

Regulatory function of plant basic region/leucine zipper transcription factors in stress, metabolism, and developmental processes

Yingjie Mao^{1#}, Ruikang Zhao^{2#}, Yali Zhang¹ and Mingxing Tu^{1*}

¹ College of Enology, College of Food Science and Engineering, Viti-viniculture Engineering Technology Center of State Forestry and Grassland Administration, Shaanxi Engineering Research Center for Viti-Viniculture, Heyang Viti-Viniculture Station, Northwest A & F University, Yangling, Shaanxi 712100, China

² State Key Laboratory of Crop Stress Biology in Arid Areas, College of Horticulture, Northwest A & F University, Yangling, Shaanxi 712100, China

[#] Authors contributed equally: Yingjie Mao, Ruikang Zhao

* Corresponding author, E-mail: tumingxing@nwfu.edu.cn

Abstract

The basic region/leucine zipper (bZIP) proteins represent one of the largest transcription factor (TF) families and have been identified in various plant species. The plant bZIPs regulate growth and development processes, are closely linked to plant metabolism, and play key roles in responses to biotic and abiotic stresses. This review specifically summarizes the regulatory functions of plant bZIP TFs in abiotic stresses (drought, salinity, temperature extremes, and heavy metals), biotic stresses (pathogens and herbivorous insects), primary and secondary metabolism, and developmental processes (seed maturation, fruit ripening, organ senescence). We highlight mechanistic models, unresolved controversies, and future research directions, providing a theoretical basis for crop improvement.

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Introduction

The transcription initiation process involves interactions among various proteins, including transcription factors (TFs). Plant TFs serve as master regulators of developmental processes and responses to environmental stimuli. On the basis of their conserved DNA-binding domains, TFs are classified into several families, such as APETALA2/ethylene-responsive element binding protein (AP2/EREBP), basic region/leucine zipper (bZIP), MYB, WRKY, and zinc finger proteins. The bZIP family represents a major group of TFs. The number of bZIP TFs varies markedly among different plant species, ranging from fewer than 20 in lower plants to nearly 200 in polyploid crops (Supplementary Table S1). This variation is primarily shaped by differences in genome size and complexity, lineage-specific duplication events, and evolutionary adaptations to ecological niches. For example, whole-genome and segmental duplications in species such as *Glycine max* (soybean) and *Gossypium hirsutum* (cotton) have greatly expanded the bZIP family, whereas woody perennials like *Vitis vinifera* (grapevine) and *Populus trichocarpa* (poplar) often have fewer but highly specialized members^[1,2]. Functional diversification following duplication has allowed bZIP subfamilies to acquire roles in stress responses, metabolism, and development, contributing to species-specific regulatory networks. Therefore, the observed differences in bZIP gene counts across taxa reflect both the genomic architecture and adaptive selection pressures.

TFs are DNA-binding proteins that regulate gene expression by interacting specifically with *cis*-acting elements in promoter regions. Transcriptional activators, including bZIPs, typically have two main functional domains: A DNA-binding domain and a transcriptional activation domain. Studies have shown that bZIPs play critical roles in plants' development and biotic and abiotic stress responses throughout their lifecycle. The DNA-binding domain of the bZIP family, known as the basic leucine zipper, determines binding

specificity. It comprises a leucine zipper that enables dimerization and a basic region that binds to DNA. In the leucine zipper, a hydrophobic leucine residue appears every seven amino acids along the same side of the α -helix, facilitating the formation of a clamp-like dimer that binds DNA. The basic region anchors to the DNA's major groove. The DNA-binding specificity of bZIP TFs is governed by this domain, which recognizes *cis*-elements containing the core ACGT motif, such as CACGTG (G-box), GACGTC (C-box), and TACGTA (A-box)^[3].

The bZIPs are involved in plants' stress responses

Plants are frequently challenged by abiotic stresses such as drought, low temperature, and salinity, as well as biotic stresses like diseases and insect pests. These stresses disrupt normal plant growth and development, often causing damage or even death. Therefore, understanding plants' responses and resistance to stress is crucial. Plant cells perceive stress signals, such as water deficit or high salinity, through receptors and activate signal transduction pathways, with abscisic acid (ABA) playing a key role in regulating gene expression under abiotic stress. Stress also affects transcriptional regulation by activating or repressing TFs and inducing epigenetic modifications. Physiologically, plants respond by synthesizing osmotic regulators like proline and betaine, activating antioxidant defense systems to scavenge reactive oxygen species (ROS), and producing secondary metabolites like anthocyanins^[4]. Additionally, stress leads to modifications in the cell wall's composition and changes in roots' development and morphology to enhance adaptability. TFs are central to these responses, regulating downstream gene expression to facilitate adaptation to environmental challenges. Among these, the bZIP family is one of the most extensively studied, with numerous reports highlighting its role in modulating stress tolerance through various mechanisms. This review summarizes the key roles of bZIP TFs in regulating plants' stress responses.

Drought stress responses

Plants can mitigate drought stress through several mechanisms, including enhanced antioxidant enzyme activity, increased osmolyte accumulation, and regulation of endogenous hormone levels (Fig. 1).

Under drought stress, the production of ROS in crops leads to lipid peroxidation of the cell membranes, disrupting normal plant metabolism. In plants, drought stress tolerance is positively correlated with ROS-scavenging capacity. To mitigate the harmful effects of ROS, plants protect the integrity of cell membranes, proteins, and metabolic enzymes by accumulating antioxidant enzymes. Therefore, peroxidase (POD) levels can, to some extent, reflect a plant's drought resistance. Osmotic substances include soluble sugars, proline, betaine, and inorganic ions such as Ca²⁺, Na⁺, and Cl⁻. These solutes, which exhibit affinity within plant cells, rapidly accumulate under abiotic stress to maintain cell turgor and protect cellular structures. Studies have shown that bZIPs can enhance drought resistance by regulating POD activity and osmotic substances. For example, the *AtbZIP62* mutant exhibited greater susceptibility to drought stress than wild-type *Arabidopsis thaliana*, as *AtbZIP62* regulates catalase (CAT), POD, and polyphenol oxidase (PPO)-like activities and increases proline and malondialdehyde (MDA) levels, thereby enhancing drought resistance^[5]. Heterologous expression of *VvABF2* in *A. thaliana* significantly improved ROS scavenging capacity, reduced membrane damage, and enhanced drought

tolerance^[6]. Similarly, *GhABF2* in cotton has been shown to boost the activity of peroxidases such as superoxide dismutase (SOD) and CAT, improving drought resistance^[7]. Likewise, *FtbZIP83*, *AhAREB1*, and *ThbZIP1* improve drought tolerance by enhancing ROS scavenging and osmoregulatory capacity^[8–10].

Hormones are key signaling molecules in plants that regulate signal transduction and play crucial roles in plants' stress responses. ABA is central to plants' stress responses, with bZIP TFs playing a vital role in ABA signaling. Specifically, the A subfamily of bZIPs acts as a master regulator in ABA-dependent responses and mediates hormone-driven drought responses. As noted earlier, several bZIP TFs bind to ABA-responsive elements (ABREs) and function in drought stress through the ABA signaling pathway. ABRE-binding factors (ABFs) and AREBs are induced by abiotic stresses and play a key role in regulating the expression of downstream stress-responsive genes via ABA signaling, thus enhancing drought resistance^[11]. ABFs/AREBs participate in ABA-mediated stress responses activated by SnRK2s (SNF1-related protein kinases). Studies have shown that ABA signaling is regulated by the phosphorylation and dephosphorylation of SnRK2s^[12]. Endogenous ABA activates receptor proteins, such as PYR (pyrabactin resistance), RCAR (regulatory components of ABA receptors), and PYL (PYR1-like), which interact with PP2Cs (Type 2C protein phosphatases) to release SnRK2s. These kinases then phosphorylate and activate ABF/AREB TFs. The activated ABFs/AREBs induce ABA-responsive genes to regulate stress responses^[13].

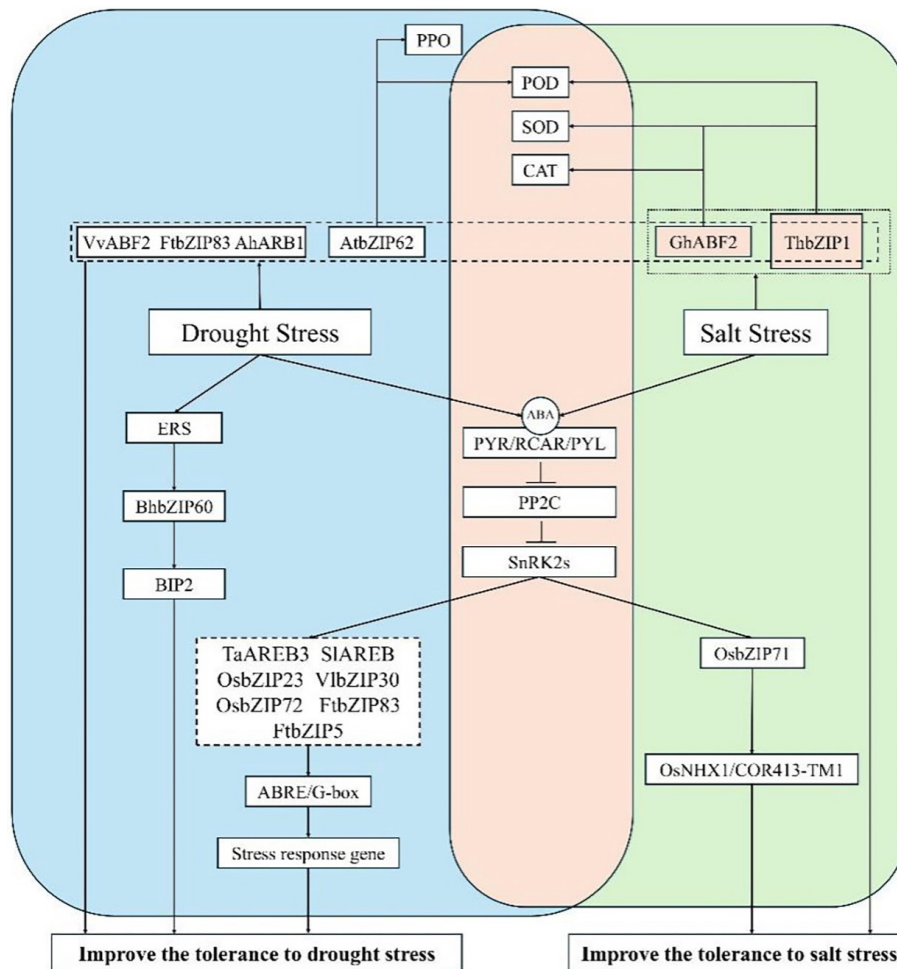


Fig. 1 bZIPs respond to drought stress and salt stress. Abbreviations: PPO, polyphenol oxidase; POD, peroxidase; SOD, superoxide dismutase; CAT, catalase; ERS, endoplasmic reticulum stress; PYR, pyrabactin resistance; PYL, PYR1-like; RCAR, regulatory component of ABA receptor; PP2C, protein phosphatase 2C; SnRK2, SNF1-related protein kinase 2; ABF, ABRE-binding factors; AREB, ABRE-binding proteins; JA, jasmonic acid.

Arabidopsis srk2d/e/i triple mutants exhibit significantly reduced drought tolerance, caused by impaired SnRK phosphorylation of AREB1. This limits the binding of AREB1 to ABREs, weakening downstream stress gene regulation. In addition to *Arabidopsis*, numerous bZIP TFs that bind ABREs have been identified as responsive to water stress in other species. For example, overexpression of *TaAREB3* in *Arabidopsis* enhances ABA sensitivity and drought resistance^[14]. In rice (*Oryza sativa*), *OsbZIP23* conferred ABA-dependent drought and salinity tolerance and shows strong potential for improving stress resistance through genetic modification^[15]. Overexpression of *OsbZIP72* in rice significantly enhances ABA-responsive genes' expression and drought resistance compared with wild-type plants^[16]. Hsieh et al. reported that the tomato (*Solanum lycopersicum*) bZIP TF *SIAREB* binds ABREs, mediating ABA signaling and improving drought resistance^[17]. Overexpressing *VlbZIP30* from grapevine in *Arabidopsis* enhances drought resistance via G-box (CACGTG)-mediated regulation of ABA-responsive genes^[18]. In tartary buckwheat (*Fagopyrum tataricum*), *FtbZIP83* and *FtbZIP5* act as positive regulators of drought resistance through ABA signaling, directly binding to *FtSnRK2.6/2.3* and *FtSnRK2.6*, respectively^[9,19]. Interestingly, bZIP TFs not only act as downstream components of ABA signaling but also contribute to its negative feedback regulation. Some studies show that ABFs can rapidly induce PP2Cs under ABA treatments, thereby negatively regulating ABA signaling. In rice, *OsbZIP23* was also found to positively regulate *OsPP2C49*, providing further evidence of negative feedback regulation in ABA signaling^[20]. Pepper (*Capsicum annuum* L.) *CaATBZ1* negatively regulated drought responses by regulating ABA-induced stomatal closure^[21].

In addition, some bZIP TFs also participate in other plant hormone signaling pathways and enhance drought resistance. Their regulatory roles involve cross-talk among the gibberellic acid (GA)/brassinosteroid (BR), salicylic acid (SA), and auxin signaling cascades, including limiting shoot growth via hormonal balance, mitigating oxidative stress through SA-induced antioxidant biosynthesis, and hydraulic adjustment mediated by auxin-driven root patterning^[22]. In *Arabidopsis*, the AtTGA2/3/5/6-NPR1 protein interaction enhances drought tolerance by activating jasmonic acid (JA)-responsive genes, boosting ROS scavenging, and reducing drought-induced oxidative membrane damage through coordinated molecular mechanisms. In maize (*Zea mays*), *ZmbZIP4* positively regulates gibberellin biosynthesis, thereby improving resistance to abiotic stress^[23]. *MdIPT5b* encodes the rate-limiting enzyme isopenyltransferase in the cytokinin biosynthesis pathway and represses gene expression, thus delaying drought-induced premature leaf senescence by reducing oxidative damage and sustaining plant growth. This contributes to drought and salt stress responses via the cytokinin pathway^[24].

Salt stress responses

As a major abiotic stressor, salt stress impairs plants' growth and development through interconnected mechanisms, including ion toxicity, osmotic imbalance, oxidative stress, and disruption of nutrient homeostasis (Fig. 1).

bZIP TFs act as central regulators in plants' responses to salinity stress. By integrating ABA signaling cascades, ROS regulatory networks, and ion homeostasis, these transcriptional modulators coordinate the expression of stress-responsive genes to mediate adaptation to ionic and osmotic challenges under saline conditions. Hsieh et al. functionally characterized the tomato bZIP TF *SIAREB*, demonstrating its role in reprogramming stress-responsive gene networks and conferring salinity tolerance in both *Arabidopsis* (heterologous) and tomato (homologous) systems^[17]. Transgenic

overexpression of *MhNPR1* from *Malus hupehensis* in tobacco (*Nicotiana tabacum*) confers dual stress tolerance by alleviating ionic imbalance and osmotic stress^[25]. Pepper (*Capsicum annuum* L.) *CabZIP25* is strongly induced by abiotic stresses and phytohormones and positively regulates salt tolerance; its expression level correlates with chlorophyll content in peppers^[26]. *Tamarix hispida* exhibits halophytic adaptation through integrated physiological mechanisms, enabling growth under saline conditions. The *Tamarix hispida* *ThbZIP1* TF enhances POD and SOD activity, mitigating oxidative stress during exposure to salinity^[27].

Salinity stress induces ABA biosynthesis, which promotes adaptation through SnRK2-mediated phosphorylation of bZIP TFs, enabling their nuclear translocation and activation of stress-responsive genes. Overexpression of *OsbZIP71* in *Oryza sativa* enhanced salinity tolerance by directly targeting the *OsNHX1* and *COR413-TM1* promoters, activating their transcription^[28]. *OsNHX1* contributes to Na⁺ homeostasis via ion compartmentalization and mediates osmotic regulation, offering dual protective mechanisms under salinity stress. *OsbZIP71*-overexpressing lines show hypersensitivity to ABA and osmotic stimuli, along with transcriptional activation of ABA-inducible markers (*RD29B*, *RAB18*, *HIS1-3*), highlighting its role as an ABA-responsive transcriptional activator that integrates stress signaling for improved crop resilience to salinity^[29]. Overexpression of *IbbZIP1* in *Arabidopsis* elevated ABA accumulation, conferring tolerance to both salinity and drought through ABA-mediated signaling^[30]. Cotton plants overexpressing *GhABF2* show increased activities of the key ROS-scavenging enzymes SOD and CAT, resulting in improved salinity tolerance via enhanced mitigation of oxidative stress^[7].

A key transcriptional regulator orchestrates salinity-responsive gene networks in plants. Rolly et al. demonstrated that *AtbZIP62* transcriptionally suppresses salinity responses beyond canonical ABA signaling. bZIP TFs also mediate salinity responses through ABA-independent mechanisms^[31]. For instance, expressing the *AtbZIP60* gene enhances salt tolerance in rice by upregulating Ca²⁺-dependent protein kinase genes (*OsCPK6/9/10/19/25/26*) under salt stress. This suggests that overexpression of *AtbZIP60* in transgenic cell lines enhances salt tolerance by modulating Ca²⁺-dependent protein kinase genes. In *Arabidopsis*, *AtbZIP17* TF is reportedly activated in response to salt stress, with genetic evidence supporting its role in a conserved stress signaling pathway. Under salinity stress, endoplasmic reticulum (ER)-localized *AtbZIP17* undergoes proteolytic processing, releasing its DNA-binding domain for nuclear translocation to activate salt-responsive targets including *ATHB-7*. An activated form of *AtbZIP17* has been shown to confer salt tolerance in *Arabidopsis*^[32]. Furthermore, *AtbZIP24* has been functionally validated as a negative regulator of salinity tolerance in *Arabidopsis* by repressing salt overly sensitive (SOS) pathway-associated genes^[31].

Notably, some studies have reported contradictory results regarding the roles of identical bZIP transcription factors under different stress conditions. For example, *AtbZIP62* has been described as a positive regulator of drought tolerance through the modulation of ROS-scavenging enzymes' activity and osmolyte accumulation^[5], whereas other studies suggest that its overexpression may impair plant growth under prolonged or high-intensity drought stress^[24]. Such inconsistencies may arise from differences in experimental design, including the severity and duration of stress treatments, as well as variations in the plants' developmental stages and tissue-specific expression patterns. Similarly, in rice, *OsbZIP23* is generally regarded as a positive regulator of ABA-dependent drought and salt responses^[15], yet feedback regulation through PP2C induction

indicates a potential negative role in ABA signaling under certain contexts^[20]. These discrepancies highlight the importance of considering the plants' genetic background, the environmental conditions, and the intensity of stress when interpreting functional studies of bZIP TFs. Future investigations integrating standardized stress treatments, multi-omics analyses, and cross-species comparisons will be essential to reconcile these divergent findings and establish a clearer mechanistic framework.

Low-temperature stress responses

Low-temperature stress severely limits plants' growth and productivity, forcing plants to activate adaptive responses through transcriptional reprogramming (Fig. 2).

bZIP transcription factors function as pivotal regulators, integrating ABA signaling, antioxidant defense, and cryoprotective gene expression to enhance cold tolerance. Within the S1-bZIP subfamily, low-temperature-induced protein 19 (*LIP19*) is strongly induced under cold conditions in monocots, and functional studies in *O. sativa* confirm its central role in cold-responsive regulatory networks^[33]. *OsOBF1*, a paralog of *LIP19* within the S1-bZIP family in *O. sativa*, exhibits temperature-dependent functional synergy with *LIP19* in thermosensory regulation. At normal temperatures (25 °C), *OsOBF1* maintains constitutive transcriptional activity to regulate basal temperature signaling homeostasis. During cold exposure (4–10 °C), *LIP19* undergoes structural changes enabling heterodimerization with *OsOBF1*, thereby allosterically modulating its DNA-binding specificity and reprogramming transcriptional networks toward cold acclimation. Mutational studies show that loss of *LIP19* function leads to reduced survival at 4 °C, whereas co-expression of *LIP19* and *OsOBF1* restores cold tolerance by activating *OsDREB1* genes and enhancing soluble sugar accumulation. In *Olea europaea*, increased expression of *OeLIP19*, together with *OeFAD8* (a fatty acid desaturase) and *OeOSM* (osmotin), has been associated with enhanced membrane fluidity and osmoprotection in cold-acclimated leaves of this evergreen perennial^[34]. Recent advances in the functional genomics of rice have identified multiple bZIP TFs (*OsZIP08*, *OsZIP35*, *OsZIP38*, *OsZIP46*, *OsZIP63*, *OsZIP72*, *OsZIP73*, and *OsZIP76*) as key regulators of chilling stress adaptation through both ABA-dependent and independent pathways^[35]. Similarly, low-temperature treatment (4 °C for 24 h) significantly

induces the expression of *RsLIP19*, a bZIP TF in *Raphanus sativus* roots, which enhanced chilling tolerance by modulating ROS scavenging systems and osmolyte biosynthesis^[36].

Additionally, bZIP TFs participate in cold stress responses in various ways. For example, in soybean, *GmbZIP44* activates the expression of the low-temperature-regulated genes *COR15A* and *COR78*, promoting proline synthesis and enhancing chilling tolerance^[37]. In *Oryza sativa*, *OsZIP73* is among the best characterized bZIPs in cold tolerance at the reproductive stage. Under chilling conditions, the *japonica* allele *bZIP73^{Jap}* physically interacts with *OsZIP71* to form heterodimers; this complex enhances the expression of sugar metabolism genes such as *OsINV4* (cell wall invertase) and *OsMST7/8* (monosaccharide transporters) to promote sucrose cleavage and hexose transport toward the anthers^[35]. In contrast, *CsbZIP18* functions as a negative regulator of ABA-mediated cold stress signaling in *Cucumis sativus*. Heterologous overexpression of *CsbZIP18* in *Arabidopsis* reduced ABA sensitivity and impaired cold acclimation compared with wild-type plants under freezing conditions^[35].

In conclusion, bZIP transcription factors function as central regulators of plants' cold stress tolerance by integrating ABA signaling, ROS detoxification, and expression of cryoprotective genes. Both positive and negative regulators exist across species, reflecting functional diversity. Future research should clarify their hierarchical networks and molecular interactions to improve crops' resilience under low-temperature conditions.

Heat stress responses

Heat stress disrupts proteins' stability, membrane integrity, and photosynthetic efficiency in plants.

To cope with high temperatures, bZIP transcription factors integrate ER stress signaling, activation of the unfolded protein response (UPR), and antioxidant defense, thereby orchestrating transcriptional networks that are essential for thermotolerance and survival under elevated temperatures. Zhang et al. (2008) reported that transgenic *Arabidopsis* lines overexpressing *ABP9* exhibited improved photosynthetic efficiency under heat shock compared with wild-type plants, attributed to *ABP9*'s role in stabilizing the Photosystem II (PSII) complex and reducing thylakoid membrane disorganization^[38]. The ER is a key organelle for protein biosynthesis in eukaryotic cells. Under biotic or abiotic stress, misfolded proteins accumulate abnormally in the ER lumen, triggering ERS through the

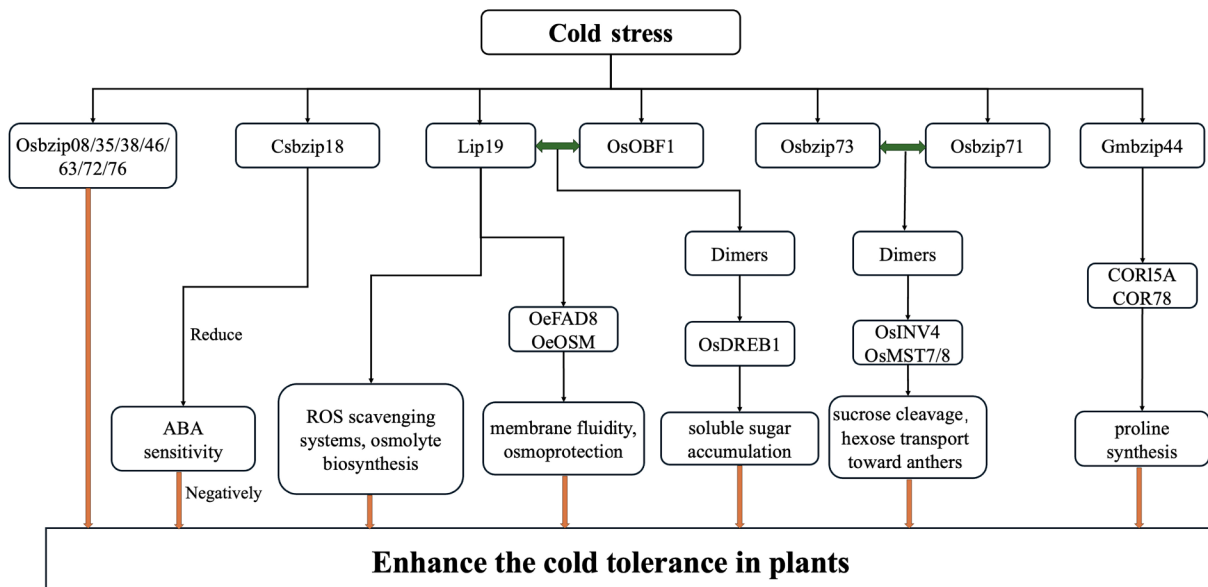


Fig. 2 bZIPs responding to low-temperature stress.

UPR. This condition leads to excess ROS production and programmed cell death. Mechanistically, the activation of canonical Endoplasmic Reticulum Stress (ERS) biomarkers, including molecular chaperones such as binding immunoglobulin protein (BiP) and calnexin (CNX), constitutes a critical adaptive response. These chaperones mitigate proteotoxic stress through two coordinated pathways: Targeted degradation of oxidatively damaged polypeptides and upregulation of antioxidant defense systems^[39,40]. Studies have shown that bZIP TFs from the K subfamily (e.g., *AtbZIP60* in *Arabidopsis* and *OsbZIP74* in rice) activate ERS-responsive genes by binding directly to ER stress elements (ERSEs) (e.g., CCAAT-N9-CCACG; ERSE-II, ATTGG-N-CCACG; UPRE-I, CAGCGTG; UPRE-II, TCATCG), thereby mitigating ERS^[41]. In *Arabidopsis*, the inositol-requiring enzyme 1 (IRE1) kinase-ribonuclease is activated by heat stress via recognition of misfolded proteins in the ER lumen. Its C-terminal domain splices *AtbZIP60* mRNA, generating a transcript that encodes an active TF—a conserved UPR mechanism that restores ER homeostasis. The IRE1-processed *AtbZIP60* mRNA isoform functions as a transcriptional activator, regulating UPR-associated genes to mitigate ERS-induced cellular damage^[42]. Maize *bZIP60* knockdown lines showed reduced thermotolerance under heat stress, indicating its essential role in heat stress adaptation via UPR activation^[43]. ERS is also involved in drought stress responses. A strong correlation has been observed between ER stress and drought resistance^[44–46]. ERS modulates drought tolerance through multiple mechanisms. For example, heterologous expression of *BhbZIP60S* in *Arabidopsis* upregulates the ER stress marker *BiP2*, enhancing drought tolerance, and deletion of ER stress-responsive cis-elements within the 5'-untranscribed region (UTR) of maize *ZmPP2C-A10* improves drought resistance. Conversely, virus-induced gene silencing (VIGS) of the ER chaperone *CaBiP1* in pepper increases ROS and MDA levels, impairing drought adaptation^[39,45,47].

Heavy metal stress responses

The toxicity of heavy metals, such as cadmium (Cd) and copper (Cu), poses severe threats to plants' growth and development by inducing oxidative stress, disrupting ion homeostasis, and damaging cellular structures. bZIP TFs function as key regulators in these processes, modulating detoxification, ion transport, and ROS scavenging systems. In tobacco, overexpression of *NtbZIP62* improves Cd tolerance by inducing antioxidant enzymes and phytochelatin synthase, which chelate Cd and reduce ROS-induced damage^[48]. Similarly, in the Cd-hyperaccumulator *Sedum plumbizincicola*, *SpbZIP60* enhances tolerance by promoting lignification of root cell walls, thereby restricting Cd influx^[49]. These results indicate that bZIP TFs orchestrate multiple protective mechanisms to mitigate heavy metal toxicity.

In addition to Cd and Cu detoxification, bZIP TFs also play a central role in zinc homeostasis. Zinc (Zn) is an essential micronutrient that functions as a structural motif and catalytic cofactor in numerous proteins, and its deficiency leads to oxidative damage, genomic instability, and reduced yield. In *Arabidopsis*, *bZIP19* and *bZIP23* act as master regulators of Zn deficiency responses, sensing Zn²⁺ via their N-terminal zinc-sensing modules (ZSMs) and activating downstream transporters such as *ZIP1*^[50]. Notably, reduced expression of *bZIP19* and *bZIP23* increases Zn and Cd accumulation in roots and shoots, further confirming their dual roles in nutrient balance and heavy metal detoxification^[51].

In summary, bZIP TFs function as versatile regulators of abiotic stress adaptation, including drought, cold, heat, salinity, and heavy metal toxicity. Their primary roles include modulating ABA signaling, ROS homeostasis, ion transport, and secondary metabolism. Both positive and negative regulatory functions exist across bZIP subgroups, highlighting their complexity and functional diversity.

Recent studies on *AtbZIP60*, *ZmbZIP29*, *OsbZIP50*, *AtbZIP17*, and *OsbZIP68* provide new insights into the molecular mechanisms of abiotic stress regulation. However, unresolved issues remain regarding pathway cross-talk, redundancy among subfamily members, and integration with epigenetic regulation. Future research should employ multi-omics and genome editing approaches to systematically dissect these mechanisms and develop stress-resilient crops.

Biotic stress responses

In addition to coordinating abiotic stress adaptation, the bZIP TF family regulates biotic stress responses. Notably, these transcriptional regulators positively influence pathogen resistance through SA- and ABA-mediated signaling modules, indicating their dual functionality in stress cross-talk. SA, synthesized as a phytoimmune signal upon pathogen attack, orchestrates systemic acquired resistance (SAR) via transcriptional reprogramming, thereby conferring broad-spectrum disease resistance in plants. SA also induces the accumulation of excess pathogenesis-related (PR) proteins in *Arabidopsis*, which play a key role in disease defense.

TGACGmotif-binding factor (TGA) TFs belonging to the D subfamily of bZIPs play a key role in the SA signaling pathway^[1]. Upon pathogen infection, the SA level in plants rises sharply, inducing the dissociation of intracellular NPR1 oligomers into monomers. As a transcriptional coactivator, the NPR1 monomer binds to TGA TFs, enhancing their binding affinity and stability with the target gene promoters (containing as-1-like elements). This process strongly activates the expression of downstream defense genes, such as *PR-1*^[52]. Zhang et al. demonstrated the redundancy and function of *TGA2*, *TGA5*, and *TGA6* in SAR. Knockout of these genes adversely affected *PR* gene expression and pathogen resistance in *Arabidopsis*^[53]. *TabZIP74* mRNA splicing is triggered by ER stress, which is induced by *Pst* (*Puccinia striiformis f. sp. tritici*) infection and wounding. Thus, *TabZIP74* participates in the ABA pathway to regulate the defenses of wheat (*Triticum aestivum*) against *Pst*^[54]. Moreover, other studies proved that *OsbZIP1* enhances rice's resistance to blast disease through the SA signaling pathway^[55]. *OsbZIP1* expression is induced by rice blast infection, and it responds rapidly to SA, JA, and ABA. Its role in enhancing resistance is mediated through an SA-dependent signaling pathway^[55].

bZIP TFs also play crucial roles in integrating stress signaling with herbivory defense. In *Arabidopsis*, *AtbZIP10* interacts with JA-regulated pathways while balancing ROS accumulation, thereby strengthening cell wall barriers and enhancing resistance to leaf-chewing pests^[56]. Similarly, *AtbZIP11* influences amino acid metabolism and the synthesis of phenylpropanoid-derived compounds, altering the nutritional and chemical profiles to deter herbivores. Furthermore, specific bZIP subfamilies act as central regulators of JA-responsive defense programs^[57]. In particular, *Arabidopsis* Class II TGAs (*TGA2/5/6*) function as essential activators of the JA/ethylene branch, driving the expression of PDF1.2 and b-CHI via the ERF1/ORAS9 node and forming a TGACG-box-centered transcriptional module for antiherbivory defenses. Supporting this, feeding assays with *Spodoptera exigua* demonstrated that the *tga2/5/6* triple mutant displays altered expression of JA/SA markers, confirming the role of TGAs in modulating JA-dependent transcription during herbivore attack^[58]. At the molecular level, herbivory-induced jasmonoyl-L-isoleucine (JA-Ile) promotes COI1-mediated degradation of the jasmonate ZIM domain (JAZ), releasing ethylene response TF (ERF) and MYC regulators, whereas TGAs cooperate at the target promoters to potentiate the expression of PDF12/VSP2^[59]. Collectively, TGA bZIPs act as hubs linking JA signaling to direct defenses, including proteinase inhibitors and antimicrobial proteins, thereby enhancing plants' resistance under insect pressure^[59].

Table 1. Roles of bZIP transcription factors in abiotic stress and biotic stress responses across species.

Stress type		Key bZIP members	Species	Subgroup	Mechanistic role
Abiotic stress	Drought	<i>AtbZIP62, VvABF2, FtbZIP83, AhAREB1, ThbZIP1, GhABF2, FtbZIP83, FtbZIP5, OsbZIP23, VlbZIP30</i>	<i>Arabidopsis thaliana, Vitis vinifera, Gossypium hirsutum, Fagopyrum tataricum</i>	A	ABA-SnRK2-AREB pathway activation; ROS scavenging; osmolyte accumulation
	Salinity	<i>OsbZIP23, OsbZIP71, SIAREB, ThbZIP1, CabZIP25</i>	<i>Oryza sativa, Solanum lycopersicum, Tamarix hispida, Capsicum annuum</i>	A/K	ABA-responsive regulation; Na ⁺ /H ⁺ homeostasis; activation of antioxidant enzymes
	Low temperature	<i>LIP19, OsOBF1, RsLIP19, GmbZIP44, CsbZIP18</i>	<i>Oryza sativa, Raphanus sativus, Glycine max, Cucumis sativus</i>	S1	Cold-inducible transcription; heterodimerization (LIP19-OBF1); ROS scavenging; proline biosynthesis
	Heat	<i>AtbZIP60, OsbZIP74, BhbZIP60, TabZIP74</i>	<i>Arabidopsis thaliana, Oryza sativa, Boea hygrometrica, Triticum aestivum</i>	K	ER stress-induced splicing; activation of UPR genes; protein folding; ROS homeostasis
	Heavy metal	<i>AtbZIP19, AtbZIP23, OsbZIP48, ThbZIP2</i>	<i>Arabidopsis thaliana, Oryza sativa, Tamarix hispida</i>	F / A	Regulation of zinc homeostasis and metal chelation; activation of antioxidant and phytochelatin genes to alleviate metal-induced oxidative stress
Biotic stress	Pathogen defense	<i>TGA2/5/6, OsbZIP1, TabZIP74</i>	<i>Arabidopsis thaliana, Oryza sativa, Triticum aestivum</i>	D	SA-dependent systemic acquired resistance (SAR); NPR1 interaction; PR gene activation
	Herbivory defense	<i>AtbZIP10, AtbZIP11, TGA2/5/6</i>	<i>Arabidopsis thaliana</i>	D/S	Integration of JA/ET signaling; modulation of ROS balance; regulation of phenylpropanoid; amino acid metabolism

Plants are constantly exposed to diverse environmental challenges, including abiotic stresses such as drought, salinity, temperature extremes, and heavy metal toxicity, as well as biotic stresses from pathogens and herbivores (Table 1). To cope, they have evolved sophisticated regulatory networks that integrate hormonal signaling, transcriptional reprogramming, and metabolic adjustments. The central mechanisms involve ABA- and JA-mediated cascades, activation of ROS scavenging systems, and induction of protective metabolites such as proline, flavonoids, and lignin. Within these networks, bZIP TFs act as pivotal hubs, linking upstream signal perception to downstream gene expression, thereby regulating osmotic balance, antioxidant defense, protein folding, and immune responses. Despite significant advances, several challenges remain. Many studies focus on single-gene characterizations, leaving the questions about functional redundancy and cross-talk among bZIP subgroups unresolved^[11]. Moreover, most findings are based on model species such as *Arabidopsis thaliana* and rice, whereas crop-specific roles and ecological contexts are less explored^[35]. Another critical gap is the limited understanding of spatiotemporal regulation, including how bZIPs integrate developmental cues with stress signaling in distinct tissues or stages^[33].

bZIP TFs hold great potential for molecular breeding and genome editing strategies aimed at developing stress-resilient crops. For example, manipulation of bZIP genes via clustered regularly interspaced short palindromic repeats (CRISPR) and CRISPR-associated protein 9 (Cas) mediation could enhance drought or salinity tolerance without compromising yield^[60]. In addition, synthetic biology approaches could exploit bZIP-controlled promoter elements to fine-tune stress-responsive genes' expression in transgenic lines. Future research should prioritize integrated multi-omics (transcriptomics, proteomics, metabolomics, and epigenomics) to construct holistic regulatory models. Comparative studies across species will be essential to identify conserved versus species-specific regulatory nodes. Finally, an important frontier lies in exploring how bZIPs participate in stress memory and priming, which may provide novel strategies for sustainable agriculture under climate change.

Functions of the bZIP family in plant metabolism

In plants, metabolism consists of primary and secondary metabolism. Primary metabolism supports growth and development,

producing essential compounds like sugars and amino acids. Secondary metabolism, derived from the primary pathways, generates products such as alkaloids and pigments, which are often associated with stress resistance. bZIP TFs are closely linked to plant metabolism (Fig. 3).

bZIPs involved in primary metabolism

The S1-group bZIP family (including *AtbZIP1, AtbZIP11, AtbZIP44, AtbZIP53*, and *bZIP63*) plays a regulatory role in glucose metabolism and sugar signaling pathways^[33]. *AtbZIP1*, a sugar-responsive gene, mediates sugar signaling and influences gene expression, plant growth, and development^[61]. Previous studies have shown that overexpression of the *tbz17* and *S1bZIP1 micro Open Reading Frame* (mORF) upregulates genes encoding sucrose phosphate synthase (SPS) and sucrose phosphate phosphatase (SPP), while silencing *tbz17* down-regulates these genes^[62]. The C/S1 network responds to low energy or nutritional starvation signals, which are mediated by SnRK1, HXK1 (HEXOKINASE 1), and the sucrose-induced repression of translation (SIRT) mechanism^[11]. For example, *bZIP63* is regulated by SnRK-mediated phosphorylation, which affects the target genes' expression and primary metabolism. A *bZIP63* knock-out mutant exhibited starvation-related phenotypes^[63]. *bZIP1* and *bZIP53* reprogram primary carbon and nitrogen metabolism after salt treatment, influencing gluconeogenesis and amino acid catabolism^[64].

Despite these advances, the regulatory mechanisms of S1-bZIPs in primary metabolism remain only partially understood. The molecular determinants of target gene specificity are not well defined, as *AtbZIP1* and *AtbZIP53* regulate overlapping pathways but display distinct outputs^[64]. It is also unclear how S1-bZIP-mediated sugar signaling is integrated with hormone pathways such as those associated with ABA and gibberellic acid (GA), leaving gaps in our understanding of stress adaptation^[11]. Furthermore, although knockout studies such as *bZIP63* suggest starvation phenotypes, it remains unresolved whether these reflect functional redundancy or unique contributions within the S1-group^[33,63]. Future work should combine structural biology, transcriptomics, and mutant analyses to clarify how S1-bZIPs balance energy status with growth.

In summary, S1-group bZIPs serve as central hubs linking sugar signaling with nutrient and stress responses, coordinating plant growth with metabolic homeostasis under fluctuating environmental conditions.

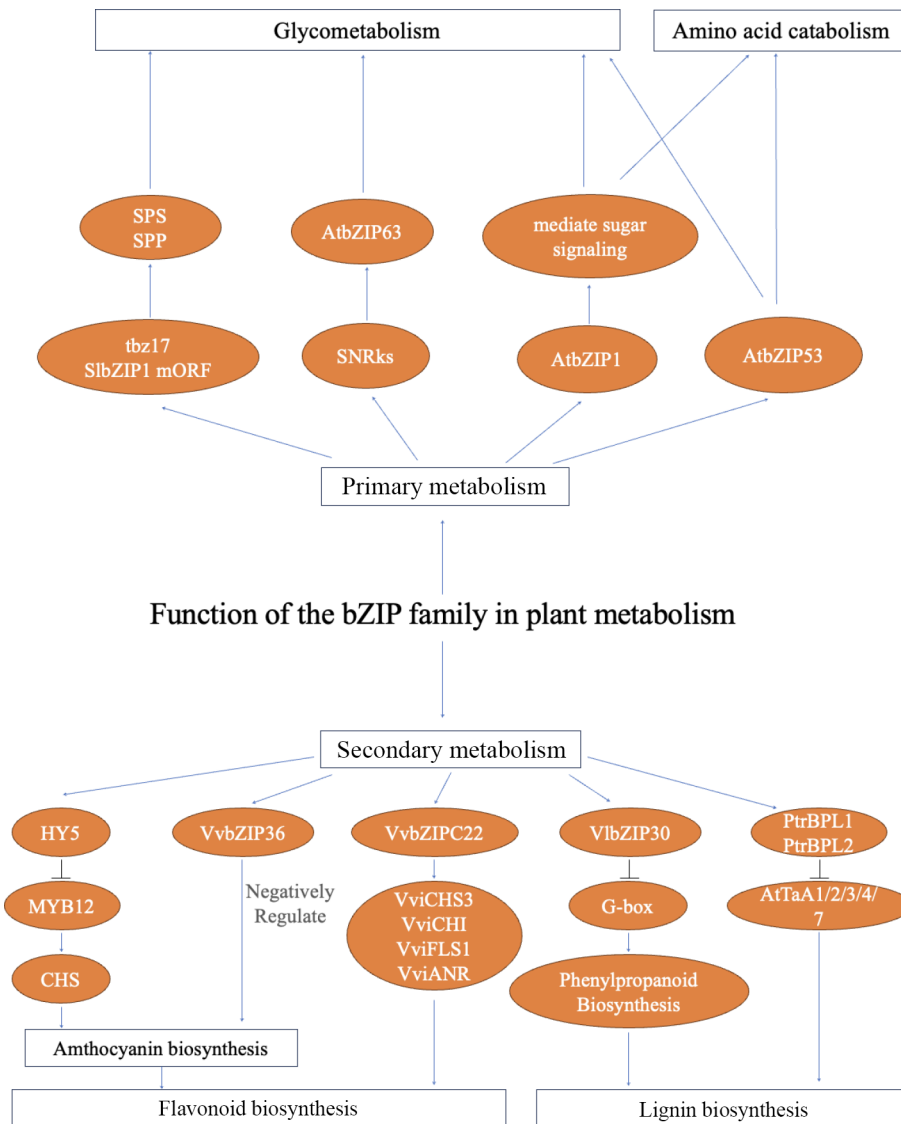


Fig. 3 The role of bZIP TFs in plant metabolism.

bZIPs involved in secondary metabolism

Flavonoids, a class of secondary metabolites, are widely distributed in plant organs and serve various biological functions^[65]. The MYB, basic helix–loop–helix (bHLH) and WD40 TF families are known to regulate flavonoid metabolism^[65]. The bZIP family of TFs also contributes to this regulatory network. *HY5* was the first bZIP TF identified as a regulator of anthocyanin biosynthesis. Experiments by Stracke et al. showed that *HY5* is required for light- and ultraviolet (UV)-B-responsive gene regulation and activates the flavonoid biosynthetic gene *CHS*. Moreover, *HY5* cooperates with *MYB12* to ensure appropriate regulation of flavonoid biosynthesis. Additionally, in grape, *VvbZIPC22* activates the transcription of flavonoid biosynthetic genes, including *VviCHS3*, *VviCHI*, *VviFLS1*, and *VviANR*, thereby contributing to flavanol production^[66]. Soybean *GmbZIP131* has been identified as a positive regulator of isoflavonoid synthesis, linking stress adaptation with legume-specific secondary metabolites^[67]. In contrast, Tu et al. (2022) reported that knockout of one allele of *VvbZIP36* negatively affects anthocyanin biosynthesis, suggesting that it functions as a negative regulator^[68].

Lignin forms the structural matrix of secondary cell walls in vascular plants and plays key physiological roles, including maintaining the xylem's hydrodynamic conductance, providing mechanical

strength, and facilitating defense against biotic and/or abiotic stressors^[69,70]. In addition to its role in ABA-mediated drought tolerance, *VlbZIP30* also directly regulates lignin biosynthesis. Tu et al. (2020) demonstrated that *VlbZIP30* binds to the promoters of phenylpropanoid pathway genes and activates peroxidase genes involved in lignin polymerization, thereby promoting strengthening of the cell wall under drought stress. This dual regulation of both ABA signaling and lignin metabolism highlights the central role of *VlbZIP30* in coordinating physiological and structural responses to dehydration^[71]. In poplar (*Populus trichocarpa*), two bZIP TFs (*PtrBPL1* and *PtrBPL2*) physically interact with *Arabidopsis* TGA proteins (*AtTGA1/3/4/7*), which are also members of the bZIP family, to synergistically activate lignin biosynthetic genes and promote lignin deposition^[72,73].

In addition, bZIPs can also regulate the biosynthesis of other secondary metabolites. *LvbZIP44* enhances anthocyanin accumulation in lily (*Lilium lancifolium* Thunb) petals under light induction^[74]. *SIAREB1* in tomato integrates ABA signaling with secondary metabolism, improving both drought resistance and carotenoid biosynthesis^[75]. In grapevine, *VvHY5* not only activates *CHS* but also coordinates UV-B-induced stilbene accumulation^[76]. In potato (*Solanum tuberosum*), *StHY5* plays a key role in light-mediated

regulation of anthocyanin biosynthesis, highlighting the broad and conserved functions of bZIPs in secondary metabolism across species^[77].

Although *an*-, *VvibZIP22*, *VvbZIP36*, and *VlbZIP30* have been identified as regulators of flavonoid and lignin biosynthesis, their hierarchical relationships remain poorly defined. Both *VvHY5* and *VvibZIP22* activate flavonoid genes, yet whether they act redundantly or cooperatively is unclear^[68,76]. New players, including *PopbZIP2*, *LvbZIP44*, *SIAREB1*, *VvHY5*, *GmbZIP131*, and *StHY5*, further complicate the network, but the mechanistic interactions and hormone cross-talk (ABA, JA, SA) remain insufficiently explored^[67,71,77,78]. These gaps underscore the need for integrative studies that dissect cooperative versus antagonistic functions of bZIPs and their links to stress-induced metabolic remodeling.

Overall, bZIP TFs function as versatile regulators in secondary metabolism, controlling the biosynthesis of flavonoids, lignin, and other phenylpropanoid-derived compounds. Their activities connect developmental cues and environmental stimuli with metabolic outputs, ultimately shaping plants' phenotype and stress adaptability.

bZIP transcription factors play multifaceted roles in both primary and secondary metabolism, linking energy homeostasis with stress adaptation and developmental processes. In primary metabolism, members of the S1-group of bZIPs such as *AtbZIP1*, *AtbZIP53*, and *bZIP63* act as central regulators of sugar signaling and nutrient reprogramming, coordinating carbon and nitrogen metabolism under fluctuating environmental conditions. In secondary metabolism, bZIPs including *HY5*, *VvibZIP22*, *VvbZIP36*, and *VlbZIP30* orchestrate flavonoid and lignin biosynthesis. Importantly, bZIPs can also regulate the biosynthesis of other secondary metabolites, such as alkaloids, terpenoids, and phenolic compounds, thereby expanding their functional scope beyond the flavonoid and lignin pathways. Despite these advances, the molecular mechanisms underlying their target gene specificity, regulatory hierarchies, and cross-talk with hormonal signaling remain poorly understood. Future research should integrate transcriptomics, metabolomics, and gene editing approaches to dissect the cooperative and antagonistic functions of bZIPs in metabolic regulation, ultimately providing new strategies for improving crops' quality and stress resilience.

bZIPs in plants' development and growth

Seed development

bZIP TFs play central roles in regulating seed maturation, accumulation of storage reserves, and germination. For example, *bZIP53* forms heterodimers with *bZIP10* and *bZIP25* to activate genes encoding seed storage proteins and proteins that are abundant in late embryogenesis, thereby coordinating maturation processes^[79]. In the ABA signaling pathway, *ABI5*, a Group A bZIP, acts as a key regulator of seed dormancy and germination by controlling ABA-responsive genes' expression and feedback regulation of ABA receptor genes^[80]. Moreover, seed-specific bZIPs such as *DPBF2* modulate oil biosynthesis and fatty acid composition, linking transcriptional control to seeds' nutritional quality^[81].

Despite these advances, current research on seed-related bZIPs is still fragmented. Most studies have focused on individual members like *ABI5* or *bZIP53*, but systematic dissection of the redundancy and specialization within the bZIP network is lacking. Furthermore, although heterodimerization is known to expand regulatory capacity, the precise "dimer code" in seed-related transcriptional regulation remains poorly understood^[11]. Bridging these knowledge gaps through cross-species comparative genomics, single-cell seed transcriptomics, and genome editing approaches will be critical for exploiting bZIPs for improving seed traits.

Light signaling and photomorphogenesis

Light is not only the energy source for plant photosynthesis but also an essential environmental signal that influences various aspects of plants' growth and development. Upon receiving light signals, plants' photoreceptors initiate a stepwise signal transduction process that activates key downstream TFs to regulate genes involved in development and photomorphogenesis.

HYH and *HY5*, two bZIP TFs, play central roles in light signaling and photomorphogenesis. *HY5* was the first TF identified in the regulation of photomorphogenesis^[82]. It binds to G-box and Z-box *cis*-elements in light-responsive promoters, inducing the expression of over 3,000 genes^[83]. In light conditions, *HY5* binds to *cis*-elements of photocontrol genes such as *CHS*, *CAB*, *PHYA*, and *RBCS*, thereby positively regulating their expression^[84–87].

As a TF, *HY5* functions by interacting with other proteins. CONSTITUTIVELY PHOTOMORPHOGENIC 1 (COP1)-*HY5* molecular modeling represents a central mechanism regulating photomorphogenesis in plants. COP1, a negative regulator of photomorphogenesis, directly interacts with *HY5* to mediate its proteasome-dependent degradation in the dark^[88]. UV-B light inhibits COP1-mediated *HY5* degradation, thereby promoting photomorphogenesis^[89]. Additionally, *HY5* interacts with and negatively regulates HFR1 and LAF1, which are crucial for Phytochrome A (phyA)-mediated photomorphogenesis^[90]. *HY5* also targets other light signaling components, including FAR-RED ELONGATED HYPOCOTYL1 (FHY1), FHY1-LIKE (FHL), and the B-BOX protein BBX25^[90–92].

Floral development and pollen germination

Floral development occurs in three successive stages: Floral induction, floral primordial formation, and floral organ development.

FLOWERING LOCUS D (FD), a bZIP family member, is a central regulator of floral transition at the shoot meristem. FLOWERING LOCUS T (FT), a conserved flowering promoter acting downstream of multiple regulatory pathways^[93], is expressed in the cotyledons and leaf vasculature but requires the bZIP TF FD at the shoot apex. Together, FT and FD promote floral transition and development via APETALA1 (AP1). FT may also function as a long-distance flowering signal. *ABI5*, another bZIP TF involved in ABA-mediated seed germination, also influences floral transition in *Arabidopsis*. Overexpression of *ABI5* delays flowering by upregulating FLOWERING LOCUS C (FLC), whereas ectopic FLC expression reverses early flowering in *ABI5* mutants.

During the formation of floral primordia, the *Arabidopsis* TF PERIANTHIA (PAN), part of the TGA family, regulates the development of floral organs' primordia, as evidenced by the abnormal pentamerous arrangement of the outer three floral whorls in the *pan* mutant^[94].

TGA TFs of Subfamily D are also critical for floral organ development. For example, *TGA9* and *TGA10* have overlapping functions in this process. *Arabidopsis* plants with mutations in both *TGA9* and *TGA10* exhibit male sterility, confirming these TFs' essential role in floral development and their necessity in anther dehiscence^[95].

In addition, bZIP TFs play a critical role in the development and functional regulation of roots and leaves. Roots, which absorb water and nutrients, anchor the plant and store nutrients, form the foundation for plants' survival and growth. In *Arabidopsis*, *bZIP29* directly binds to and activates cell-cycle-regulatory genes (e.g., *CYCB1;2*) and inhibitory genes (e.g., *SMR4*), maintaining root apical meristem homeostasis by balancing cell proliferation and differentiation^[96]. In maize, *ZmbZIP4* positively regulates abiotic stress responses and modulates root development by influencing auxin signaling and cyclin-related genes^[23].

Fruit development and organ senescence

Fruit development and organ senescence are complex biological processes that require precise transcriptional regulation. Increasing evidence has demonstrated that bZIPs play a central role in coordinating hormonal, metabolic, and environmental cues to regulate these developmental events. In fleshy fruit crops, bZIPs are directly linked to the control of ripening-associated pigmentation. For instance, in strawberry (*Fragaria × ananassa*), *FaHY5*, a light-responsive bZIP protein, acts within the *FaCRY1–FaCOP1–FaHY5* regulatory module to activate anthocyanin biosynthesis and transporter genes, thereby promoting pigment accumulation and fruit coloration at the molecular level. Silencing *FaHY5* leads to impaired anthocyanin production and delayed ripening, although its overexpression enhances fruit pigmentation^[97]. This highlights the critical role of bZIPs in integrating light signals with secondary metabolism to fine-tune fruit quality.

In addition to fruit development, bZIP transcription factors are key regulators of organ senescence, a genetically programmed process that is essential for nutrient remobilization and survival. In *Arabidopsis*, ABA-activated ABF/AREB bZIPs directly induce the expression of chlorophyll catabolic genes and senescence-associated genes (SAGs), thereby promoting the degradation of chlorophyll and accelerating leaf yellowing^[98]. These findings underscore that bZIPs function as transcriptional activators connecting hormonal signals with age-dependent transcriptional programs. Furthermore, recent reviews have emphasized the hub-like role of bZIPs in coordinating ABA, light, and stress signals with developmental outcomes, making them critical players in both fruit maturation and senescence pathways^[99]. The molecular mechanisms revealed in strawberry and *Arabidopsis* exemplify how bZIP TFs act as central regulators, integrating environmental cues and endogenous signals to control developmental plasticity.

bZIPs activate metabolic pathways for pigment biosynthesis during fruit ripening, modulate hormone signaling to regulate maturation, and promote senescence programs by controlling the degradation of chlorophyll and SAG expression. These recent advances highlight their importance as molecular switches in balancing fruit quality traits and organ longevity, providing promising targets for genetic improvement of crops.

Overall, bZIP TFs orchestrate diverse aspects of plants' development, including seed maturation, photomorphogenesis, flowering transition, organ growth, fruit ripening, and senescence, by integrating hormonal, metabolic, and environmental cues into transcriptional programs^[79,83,100]. Through context-specific dimerization and interaction with signaling pathways such as ABA, light, and carbon metabolism, bZIPs act as crucial molecular hubs linking developmental processes to stress adaptation^[11]. However, most studies have concentrated on a few representative members such as *HY5*, *ABI5*, and *FD*, leaving the broader regulatory dimer code and network redundancy insufficiently understood^[93]. Future research should therefore focus on elucidating the spatiotemporal specificity of bZIP dimers, integrating single-cell and spatial omics to map the developmental networks, and clarify the cross-talk among hormone, light, sugar, and ER stress signaling during growth transitions. In addition, dissecting the post-translational modifications and chromatin remodeling complexes that fine-tune bZIP activity will help link developmental plasticity with resilience^[85]. Finally, the application of genome editing and synthetic biology offers promising opportunities to engineer bZIP pathways in a precise and organ-specific manner, thereby improving flowering control, fruit quality, and postharvest longevity in crops.

The activity and specificity of bZIP TFs are tightly regulated by post-translational modifications (PTMs) and epigenetic mechanisms.

Phosphorylation, ubiquitination, and acetylation dynamically modulate the stability, localization, and DNA-binding activity of bZIP proteins during development and stress adaptation. For instance, SnRK2- and Calcium-Dependent Protein Kinase (CDPK)-mediated phosphorylation enhances the transactivation capacity of AREB/ABF members under drought and ABA treatments, whereas casein kinase II-dependent phosphorylation of *HY5* affects its stability and photomorphogenic function^[13,85]. Conversely, ubiquitin-proteasome-mediated degradation, such as the COP1-dependent turnover of *HY5*, represents a crucial feedback mechanism linking light and temperature cues^[100].

In parallel, bZIPs' transcriptional networks are modulated by epigenetic modifications that shape chromatin's accessibility at target loci. Histone acetylation and methylation influence recruitment of bZIPs to promoter regions of stress-responsive and metabolic genes. The *SNL–HDA19* histone deacetylase complex antagonizes *HY5* activity, repressing light-induced gene expression and photomorphogenesis^[85]. Furthermore, interactions between bZIPs and chromatin remodelers such as Switch/Sucrose Non-Fermentable (SWI/SNF) complexes integrate hormonal and developmental signals to fine-tune gene expression. These multi-layered regulatory networks illustrate how PTMs and epigenetic modifications cooperate to orchestrate bZIP-mediated transcription in growth, stress responses, and secondary metabolism. Understanding these layers of control will be critical for elucidating the context-specific regulatory hierarchies and for leveraging bZIPs in precision engineering of crops.

The bZIP TF family, as highly conserved regulatory elements in plants, plays a central role in developmental regulation by participating in multiple physiological processes, including the initiation of seed maturation, establishment of the root system's architecture, light signal perception, and photomorphogenesis. Molecular studies have revealed that this family establishes complex signal cross-talk networks by integrating various hormone signaling pathways (e.g., ABA and JA) and secondary metabolic pathways (e.g., lignin biosynthesis in phenylpropanoid metabolism), thereby coordinating plants' adaptive responses to abiotic stresses such as drought and salinity and enhancing stress resistance. Although current molecular breeding strategies based on bZIP TFs have shown promise for developing stress-resistant germplasm, the functional redundancy among family members and subfamily-specific regulatory cascades remain to be fully understood. Future research should integrate multi-omics analyses with CRISPR/Cas9-mediated gene editing to systematically identify the key functional modules involved in improving crop nutrition and extreme environmental adaptation, with attention to the three-dimensional genomic architecture and epigenetic modifications. Therefore, this approach will support the development of multidimensional stress-resistance models, offering theoretical frameworks and technical strategies for breeding smart crops with complex stress-resistant traits.

Author contributions

The authors confirm their contributions to the paper as follows: writing – original draft: Mao Y, Tu M, Zhao R; writing – review and editing: Zhang Y. All authors reviewed the results and approved the final version of the manuscript.

Data availability

Data sharing is not applicable to this article, as no new datasets were generated or analyzed during the current study.

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Conflict of interest

The authors declare that they have no conflict of interest.

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References

- Dröge-Laser W, Snoek BL, Snel B, Weiste C. 2018. The *Arabidopsis* bZIP transcription factor family—an update. *Current Opinion in Plant Biology* 45:36–49
- Zhang B, Feng C, Chen L, Li B, Zhang X, et al. 2022. Identification and functional analysis of bZIP genes in cotton response to drought stress. *International Journal of Molecular Sciences* 23:14894
- Duan L, Mo Z, Fan Y, Li K, Yang M, et al. 2022. Genome-wide identification and expression analysis of the bZIP transcription factor family genes in response to abiotic stress in *Nicotiana tabacum* L. *BMC Genomics* 23:318
- Gill SS, Tuteja N. 2010. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry* 48:909–30
- Rolly NK, Imran QM, Shahid M, Imran M, Khan M, et al. 2020. Drought-induced *AtbZIP62* transcription factor regulates drought stress response in *Arabidopsis*. *Plant Physiology and Biochemistry* 156:384–95
- Liu J, Chu J, Ma C, Jiang Y, Ma Y, et al. 2019. Overexpression of an ABA-dependent grapevine bZIP transcription factor, *VvABF2*, enhances osmotic stress in *Arabidopsis*. *Plant Cell Reports* 38:587–96
- Liang C, Meng Z, Meng Z, Malik W, Yan R, et al. 2016. *GhABF2*, a bZIP transcription factor, confers drought and salinity tolerance in cotton (*Gossypium hirsutum* L.). *Scientific Reports* 6:35040
- Ji X, Liu G, Liu Y, Zheng L, Nie X, et al. 2013. The bZIP protein from *Tamarix hispida*, *ThbZIP1*, is ACGT elements binding factor that enhances abiotic stress signaling in transgenic *Arabidopsis*. *BMC Plant Biology* 13:151
- Li Q, Wu Q, Wang A, Lv B, Dong Q, et al. 2019. Tartary buckwheat transcription factor *FtbZIP83* improves the drought/salt tolerance of *Arabidopsis* via an ABA-mediated pathway. *Plant Physiology and Biochemistry* 144:312–23
- Li XY, Liu X, Yao Y, Li YH, Liu S, et al. 2013. Overexpression of *Arachis hypogaea* *AREB1* gene enhances drought tolerance by modulating ROS scavenging and maintaining endogenous ABA content. *International Journal of Molecular Sciences* 14:12827–42
- Dröge-Laser W, Weiste C. 2018. The C/S₁ bZIP network: a regulatory hub orchestrating plant energy homeostasis. *Trends in Plant Science* 23:422–33
- Lin Z, Li Y, Wang Y, Liu X, Ma L, et al. 2021. Initiation and amplification of SnRK2 activation in abscisic acid signaling. *Nature Communications* 12:2456
- Song J, Sun P, Kong W, Xie Z, Li C, et al. 2023. SnRK2.4-mediated phosphorylation of ABF2 regulates ARGININE DECARBOXYLASE expression and putrescine accumulation under drought stress. *New Phytologist* 238:216–36
- Wang J, Li Q, Mao X, Li A, Jing R. 2016. Wheat transcription factor TaAREB3 participates in drought and freezing tolerances in *Arabidopsis*. *International Journal of Biological Sciences* 12:257–69
- Xiang Y, Tang N, Du H, Ye H, Xiong L. 2008. Characterization of OsbZIP23 as a key player of the basic leucine zipper transcription factor family for conferring abscisic acid sensitivity and salinity and drought tolerance in rice. *Plant Physiology* 148:1938–52
- Lu G, Gao C, Zheng X, Han B. 2009. Identification of OsbZIP72 as a positive regulator of ABA response and drought tolerance in rice. *Planta* 229:605–15
- Hsieh TH, Li CW, Su RC, Cheng CP, Sanjaya, et al. 2010. A tomato bZIP transcription factor, SIAREB, is involved in water deficit and salt stress response. *Planta* 231:1459–73
- Tu M, Wang X, Zhu Y, Wang D, Zhang X, et al. 2018. *VibZIP30* of grapevine functions in dehydration tolerance via the abscisic acid core signaling pathway. *Horticulture Research* 5:49
- Li Q, Zhao H, Wang X, Kang J, Lv B, et al. 2020. Tartary buckwheat transcription factor *FtbZIP5*, regulated by FtSnRK2.6, can improve salt/drought resistance in transgenic *Arabidopsis*. *International Journal of Molecular Sciences* 21:1123
- Zong W, Tang N, Yang J, Peng L, Ma S, et al. 2016. Feedback regulation of ABA signaling and biosynthesis by a bZIP transcription factor targets drought-resistance-related genes. *Plant Physiology* 171:2810–25
- Joo H, Lim CW, Lee SC. 2019. Roles of pepper bZIP transcription factor CaATBZ1 and its interacting partner RING-type E3 ligase CaASRF1 in modulation of ABA signalling and drought tolerance. *Plant Journal* 100:399–410
- Lu C, Liu X, Tang Y, Fu Y, Zhang J, et al. 2024. A comprehensive review of TGA transcription factors in plant growth, stress responses, and beyond. *International Journal of Biological Macromolecules* 258:128880
- Ma H, Liu C, Li Z, Ran Q, Xie G, et al. 2018. *ZmbZIP4* contributes to stress resistance in maize by regulating ABA synthesis and root development. *Plant Physiology* 178:753–70
- Feng Y, Wang Y, Zhang G, Gan Z, Gao M, et al. 2021. Group-C/S1 bZIP heterodimers regulate *MdIPT5b* to negatively modulate drought tolerance in apple species. *Plant Journal* 107:399–417
- Zhang JY, Qu SC, Qiao YS, Zhang Z, Guo ZR. 2014. Overexpression of the *Malus hupehensis* *MhNPR1* gene increased tolerance to salt and osmotic stress in transgenic tobacco. *Molecular Biology Reports* 41:1553–61
- Gai WX, Ma X, Qiao YM, Shi BH, Haq SU, et al. 2020. Characterization of the bZIP transcription factor family in pepper (*Capsicum annuum* L.): *CabZIP25* positively modulates the salt tolerance. *Frontiers in Plant Science* 11:139
- Wang Y, Gao C, Liang Y, Wang C, Yang C, et al. 2010. A novel bZIP gene from *Tamarix hispida* mediates physiological responses to salt stress in tobacco plants. *Journal of Plant Physiology* 167:222–30
- Liu C, Mao B, Ou S, Wang W, Liu L, et al. 2014. *OsbZIP71*, a bZIP transcription factor, confers salinity and drought tolerance in rice. *Plant Molecular Biology* 84:19–36
- Ying S, Zhang DF, Fu J, Shi YS, Song YC, et al. 2012. Cloning and characterization of a maize bZIP transcription factor, *ZmbZIP72*, confers drought and salt tolerance in transgenic *Arabidopsis*. *Planta* 235:253–66
- Kang C, Zhai H, He S, Zhao N, Liu Q. 2019. A novel sweetpotato bZIP transcription factor gene, *IbbZIP1*, is involved in salt and drought tolerance in transgenic *Arabidopsis*. *Plant Cell Reports* 38:1373–82
- Rolly NK, Imran QM, Lee IJ, Yun BW. 2020. Salinity stress-mediated suppression of expression of salt overly sensitive signaling pathway genes suggests negative regulation by *AtbZIP62* transcription factor in *Arabidopsis thaliana*. *International Journal of Molecular Sciences* 21:1726
- Liu JX, Srivastava R, Howell SH. 2008. Stress-induced expression of an activated form of *AtbZIP17* provides protection from salt stress in *Arabidopsis*. *Plant, Cell & Environment* 31:1735–43
- Wang H, Zhang Y, Norris A, Jiang CZ. 2022. S1-bZIP transcription factors play important roles in the regulation of fruit quality and stress response. *Frontiers in Plant Science* 12:802802
- D'Angeli S, Matteucci M, Fattorini L, Gismondi A, Ludovici M, et al. 2016. *OeFAD8*, *OeLIP* and *OeOSM* expression and activity in cold-acclimation of *Olea europaea*, a perennial dicot without winter-dormancy. *Planta* 243:1279–96
- Liu C, Schläppi MR, Mao B, Wang W, Wang A, et al. 2019. The bZIP73 transcription factor controls rice cold tolerance at the reproductive stage. *Plant Biotechnology Journal* 17:1834–49

36. Abro AA, Qasim M, Abbas M, Muhammad N, Ali I, et al. 2025. Integrating physiological and molecular insights in cotton under cold stress conditions. *Genetic Resources and Crop Evolution* 72:2561–91
37. Liao Y, Zou HF, Wei W, Hao YJ, Tian AG, et al. 2008. Soybean GmbZIP44, GmbZIP62 and GmbZIP78 genes function as negative regulator of ABA signaling and confer salt and freezing tolerance in transgenic *Arabidopsis*. *Planta* 228:225–40
38. Zhang X, Wollenweber B, Jiang D, Liu F, Zhao J. 2008. Water deficits and heat shock effects on photosynthesis of a transgenic *Arabidopsis thaliana* constitutively expressing ABP9, a bZIP transcription factor. *Journal of Experimental Botany* 59:839–48
39. Wang B, Du H, Zhang Z, Xu W, Deng X. 2017. BhbZIP60 from resurrection plant *Boea hygrometrica* is an mRNA splicing-activated endoplasmic reticulum stress regulator involved in drought tolerance. *Frontiers in Plant Science* 8:245
40. Davoudi M, Chen J, Lou Q. 2022. Genome-wide identification and expression analysis of heat shock protein 70 (*HSP70*) gene family in pumpkin (*Cucurbita moschata*) rootstock under drought stress suggested the potential role of these chaperones in stress tolerance. *International Journal of Molecular Sciences* 23:1918
41. Lu SJ, Yang ZT, Sun L, Sun L, Song ZT, et al. 2012. Conservation of IRE1-regulated bZIP74 mRNA unconventional splicing in rice (*Oryza sativa* L.) involved in ER stress responses. *Molecular Plant* 5:504–14
42. Nagashima Y, Mishiba KI, Suzuki E, Shimada Y, Iwata Y, et al. 2011. *Arabidopsis* IRE1 catalyses unconventional splicing of bZIP60 mRNA to produce the active transcription factor. *Scientific Reports* 1:29
43. Li Z, Tang J, Srivastava R, Bassham DC, Howell SH. 2020. The transcription factor bZIP60 links the unfolded protein response to the heat stress response in maize. *The Plant Cell* 32:3559–75
44. Reis PAB, Carpinetti PA, Freitas PPJ, Santos EGD, Camargos LF, et al. 2016. Functional and regulatory conservation of the soybean ER stress-induced DCD/NRP-mediated cell death signaling in plants. *BMC Plant Biology* 16:156
45. Wang H, Niu H, Zhai Y, Lu M. 2017. Characterization of BiP genes from pepper (*Capsicum annuum* L.) and the role of *CaBiP1* in response to endoplasmic reticulum and multiple abiotic stresses. *Frontiers in Plant Science* 8:1122
46. Coutinho FS, Dos Santos DS, Lima LL, Vital CE, Santos LA, et al. 2019. Mechanism of the drought tolerance of a transgenic soybean overexpressing the molecular chaperone BiP. *Physiology and Molecular Biology of Plants* 25:457–72
47. Xiang Y, Sun X, Gao S, Qin F, Dai M. 2017. Deletion of an endoplasmic reticulum stress response element in a *ZmPP2C-A* gene facilitates drought tolerance of maize seedlings. *Molecular Plant* 10:456–69
48. Zhang H, Lu L. 2024. Transcription factors involved in plant responses to cadmium-induced oxidative stress. *Frontiers in Plant Science* 15:1397289
49. Lu Z, Yu M, Han X, Qiao G, Xu J, et al. 2024. *SpbZIP60* confers cadmium tolerance by strengthening the root cell wall compartmentalization in *Sedum plumbizincicola*. *Journal of Hazardous Materials* 480:135936
50. Lilay GH, Persson DP, Castro PH, Liao F, Alexander RD, et al. 2021. *Arabidopsis* bZIP19 and bZIP23 act as zinc sensors to control plant zinc status. *Nature Plants* 7:137–43
51. Spielmann J, Schloesser M, Hanikenne M. 2024. Reduced expression of bZIP19 and bZIP23 increases zinc and cadmium accumulation in *Arabidopsis halleri*. *Plant, Cell & Environment* 47:2093–108
52. Zavaliev R, Dong X. 2024. NPR1, a key immune regulator for plant survival under biotic and abiotic stresses. *Molecular Cell* 84:131–41
53. Zhang Y, Tessaro MJ, Lassner M, Li X. 2003. Knockout analysis of *Arabidopsis* transcription factors TGA2, TGA5, and TGA6 reveals their redundant and essential roles in systemic acquired resistance. *The Plant Cell* 15:2647–53
54. Wang F, Lin R, Li Y, Wang P, Feng J, et al. 2019. *TabZIP74* acts as a positive regulator in wheat stripe rust resistance and involves root development by mRNA splicing. *Frontiers in Plant Science* 10:1551
55. Meng XB, Zhao WS, Lin RM, Wang M, Peng YL. 2005. Identification of a novel rice bZIP-type transcription factor gene, *OsbZIP1*, involved in response to infection of *Magnaporthe grisea*. *Plant Molecular Biology Reporter* 23:301–2
56. Kaminaka H, Näke C, Eppele P, Dittgen J, Schütze K, et al. 2006. bZIP10-LSD1 antagonism modulates basal defense and cell death in *Arabidopsis* following infection. *EMBO Journal* 25:4400–11
57. Hanson J, Hanssen M, Wiese A, Hendriks MMWB, Smeekens S. 2008. The sucrose regulated transcription factor bZIP11 affects amino acid metabolism by regulating the expression of *ASPARAGINE SYNTHETASE1* and *PROLINE DEHYDROGENASE2*. *The Plant Journal* 53:935–49
58. Paudel J, Copley T, Amirizian A, Prado A, Bede JC. 2013. *Arabidopsis* redox status in response to caterpillar herbivory. *Frontiers in Plant Science* 4:113
59. Hickman R, Van Verk MC, Van Dijken AJH, Mendes MP, Vroegop-Vos IA, et al. 2017. Architecture and dynamics of the jasmonic acid gene regulatory network. *The Plant Cell* 29:2086–105
60. Kaushal C, Sachdev M, Parekh M, Gowrishankar H, Jain M, et al. 2025. Transcriptional engineering for value enhancement of oilseed crops: a forward perspective. *Frontiers in Genome Editing* 6:1488024
61. Kang SG, Price J, Lin PC, Hong JC, Jang JC. 2010. The *Arabidopsis* bZIP1 transcription factor is involved in sugar signaling, protein networking, and DNA binding. *Molecular Plant* 3:361–73
62. Thalor SK, Berberich T, Lee SS, Yang SH, Zhu X, et al. 2012. Deregulation of sucrose-controlled translation of a bZIP-type transcription factor results in sucrose accumulation in leaves. *PLoS One* 7:e33111
63. Mair A, Pedrotti L, Wurzing B, Anrather D, Simeunovic A, et al. 2015. SnRK1-triggered switch of bZIP63 dimerization mediates the low-energy response in plants. *elife* 4:e05828
64. Hartmann L, Pedrotti L, Weiste C, Fekete A, Schierstaedt J, et al. 2015. Crosstalk between two bZIP signaling pathways orchestrates salt-induced metabolic reprogramming in *Arabidopsis* roots. *The Plant Cell* 27:2244–60
65. Xu W, Dubos C, Lepiniec L. 2015. Transcriptional control of flavonoid biosynthesis by MYB-bHLH-WDR complexes. *Trends in Plant Science* 20:176–85
66. Malacarne G, Coller E, Czemplak S, Vrhovsek U, Engelen K, et al. 2016. The grapevine VvibZIP22 transcription factor is involved in the regulation of flavonoid biosynthesis. *Journal of Experimental Botany* 67:3509–22
67. Ni X, Wang Y, Dai L, Jiang K, Zeng S, et al. 2025. The transcription factor GmbZIP131 enhances soybean salt tolerance by regulating flavonoid biosynthesis. *Plant Physiology* 197:kiaf092
68. Tu M, Fang J, Zhao R, Liu X, Yin W, et al. 2022. CRISPR/Cas9-mediated mutagenesis of *VvbZIP36* promotes anthocyanin accumulation in grapevine (*Vitis vinifera*). *Horticulture Research* 9:uhac022
69. Tang W, Page M. 2013. Transcription factor *AtbZIP60* regulates expression of Ca²⁺-dependent protein kinase genes in transgenic cells. *Molecular Biology Reports* 40:2723–32
70. Liu Q, Luo L, Zheng L. 2018. Lignins: biosynthesis and biological functions in plants. *International Journal of Molecular Sciences* 19:335
71. Tu M, Wang X, Yin W, Wang Y, Li Y, et al. 2020. Grapevine *VlbZIP30* improves drought resistance by directly activating *VvNAC17* and promoting lignin biosynthesis through the regulation of three peroxidase genes. *Horticulture Research* 7:150
72. Wang Y, Salasini BC, Khan M, Devi B, Bush M, et al. 2019. Clade I TGACG-motif binding basic leucine zipper transcription factors mediate BLADE-ON-PETIOLE-dependent regulation of development. *Plant Physiology* 180:937–51
73. Li S, Devi B, Allam G, Bhullar A, Murmu J, et al. 2023. Regulation of secondary growth by poplar *BLADE-ON-PETIOLE* genes in *Arabidopsis*. *Frontiers in Plant Science* 14:1244583
74. Gao Z, Sun Y, Zhu Z, Ni N, Sun S, et al. 2024. Transcription factors LvBBX24 and LvbZIP44 coordinated anthocyanin accumulation in response to light in lily petals. *Horticulture Research* 11:uhae211
75. He Y, Wu Q, Cui C, Tian Q, Zhang D, et al. 2023. ChIP-seq analysis of *SIAREB1* downstream regulatory network during tomato ripening. *Foods* 12:2357
76. Ali Z, Sun Y, Ma Z, Zheng Y, Liu Y. 2025. *VvHY5* and *VvBEE1* antagonistically control resveratrol biosynthesis to mitigate high light-induced damage in grapevine. *Journal of Integrative Plant Biology* 67:993–1008
77. Zhang H, Liu J, Zhao Y, Wang E, Zhou J, et al. 2025. Comprehensive metabolomics and transcriptomics analysis reveal the regulatory

- mechanism of StHY5 on anthocyanin accumulation in potato tubers. *Plant Science* 356:112500
78. Zhao K, Chen S, Yao W, Cheng Z, Zhou B, et al. 2021. Genome-wide analysis and expression profile of the bZIP gene family in poplar. *BMC Plant Biology* 21:122
 79. Alonso R, Oñate-Sánchez L, Weltmeier F, Ehlert A, Diaz I, et al. 2009. A pivotal role of the basic leucine zipper transcription factor bZIP53 in the regulation of *Arabidopsis* seed maturation gene expression based on heterodimerization and protein complex formation. *The Plant Cell* 21:1747–61
 80. Ali F, Qanmber G, Li F, Wang Z. 2022. Updated role of ABA in seed maturation, dormancy, and germination. *Journal of Advanced Research* 35:199–214
 81. Kim I, Lee KR, Park ME, Kim HU. 2022. The seed-specific transcription factor DPBF2 modulates the fatty acid composition in seeds. *Plant Direct* 6:e395
 82. Zhang Y, Li C, Zhang J, Wang J, Yang J, et al. 2017. Dissection of HY5/HYH expression in *Arabidopsis* reveals a root-autonomous HY5-mediated photomorphogenic pathway. *PLoS One* 12:e0180449
 83. Lee J, He K, Stolc V, Lee H, Figueroa P, et al. 2007. Analysis of transcription factor HY5 genomic binding sites revealed its hierarchical role in light regulation of development. *The Plant Cell* 19:731–49
 84. Maxwell BB, Andersson CR, Poole DS, Kay SA, Chory J. 2003. HY5, Circadian Clock-Associated 1, and a cis-element, DET1 dark response element, mediate DET1 regulation of chlorophyll a/b-binding protein 2 expression. *Plant Physiology* 133:1565–77
 85. Jing Y, Guo Q, Lin R. 2021. The SNL-HDA19 histone deacetylase complex antagonizes HY5 activity to repress photomorphogenesis in *Arabidopsis*. *New Phytologist* 229:3221–36
 86. Chattopadhyay S, Ang LH, Puente P, Deng XW, Wei N. 1998. *Arabidopsis* bZIP protein HY5 directly interacts with light-responsive promoters in mediating light control of gene expression. *The Plant Cell* 10:673–83
 87. Huang Y, Xiong H, Xie Y, Lyu S, Miao T, et al. 2022. BBX24 interacts with DELLA to regulate UV-B-induced photomorphogenesis in *Arabidopsis thaliana*. *International Journal of Molecular Sciences* 23:7386
 88. Osterlund MT, Hardtke CS, Wei N, Deng XW. 2000. Targeted destabilization of HY5 during light-regulated development of *Arabidopsis*. *Nature* 405:462–66
 89. Jiang L, Wang Y, Li QF, Björn LO, He JX, et al. 2012. *Arabidopsis* STO/BBX24 negatively regulates UV-B signaling by interacting with COP1 and repressing HY5 transcriptional activity. *Cell Research* 22:1046–57
 90. Jang IC, Henriques R, Chua NH. 2013. Three transcription factors, HFR1, LAF1 and HY5, regulate largely independent signaling pathways downstream of phytochrome A. *Plant & Cell Physiology* 54:907–16
 91. Li J, Li G, Gao S, Martinez C, He G, et al. 2010. *Arabidopsis* transcription factor ELONGATED HYPOCOTYL5 plays a role in the feedback regulation of phytochrome a signaling. *The Plant Cell* 22:3634–49
 92. Gangappa SN, Crocco CD, Johansson H, Datta S, Hettiarachchi C, et al. 2013. The *Arabidopsis* B-BOX protein BBX25 interacts with HY5, negatively regulating BBX22 expression to suppress seedling photomorphogenesis. *The Plant Cell* 25:1243–57
 93. Collani S, Neumann M, Yant L, Schmid M. 2019. FT modulates genome-wide DNA-binding of the bZIP transcription factor FD. *Plant Physiology* 180:367–80
 94. Gutsche N, Zachgo S. 2016. The N-terminus of the floral *Arabidopsis* TGA transcription factor PERIANTHIA mediates redox-sensitive DNA-binding. *PLoS One* 11:e0153810
 95. Murmu J, Bush MJ, DeLong C, Li S, Xu M, et al. 2010. *Arabidopsis* basic leucine-zipper transcription factors TGA9 and TGA10 interact with floral glutaredoxins ROXY1 and ROXY2 and are redundantly required for anther development. *Plant Physiology* 154:1492–504
 96. Van Leene J, Blomme J, Kulkarni SR, Cannoot B, De Winne N, et al. 2016. Functional characterization of the *Arabidopsis* transcription factor bZIP29 reveals its role in leaf and root development. *Journal of Experimental Botany* 67:5825–40
 97. Liu Y, Tang L, Wang Y, Zhang L, Xu S, et al. 2023. The blue light signal transduction module FaCRY1-FaCOP1-FaHY5 regulates anthocyanin accumulation in cultivated strawberry. *Frontiers in Plant Science* 14:1144273
 98. Cao J, Liu H, Tan S, Li Z. 2023. Transcription factors-regulated leaf senescence: current knowledge, challenges and approaches. *International Journal of Molecular Sciences* 24:9245
 99. Guo Z, Dzinyela R, Yang L, Hwarari D. 2024. bZIP transcription factors: structure, modification, abiotic stress responses and application in plant improvement. *Plants* 13:2058
 100. Liu S, Wang Q, Zhong M, Lin G, Ye M, et al. 2025. The CRY1-COP1-HY5 axis mediates blue-light regulation of *Arabidopsis* thermotolerance. *Plant Communications* 6:101264



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