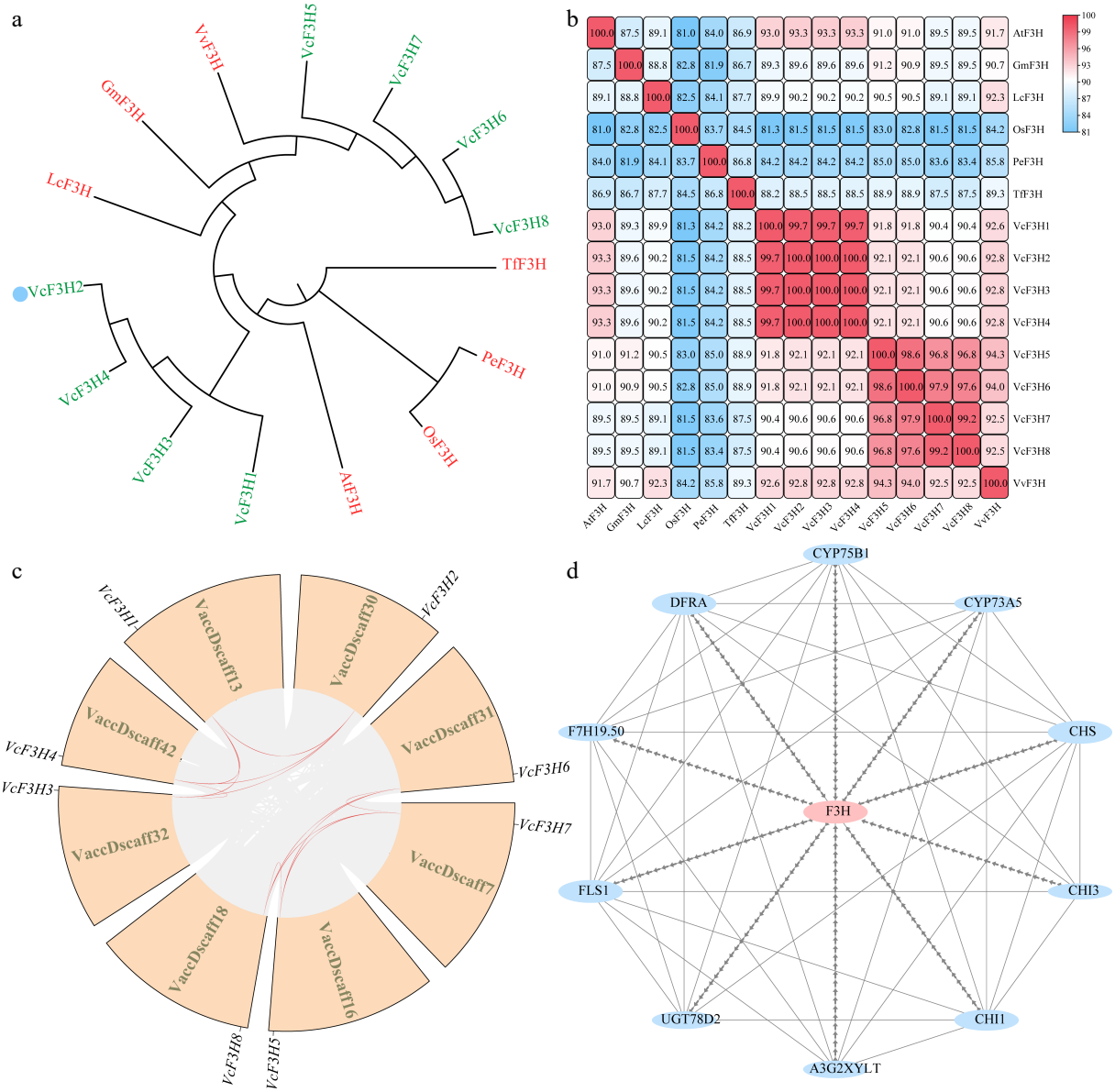

Fig. 2 (a) Phylogenetic, (b) similarity, (c) synteny, and (d) interacting protein analysis results of VcF3Hs.

elements and some functional unknown elements, 39 kinds of elements (in amount of 334) are identified on their promoters. These elements can be further classified into four categories: light responsive, growth, and development-related, phytohormone responsive, and stress responsive (Fig. 3a). All *VcF3Hs* promoters contained light responsive G-box element, ABA-responsive related ABRE element, drought-inducibility related MYC element, defense and stress-related MYB element, high-temperature responsive STRE elements, and endosperm expression related GCN4_motif elements.

We also identified binding sites for TFs from 29 families (Fig. 3b), such as MYB, bHLH, ERF, TCP, and NAC, in promoters of anthocyanin-related *VcF3Hs*. Among them, binding sites for MYB accounted for the largest (149), followed by ERF (83), and bHLH (62). The number of TFBSs on *VcF3H1* promoter was the highest (105), and the number of binding sites on *VcF3H6* promoter was the lowest (59). There are MYB, bHLH, MYB_related, BES1, WRKY, and TCP binding sites on all *VcF3Hs* promoters. It is worth noting that each *VcF3H* promoter contains at least two binding sites for MYB, bHLH, BES1, and TCP. In addition, the MYB and bHLH binding sites amounts on *VcF3H1~4* promoters were much higher than that on *VcF3H5~8* promoters.

Gene expression analysis results

According to our blueberry fruit transcriptome data, the expression patterns of anthocyanin-related *VcF3Hs* in fruits at five ripening stages were first studied (Fig. 4a). Results showed that *VcF3H1~4* all exhibited a 'fall-rise' expression pattern during fruit ripening, and their expression levels all peaked in purple fruits (PF). The expression of *VcF3H7~8* increased as fruit ripened. However, the *VcF3H5~6* expression levels in fruits at different ripening stages followed the order: green fruit (GF) < pink fruit (PiF) < PF < red fruit (RF) < blue fruit (BF). The expression of *VcF3H2*, *VcF3H6,* and *VcF3H8* were further verified by qRT-PCR (Fig. 4b). Results showed that the expression pattern of *VcF3H2* was consistent with our transcriptome results. Its expression level in PiF, RF, PF, and BF was about 5.93-, 33.58-, 38.96-, and 20.00-fold of GF, respectively. Although the qRT-PCR results for *VcF3H6* and *VcF3H8* were not the same as our transcriptome data. Their relative expression levels in PiF, RF, PF, and BF were significantly higher than those in GF.

There are many stress- and phytohormone-responsive elements in promoters of anthocyanin-related *VcF3Hs*, suggesting that their expression may be significantly affected by various stresses and phytohormone treatments. Consistently, our leaf transcriptome data showed that ABA, MeJA, alkali, salt, and LT treatments significantly upregulated the expression of all anthocyanin-related *VcF3Hs* (Fig. 4c). Interestingly, HT treatment significantly upregulated the expression of *VcF3H1~4* but downregulated the expression of *VcF3H5~8*. The expression of *VcF3H2* was significantly upregulated by ABA, MeJA, alkali, salt, HT, and LT (Fig. 4d), accounting for 4.28-, 1.48-, 3.57-, 2.24-, 3.58-, and 5.36-fold of CK, respectively. ABA, alkali, salt, and LT treatments respectively upregulated *VcF3H6* expression by 3.05-, 1.84-, 2.60-, and 2.10-fold compared to CK, while HT and JA

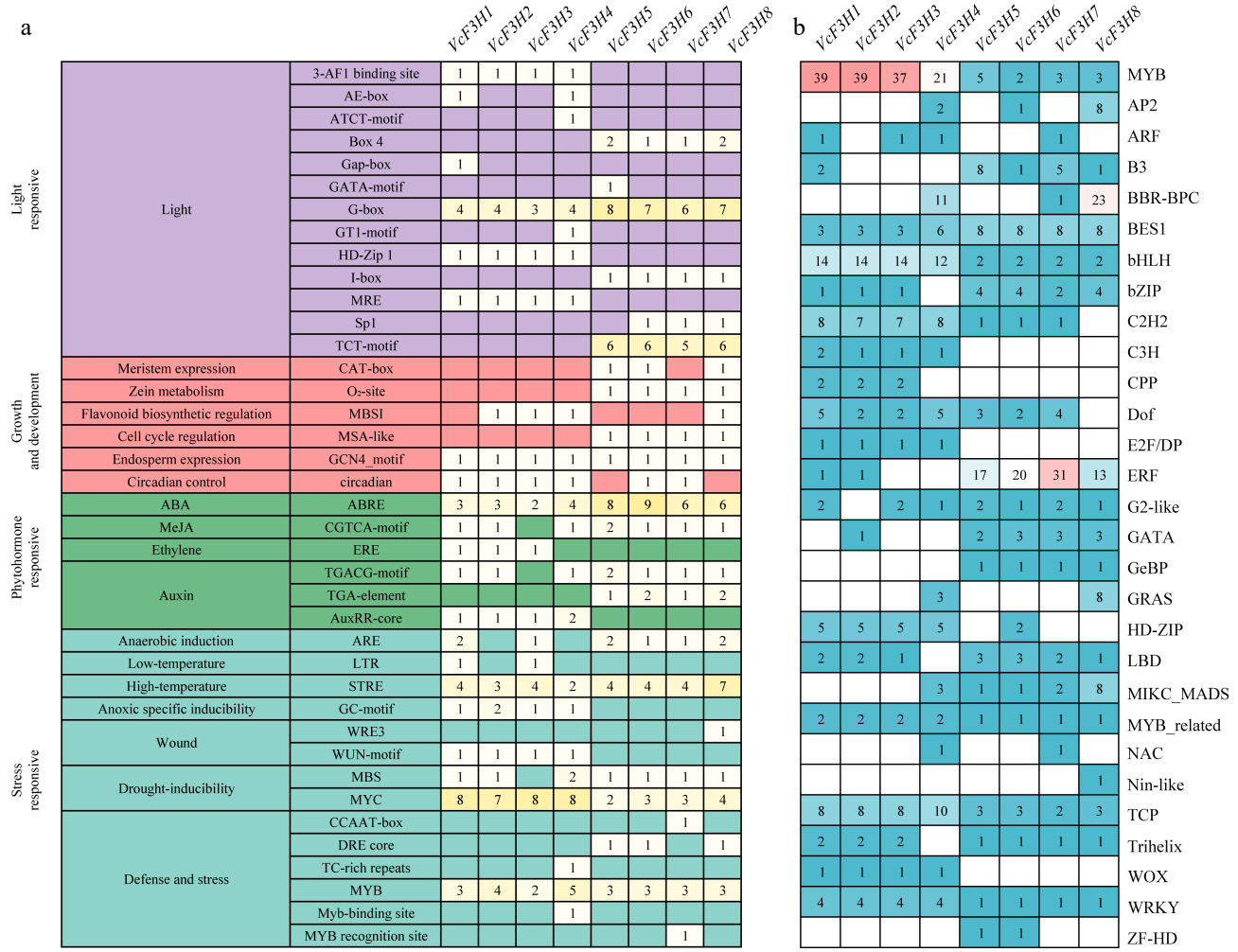

Fig. 3 Predicted (a) *cis*-acting elements, and (b) transcription factor binding sites in promoters of *VcF3H*.

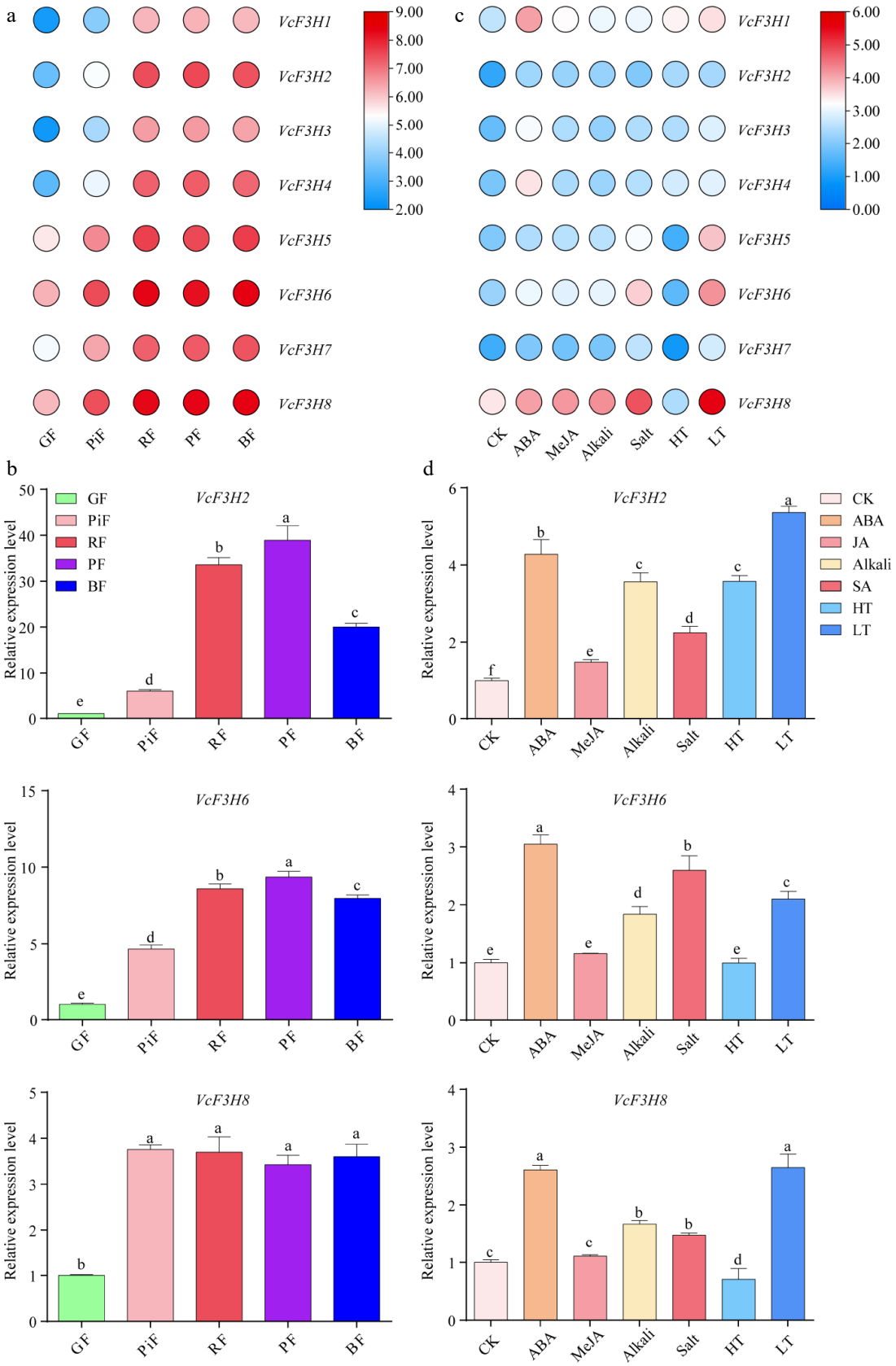

Fig. 4 Gene expression analysis results of *VcF3Hs* in (a), (b) fruits at five different ripening stages, and (c), (d) in leaves treated with different phytohormones and abiotic stresses. (a) and (c) are transcriptome data analysis results. The Log_2 (FPKM + 1) value is used for heatmap drawing. The redder the color, the higher the gene expression, and the bluer the color, the lower the gene expression. (b) and (d) are qRT-PCR analysis results. Different letters above columns represent a significant difference at the $p < 0.05$ level. GF: green fruit; PiF: pink fruit; RF: red fruit; PF: purple fruit; BF: blue fruit; CK: nontreated control leaf; ABA, MeJA, Alkali, Salt, HT, and LT represents leaf treated with 100 mM ABA, 100 mM MeJA, 200 mM NaCl, and Na_2SO_4, 200 mM $NaHCO_3$ and Na_2CO_3, 4 °C low temperature and 40 °C high temperature, respectively.

treatments did not significantly influence its expression. *VcF3H8* expression was upregulated by 2.61-, 1.11-, 1.67-, 1.48-, and 2.65-fold by ABA, MeJA, alkali, salt, and LT treatment, respectively. However, HT downregulated its expression to approximately 71% of CK, and MeJA treatment slightly upregulated its expression.

Transient overexpression analysis of *VcF3H2* in apple and blueberry fruits

VcF3H2 expressed highly in blueberry fruit and exhibited the most significant upregulation during fruit ripening compared to other

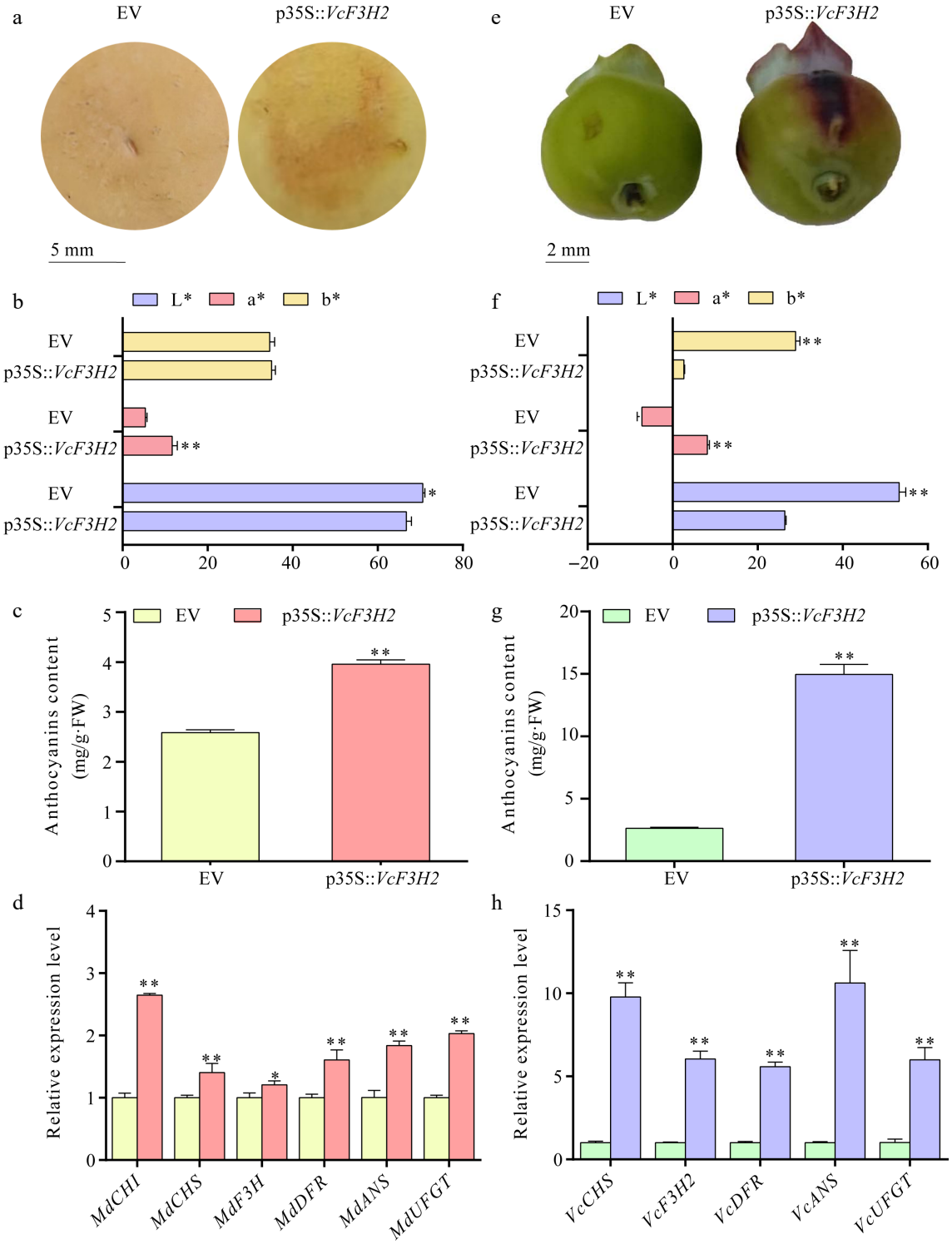

Fig. 5 The influences of *VcF3H2* overexpression in anthocyanins biosynthesis and expression of corresponding structural genes in apple and blueberry fruits. (a) Transient overexpression of *VcF3H2* promoted anthocyanins accumulation in apple peel; (b)–(d) Effects of overexpression of *VcF3H2* on color parameters, anthocyanins content and expression of anthocyanins biosynthesis structural genes in apple peels. (e) Transient overexpression of *VcF3H2* promoted anthocyanins accumulation in blueberry fruit peels; (f)–(h) Effects of overexpression of *VcF3H2* on color parameters, anthocyanin content, and expression of anthocyanin biosynthesis structural genes in blueberry fruit peels. Data were presented as the means ± standard deviations of three biological replicates, with asterisks showing significant differences (*, $p < 0.05$; **, $p < 0.01$). In (b) and (f), L* indicates lightness from 0 (black) to 100 (white); a* indicates red/green (+a* = redder, −a = greener); b* indicates yellow/blue (+b* = yellower, −b* = bluer).

anthocyanin-related *VcF3Hs*. This suggested that it might play a key role in anthocyanin biosynthesis in blueberry fruits during ripening. By using transient overexpression assays in apple and blueberry fruits, its functions were further studied (Fig. 5). Results showed that its overexpression resulted in obvious pigment accumulation in apple peels (Fig. 5a & b). The anthocyanin content in apple peels overexpressing *VcF3H2* was found to be 1.53-fold of EV (Fig. 5c) QRT-PCR analysis showed that its overexpression significantly upregulated the expression levels of *MdCHI*, *MdCHS*, *MdDFR*, *MdANS*, and *MdUFGT* in apple peels (Fig. 5d), accounting for 2.65-, 1.40-, 1.61-, 1.83-, and 2.03-fold of EV, respectively.

The overexpression of *VcF3H2* also led to obvious pigmentation in blueberry fruit peels (Fig. 5e & f). The anthocyanin content in blueberry fruit peels overexpressing *VcF3H2* was about 5.68-fold of EV (Fig. 5g), and the expression level of *VcF3H2* accounted for 6.04-fold of EV (Fig. 5h). Its overexpression significantly upregulated the expressions of *VcCHS*, *VcDFR*, *VcANS*, and *VcUFGT* in blueberry peel, accounting for 9.78-, 5.58-, 5.99-, and 10.62-fold of EV (Fig. 5h), respectively.

The binding and activation effects of VcMYB-1 and VcAN1 on *VcF3H2* promoter

Promoter analysis showed that all anthocyanin-related *VcF3Hs* contained G-box, Myb, and MYC elements, as well as the binding sites for MYB and bHLH TFs in their promoters. These indicated that the expression of *VcF3Hs* might be regulated by anthocyanin-regulatory MYBs and bHLHs. By using Y1H, we investigated the binding ability of VcAN1 and VcMYB-1 to the *VcF3H2* promoter. Results showed that the growth of yeast carrying $pVcF3H2_{1960}$ was inhibited on SD/-Ura media containing 250 mM Aureobasidin A (AbA) (Fig. 6a), indicating that $pVcF3H2_{1960}$ exhibited no self-activation. Yeast co-transformed with $pVcF3H2_{1960}$ and pGADT7 could not grow on SD/-Leu medium containing 250 mM AbA (Fig. 6b), while strains co-transformed with $pVcF3H2_{1960}$ and pGADT7-VcAN1/VcMYB-1 grew well. These results indicate that both VcAN1 and VcMYB-1 can bind to the *VcF3H2* promoter.

To further investigate the binding ability of VcMYB-1 and VcAN1 to the *VcF3H2* promoter, dual-luciferase assays (LUC) were performed (Fig. 6c). Both SK-MYB-1 and SK-AN1 were found to significantly enhanced the fluorescence of LUC-pVcF3H2$_{1960}$ in tobacco leaves (Fig. 6d & e), indicating that both VcMYB-1 and VcAN1 can promote anthocyanins accumulation by directly binding to and activating the promoter of *VcF3H2*.

Discussion

F3H, catalyzing naringenin into dihydroflavonols, intermediates in the biosynthesis of both flavonols and anthocyanidins, is a key

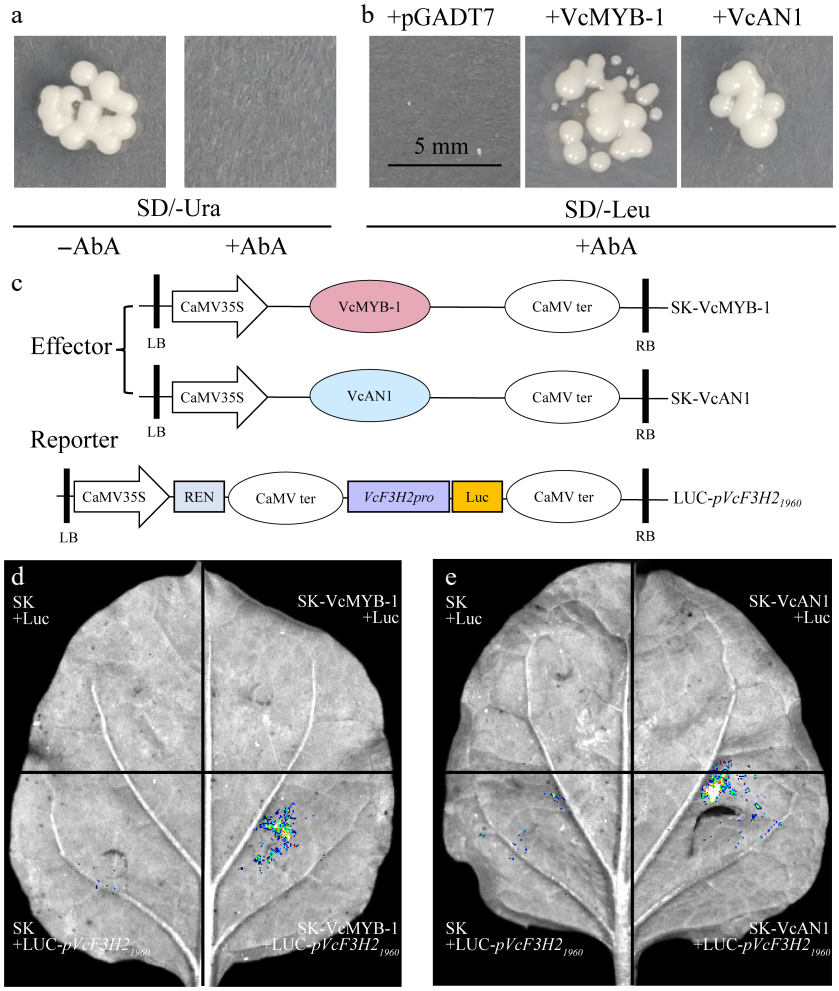

Fig. 6 Regulation of VcMYB-1 and VcAN1 on *VcF3H2* promoter. (a) Self-activation verification of the *VcF3H2* promoter. AbA: Aureobasidin A. (b) Y1H validation results. (c) Schematic diagrams for the effector and reporter constructs used for LUC; REN: Renilla luciferase. The stop codon removed CDS sequences of *VcMYB-1* and *VcAN1* were inserted into the pNC-Green-SK vector to generate effector constructs. The 1960 bp *VcF3H2* promoter sequence was cloned into pNC-Green-Luc vector to generate the reporter construct (LUC-$pVcF3H2_{1960}$). LB: left border; RB: right border. (d) and (e) LUC imaging results for the binding and activation abilities of VcMYB-1 and VcAN1 on the *VcF3H2* promoter, respectively.

enzyme of the plant flavonoids/anthocyanins biosynthesis process. In this study, we identified eight anthocyanin-related *F3Hs* from blueberry. All the identified VcF3Hs shared more than 80% similarity with several reported flavonoid/anthocyanin-related F3Hs from other plants, such as AtF3H[25], OsF3H[26], VvF3H[9], PeF3H[29], GmF3H[27], and so on. This indicated that plant anthocyanin-related F3Hs were highly conserved. Several key enzymes in the flavonoid/anthocyanin biosynthesis pathway, including C4H, CHI, CHS, F3H, FLS, DFR, and UFGT, were predicted to be interacting proteins of these VcF3Hs, indicating again that they play important roles in blueberry flavonoid/anthocyanin biosynthesis. The protein sequences of VcF3H2~4 were the same, with only one amino acid difference from VcF3H1. Moreover, their expression patterns in fruits at different ripening stages were almost the same. The other four VcF3Hs shared similarity greater than 96.78% with each other, but only slightly over 90% with VcF3H1~4. All these eight *VcF3Hs* were highly expressed in fruits at late ripening stages, but *VcF3H5~8* had different expression patterns from *VcF3H1~4*. Additionally, among the 12 duplicated gene pairs identified among the eight *VcF3Hs*, none were found between *VcF3H1~4* members and *VcF3H5~8* members. Collectively, it can be concluded that the eight anthocyanin-related *VcF3Hs* can be further categorized into two groups that play somewhat different roles in anthocyanin biosynthesis.

F3Hs play important roles in plant stress responses and their expression could be significantly affected by various stresses, and phytohormones[42,43]. The promoters of all anthocyanin-related *VcF3Hs* had ABA-responsive, drought-inducibility-related, defense, and stress related, and high-temperature responsive elements. Consistently, gene expression analysis revealed that ABA, alkali, salt, and LT, especially ABA, significantly upregulated the expression of almost all *VcF3Hs*. The upregulation of *D. officinale F3H* could increase anthocyanin and flavonol accumulation and confer plants with tolerance to salt and cold[12]. The overexpression of *L. chinense F3H* greatly enhanced the antioxidant ability of transgenic tobacco plants[13]. The ABA- and stress-inducible nature of these *VcF3Hs* may enhance the stress-resistance of blueberry in an anthocyanin-dependent way. Interestingly, high temperatures upregulated the expression of *VcF3H1~4*, but downregulated the expression of *VcF3H5~8*, suggesting that the two *VcF3Hs* groups play different roles in response to heat stress.

The upregulation or overexpression of *F3H* genes have been frequently reported to help in increasing anthocyanin accumulation[44]. Similarly, our study revealed that the transient overexpression of *VcF3H2*, the most differentially expressed *VcF3H* member during fruit ripening, resulted in significantly promoted anthocyanin accumulation and greatly upregulated expression of anthocyanin biosynthesis-related structural genes in apple and blueberry peels. Many studies have shown that TFs, such as MYB and bHLH, play important regulatory roles in anthocyanin biosynthesis by directly or indirectly influencing the expression of corresponding structural genes[45–47]. The rice MYC2 can activate the promoters of several anthocyanin biosynthesis genes, including *CHI*, *CHS*, *F3H*, *F3'H*, *DFR*, and *3GT*[48]. *Ginkgo biloba* MYB can directly bind to the promoters of *F3H* and some other structural genes, and influence the flavonol biosynthesis and accumulation in leaves[49]. In our study, binding sites for MYB and bHLH TFs were identified on the promoters of all eight anthocyanin-related *VcF3Hs*. Notably, MYB and bHLH binding sites on the *VcF3H2* promoter made up more than half of all TFBSs (accounting for 57.3%), suggesting that *VcF3H2* may be regulated by these two kinds of TFs. Consistently, our Y1H and LUC assay results showed that both the anthocyanin regulatory factors VcMYB-1 and VcAN1 could bind to and activate the *VcF3H2* promoter. Thus, the 'VcMYB-1/VcAN1-VcF3H2' module is concluded to play an important role in blueberry anthocyanins biosynthesis (Fig. 7).

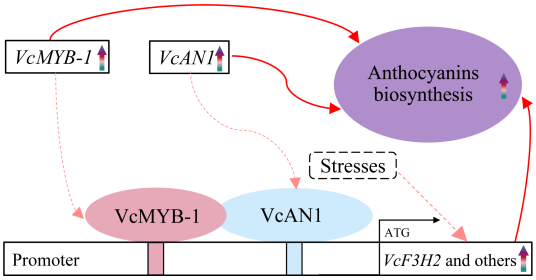

Fig. 7 The 'VcMYB/VcAN1-VcF3H2' module in regulating anthocyanin biosynthesis in blueberry. Rainbow arrows represent upregulation or increase during fruit ripening. Red arrows represent promoting effects. Solid arrows represent results have been validated in this study or previous studies, and dotted arrows represent results that need to be further confirmed.

In summary, we identified and characterized the eight anthocyanin-related *F3H* genes in blueberry, and demonstrated the contribution of *VcF3H2* to anthocyanin biosynthesis. By using Y1H and LUC assays, we validated the binding activities of two anthocyanin regulatory TFs (VcMYB-1 and VcAN1) on the *VcF3H2* promoter. These results will help elucidate the anthocyanin metabolism in blueberry. *VcF3H2* promoter has many stress-responsive elements and the gene's expression could be significantly influenced by multiple abiotic stresses and phytohormones, implying *VcF3H2* might be involved in stress-induced anthocyanin accumulation. Additionally, many binding sites for some other TFs were also identified in the *VcF3H2* promoter. In the future, further research should be conducted to verify the roles of *VcF3H2* in blueberry stress responses and to reveal the regulatory roles of other TFs on *VcF3H2* and other anthocyanin biosynthesis-related structural genes.

Author contributions

The authors confirm contribution to the paper as follows: study conception and design: Cheng C; data collection: Li R, Guo S, Hao S, Du S, Zhang J; analysis and interpretation of results: Li R, Guo S, Hao S, Du S, Liu R, Li J; technical assistance: Cheng C, Zhang Y; draft manuscript preparation: Li R, Cheng C. All authors reviewed the results and approved the final version of the manuscript.

Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

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Conflict of interest

The authors declare that they have no conflict of interest.

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References

1. Zhao L, Liu Y, Chen X, Zhang X, Liao L, et al. 2023. Visible light induces the *PpHYH* transcription to promote anthocyanin pigmentation in peach peel. *Fruit Research* 3:25
2. Liu R, Wang Y, Shu B, Xin J, Yu B, et al. 2025. SmHSFA8 enhances the heat tolerance of eggplant by regulating the SmEGY3 - SmCSD1 module and promoting SmF3H - mediated flavonoid biosynthesis. *Plant, Cell & Environment* 48:3085–104
3. Dai M, Kang X, Wang Y, Huang S, Guo Y, et al. 2022. Functional characterization of flavanone 3-hydroxylase (F3H) and its role in anthocyanin and flavonoid biosynthesis in mulberry. *Molecules* 27:3341
4. Ma G, Zhang L, Yamamoto R, Kojima N, Yahata M, et al. 2023. Molecular characterization of a flavanone 3-hydroxylase gene from citrus fruit reveals its crucial roles in anthocyanin accumulation. *BMC Plant Biology* 23:233
5. Maloney GS, DiNapoli KT, Muday GK. 2014. The *anthocyanin reduced* tomato mutant demonstrates the role of flavonols in tomato lateral root and root hair development. *Plant Physiology* 166:614–31
6. Jan R, Khan MA, Asaf S, Lee IJ, Kim KM. 2020. Overexpression of OsF_3H modulates WBPH stress by alteration of phenylpropanoid pathway at a transcriptomic and metabolomic level in *Oryza sativa*. *Scientific Reports* 10:14685
7. Li C, Liu S, Yao X, Wang J, Wang T, et al. 2017. PnF3H, a flavanone 3-hydroxylase from the Antarctic moss *Pohlia nutans*, confers tolerance to salt stress and ABA treatment in transgenic *Arabidopsis*. *Plant Growth Regulation* 83:489–500
8. Azuma A, Yakushiji H, Koshita Y, Kobayashi S. 2012. Flavonoid biosynthesis-related genes in grape skin are differentially regulated by temperature and light conditions. *Planta* 236:1067–80
9. Zheng Y, Tian L, Liu H, Pan Q, Zhan J, et al. 2009. Sugars induce anthocyanin accumulation and flavanone 3-hydroxylase expression in grape berries. *Plant Growth Regulation* 58:251–60
10. Teng S, Keurentjes J, Bentsink L, Koornneef M, Smeekens S. 2005. Sucrose-specific induction of anthocyanin biosynthesis in Arabidopsis requires the MYB75/PAP1 gene. *Plant Physiology* 139:1840–52
11. Loreti E, Povero G, Novi G, Solfanelli C, Alpi A, et al. 2008. Gibberellins, jasmonate and abscisic acid modulate the sucrose-induced expression of anthocyanin biosynthetic genes in *Arabidopsis*. *New Phytologist* 179:1004–16
12. Si C, Dong W, Teixeira Da Silva JA, He C, Yu Z, et al. 2023. Functional analysis of flavanone 3-hydroxylase (F3H) from *Dendrobium officinale*, which confers abiotic stress tolerance. *Horticultural Plant Journal* 9:356–64
13. Song X, Diao J, Ji J, Wang G, Guan C, et al. 2016. Molecular cloning and identification of a flavanone 3-hydroxylase gene from *Lycium chinense*, and its overexpression enhances drought stress in tobacco. *Plant Physiology and Biochemistry* 98:89–100
14. Mo R, Han G, Zhu Z, Essemine J, Dong Z, et al. 2022. The ethylene response factor ERF5 regulates anthocyanin biosynthesis in 'Zijin' mulberry fruits by interacting with *MYBA* and *F3H* genes. *International Journal of Molecular Sciences* 23:7615
15. Zou T, Wang X, Sun T, Rong H, Wu L, et al. 2023. MYB transcription factor $OsC1^{PLSr}$ involves the regulation of purple leaf sheath in rice. *International Journal of Molecular Sciences* 24:6655
16. Zhao Y, Zhang G, Tang Q, Song W, Gao Q, et al. 2022. *EbMYBP1*, a R2R3-MYB transcription factor, promotes flavonoid biosynthesis in *Erigeron breviscapus*. *Frontiers in Plant Science* 13:946827
17. Suresh S, Begum RF, Ankul Singh S, Chitra V. 2022. Anthocyanin as a therapeutic in Alzheimer's disease: a systematic review of preclinical evidences. *Ageing Research Reviews* 76:101595
18. Tran PHL, Tran TTD. 2021. Blueberry supplementation in neuronal health and protective technologies for efficient delivery of blueberry anthocyanins. *Biomolecules* 11:102
19. Zhang C, Guo Q, Liu Y, Liu H, Wang F, et al. 2017. Molecular cloning and functional analysis of a flavanone 3-hydroxylase gene from blueberry. *The Journal of Horticultural Science and Biotechnology* 92:57–64
20. Lin Y, Wang Y, Li B, Tan H, Li D, et al. 2018. Comparative transcriptome analysis of genes involved in anthocyanin synthesis in blueberry. *Plant Physiology and Biochemistry* 127:561–72
21. Xu L, Gao Z, Li L, Guo J. 2024. Impact of dielectric barrier discharge cold plasma on anthocyanin metabolism in blueberries: a targeted metabonomic and transcriptomic analysis. *Postharvest Biology and Technology* 213:112963
22. Zhang Y, Huang D, Wang B, Yang X, Wu H, et al. 2023. Characterization of highbush blueberry (*Vaccinium corymbosum* L.) anthocyanin biosynthesis related MYBs and functional analysis of *VcMYB* gene. *Current Issues in Molecular Biology* 45:379–99
23. Zhang Y, Liu F, Wang B, Wu H, Wu J, et al. 2021. Identification, characterization and expression analysis of anthocyanin biosynthesis-related *bHLH* genes in blueberry (*Vaccinium corymbosum* L.). *International Journal of Molecular Sciences* 22:13274
24. Zhang Y, Zhang Z, Guo S, Qu P, Liu J, et al. 2024. Characterization of blueberry *glutathione S-transferase* (*GST*) genes and functional analysis of *VcGSTF8* reveal the role of 'MYB/bHLH-GSTF' module in anthocyanin accumulation. *Industrial Crops and Products* 218:119006
25. Aguadé M. 2001. Nucleotide sequence variation at two genes of the phenylpropanoid pathway, the *FAH1* and *F3H* genes, in *Arabidopsis thaliana*. *Molecular Biology and Evolution* 18:1–9
26. Jan R, Asaf S, Paudel S, Lubna, Lee S, et al. 2021. Discovery and validation of a novel step catalyzed by *OsF3H* in the flavonoid biosynthesis pathway. *Biology* 10:32
27. Lim YJ, Lyu JI, Kwon SJ, Eom SH. 2021. Effects of UV-A radiation on organ-specific accumulation and gene expression of isoflavones and flavonols in soybean sprout. *Food Chemistry* 339:128080
28. Qiao F, Zhang K, Zhou L, Qiu QS, Chen Z, et al. 2022. Analysis of flavonoid metabolism during fruit development of *Lycium chinense*. *Journal of Plant Physiology* 279:153856
29. Hsu CC, Chen YY, Tsai WC, Chen WH, Chen HH. 2015. Three R2R3-MYB Transcription factors regulate distinct floral pigmentation patterning in *Phalaenopsis* spp. *Plant Physiology* 168:175–91
30. Yuan Y, Ma X, Shi Y, Tang D. 2013. Isolation and expression analysis of six putative structural genes involved in anthocyanin biosynthesis in *Tulipa fosteriana*. *Scientia Horticulturae* 153:93–102
31. Chen C, Chen H, Zhang Y, Thomas HR, Frank MH, et al. 2020. TBtools: an integrative toolkit developed for interactive analyses of big biological data. *Molecular Plant* 13:1194–202
32. Wang Y, Tang H, DeBarry JD, Tan X, Li J, et al. 2012. *MCScanX*: a toolkit for detection and evolutionary analysis of gene synteny and collinearity. *Nucleic Acids Research* 40:e49
33. Chou KC, Shen HB. 2008. Cell-PLoc: a package of Web servers for predicting subcellular localization of proteins in various organisms. *Nature Protocols* 3:153–62
34. Jaakola L, Määttä K, Pirttilä AM, Törrönen R, Kärenlampi S, et al. 2002. Expression of genes involved in anthocyanin biosynthesis in relation to anthocyanin, proanthocyanidin, and flavonol levels during bilberry fruit development. *Plant Physiology* 130:729–39
35. von Mering C, Jensen LJ, Snel B, Hooper SD, Krupp M, et al. 2005. STRING: known and predicted protein–protein associations, integrated and transferred across organisms. *Nucleic Acids Research* 33:D433–D437
36. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, et al. 2003. Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Research* 13:2498–504
37. Lv Y, Zhang M, Wu T, Wu T, Zhong Y. 2019. The infiltration efficiency of *Agrobacterium*-mediated transient transformation in four apple cultivars. *Scientia Horticulturae* 256:108597
38. Zhang Z, Qu P, Hao S, Li R, Zhang Y, et al. 2023. Characterization and functional analysis of *chalcone synthase* genes in highbush blueberry (*Vaccinium corymbosum*). *International Journal of Molecular Sciences* 24:13882

39. Hu Y, Gong Z, Yan Y, Zhang J, Shao A, et al. 2024. ChBBX6 and ChBBX18 are positive regulators of anthocyanins biosynthesis and carotenoids degradation in *Cerasus humilis*. *International Journal of Biological Macromolecules* 282:137195
40. Wang N, Liu W, Zhang T, Jiang S, Xu H, et al. 2018. Transcriptomic analysis of red-fleshed apples reveals the novel role of MdWRKY11 in flavonoid and anthocyanin biosynthesis. *Journal of Agricultural and Food Chemistry* 66:7076−86
41. Chen H, Zou Y, Shang Y, Lin H, Wang Y, et al. 2008. Firefly luciferase complementation imaging assay for protein-protein interactions in plants. *Plant Physiology* 146:323−24
42. Bai Y, Zou R, Zhang H, Li J, Wu T. 2024. Functional characterization of *CsF3Ha* and its promoter in response to visible light and plant growth regulators in the tea plant. *Plants* 13:196
43. Jan R, Aaqil Khan M, Asaf S, Lubna, Park JR, et al. 2021. Flavonone 3-hydroxylase relieves bacterial leaf blight stress in rice via overaccumulation of antioxidant flavonoids and induction of defense fenes and hormones. *International Journal of Molecular Sciences* 22:6152
44. Yang C, Chen W, Tang D, Tan X, Tan L, et al. 2024. Metabolomic and transcriptomic insights into anthocyanin biosynthesis in 'Ziyan' tea plants under varied photoperiod and temperature conditions. *Agronomy* 14:56
45. Dubos C, Le Gourrierec J, Baudry A, Huep G, Lanet E, et al. 2008. MYBL2 is a new regulator of flavonoid biosynthesis in *Arabidopsis thaliana*. *The Plant Journal* 55:940−53
46. Schaart JG, Dubos C, Romero De La Fuente IR, van Houwelingen AMML, de Vos RCH, et al. 2013. Identification and characterization of MYB-bHLH-WD40 regulatory complexes controlling proanthocyanidin biosynthesis in strawberry (*Fragaria* × *ananassa*) fruits. *New Phytologist* 197:454−67
47. Liu X, Zhai Y, Liu J, Xue J, Markovic T, et al. 2023. Comparative transcriptome sequencing analysis to postulate the scheme of regulated leaf coloration in *Perilla frutescens*. *Plant Molecular Biology* 112:119−42
48. Li N, Xu Y, Lu Y. 2024. A regulatory mechanism on pathways: modulating roles of MYC2 and BBX21 in the flavonoid network. *Plants* 13:1156
49. Wang G, Zhang L, Wang G, Cao F. 2022. Growth and flavonol accumulation of *Ginkgo biloba* leaves affected by red and blue light. *Industrial Crops and Products* 187:115488