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The addition of spent oyster mushroom substrates has positive effects on alfalfa growth and soil available nutrients

Yuchuan Shi, Xinran Cui, Yingjun Zhang^{*} and Mohan Liu^{*}

College of Grassland Science and Technology, China Agricultural University, Beijing 100193, China * Corresponding authors, E-mail: zhangyj@cau.edu.cn; liumh@cau.edu.cn

Abstract

This study aimed to investigate the effects of spent mushroom substrate (SMS, 0%, 10%, 20%, and 30%, w/w) addition to degraded grassland soil on the growth and nutrient uptake of alfalfa through a greenhouse pot experiment. Meanwhile, we compared with the inorganic fertilizer (CF, 200 N mg/kg and 30 P mg/kg) application treatment, and explored the most suitable SMS addition amount for alfalfa yield. Our results showed, that compared with the control treatment (CK), 10% SMS, 20% SMS, 30% SMS, and CF treatments increased alfalfa shoot biomass by 1.19, 1.67, 1.77, and 1.77 times, respectively. Total carbon content in leaves and total nitrogen content in stems of 20% SMS treatment were significantly higher than other treatments. Adding SMS increased the nodule number, especially the 20% SMS treatment. In addition, the concentrations of dissolved organic carbon, available phosphorus, and microbial biomass carbon and nitrogen were significantly enhanced with increasing SMS addition, and there was no significant difference between CF and CK treatments except for available phosphorus. Shoot biomass was significantly correlated with available phosphorus. In summary, adding SMS (20% and 30%) to degraded grassland soil can significantly improve soil nutrients and microenvironment to increase alfalfa yield, but considering economic benefits, 20% SMS is the most suitable application amount. This study provides the theoretical basis and technical support for the large-scale application of SMS in the field.

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Introduction

According to the survey by the China Edible Fungi Association, China is the largest country in the production and export of edible mushrooms^[1,2]. Edible mushrooms are rich in minerals and are often considered an alternative source of meat, fish, and vegetables^[3,4]. In particular, the demand for Pleurotus mushrooms is increasing year by year because of its high protein, dietary fiber, as well as essential and non-essential amino acids^[5,6]. It has been known that each kilogram of mushroom can produce about 2.5–5 kg of the spent mushroom substrate (SMS)^[7,8]. Currently, SMS is usually treated as agricultural waste and is landfilled, burned in the open on land, or composted with animal manure. These disposals may lead to environmental issues such as soil, air, and water pollution^[9].

SMS is the lignocellulosic by-product of mushroom cultivation mainly referring to biomass waste which is not completely degraded as the degradation efficiency of edible fungi species only reached 40%–80%^[10,11]. The term 'spent mushroom compost' (SMC) can be used interchangeably with SMS describing the agro-residues and fungal mycelium left after mushroom harvest^[12]. SMS mainly consisted of residual fungal mycelium, various disintegrated lignocellulosic biomass (such as corn cob, sawdust, livestock litter and manure, cottonseed hull, wood chip, straw), amendments (such as lime, peat, gypsum), nutrients, as well as a high level of enzymes and organic matter^[13–15]. A previous study has found that SMS has the characteristics of loose texture, small bulk density, rich particle structure, good air permeability, high nutrient, and water retention rate, which can effectively improve soil physical structure and soil microbial ecological environment^[16].

After harvesting mushrooms, the SMS contains nutrients that are useful for crop, vegetable, fruit tree production and soil improvement^[17]. Nowadays, there have been various studies using SMS as a biofertilizer to cultivate plants, including tomato, barley, pineapple, and so on. Lopes et al.[18] concluded that the use of SMS (Agaricus subrufescens) for seedling production had a higher total tomato production compared to previously reported production levels. Some experiments have also shown that cultivating pineapple in soil amended with SMS of oyster mushroom (Pleurotus ostreatus) increased its fruit number and optimized soil properties^[19]. Field experiment results showed that the application of SMS at 50 and 100 t/ha significantly increased barley grain yield and soil nutrients, especially soil nitrogen and potassium content^[20]. Vahid Afagh et al.^[21] revealed that 10%–15% of SMS addition in the growing substrate significantly increased plant growth, flower yield, essential macronutrient uptake and soluble sugar content as well as essential oil percentages on German chamomile (Matricaria recursta L.) compared to the control treatment (no SMS). In addition, a previous report showed that SMS improved soil physical and chemical properties, and the speed of lawn establishment in turf production^[22]. However, research on using SMS to cultivate forage has not yet been in-depth. Improving the nutrient content of grasslands in grassland management is one of the important measures to promote forage production. Therefore, exploring the impact mechanism of SMS on forage in degraded grassland soil can help the editable mushroom

industry to achieve green development, transformation, and upgrading^[23].

Alfalfa has been grown on approximately 30 million hectares for hay, silage processing and grazing globally^[24]. With its long cultivation history and adaptability to a wide range of territories, alfalfa has been commonly used for animal feed^[25]. It has also been utilized as a medicinal herb since it is a good source of vitamins (A, C, E, and K), protein, and minerals such as calcium, iron, potassium, and phosphorus^[26]. Apart from these uses, alfalfa is an ideal natural resource and model plant for the remediation of contaminated soils, offering a variety of elite characteristics, including a highly productive biomass, drought tolerance, a fast-growing and prosperous root system, and availability in large amounts over several months of the year^[27]. Based on the current cultivation status and application prospects of alfalfa mentioned above, it is quite important to study the application of SMS in alfalfa cultivation. Therefore, this research is based on the idea that SMS is expected to be used for alfalfa cultivation, filling the gap in this research field. Using SMS and soil mixed in proportion as a nutrient substrate. This research aims to explore the effect of SMS on the growth of alfalfa in degraded grassland soil, while improving the reuse efficiency of mushroom substrate.

Materials and methods

Soil and spent mushroom substrate properties

The topsoil (0–20 cm) used in the experiment was taken from Duolun Restoration Ecology Research Station(42°20'N, 116°17'E), Inner Mongolian, China^[28]. Soil properties were: pH 6.51, 84,066.46 mg/kg available nitrogen, 48.55 mg/kg dissolved organic carbon (DOC), and 232.79 mg/kg microbial biomass carbon (MBC). The type of experimental soil was classified as Haplic calisols according to the Food and Agriculture Organization of the United Nation (FAO) classification.

The experimental spent mushroom substrate (SMS) was the waste residue from 4–5 generations of *Pleurotus ostreatus* from Beijing Academy of Agriculture and Forestry Sciences, and its main components include 50% cottonseed hull, 40% corncob, a small amount of bran, and soybean meal (about 2%–5%). The spent oyster mushroom substrates properties were: pH 6.04, 90,650.61 mg/kg available nitrogen, 50,426.34 mg/kg DOC, and 50,626.21 mg/kg MBC. The spent mushroom substrate was thoroughly air-dried, crushed, and mixed before use.

Experimental design

The experiment included four substrate ratio treatments and one inorganic fertilizer with four replications. Therefore, these five treatments were 100% soil (CK), 90% soil and 10% SMS spent oyster mushroom substrates (10%SMS), 80% soil and 20% SMS spent oyster mushroom substrates (20%SMS), 70% soil and 30% SMS spent oyster mushroom substrates (30% SMS), and 100% soil with adding inorganic fertilizers (CF). Mix the crushed SMS and soil passing through a 2 mm sieve in proportion to each other and put them into disinfected pots one by one. Each plastic pot was filled with 0.4 kg dry weight of mixed substrate. According to previous studies, 200 mg/kg N and 30 mg/kg P (the actual fertilizer substances were (NH₄)₂SO₄ and KH₂PO₄) were applied in the inorganic fertilizer treatment^[29,30].

Use a 10% H₂O₂ (SCRC, Shanghai, China) solution to sterilize the surface of alfalfa (*Medicago sativa* L.) seeds with uniform

size and full particles. Rinse them with sterile water several times and then sow them in plastic pots, with a few seeds per pot. Water them every two days. After the emergence of forage grass, eight plants per pot were established, cultivated in an artificial climate chamber with 16 h light, 8 h darkness, and a constant temperature of 25 °C. Under these conditions, alfalfa can have suitable growth conditions and good cultivation effects^[31]. The experiment was conducted from April 2022 to December 2022 and harvested 40 d later.

Experimental method

This experiment measured the plants and soil of five treatments of alfalfa, including plant physiological indicators such as shoot biomass, root biomass, alfalfa nodule number, and total carbon (TC) and nitrogen (TN) of plants (including roots, stems, and leaves respectively), as well as soil microbial biomass carbon (MBC), microbial biomass nitrogen (MBN), dissolved organic carbon (DOC), and ammonium nitrogen (NH_4^+ -N), nitrate nitrogen (NO_3^- -N), soil pH and available phosphorus (P).

We used scissors to cut aboveground alfalfa from the substrate surface, and then, the belowground was thoroughly washed with deionized water to remove soil particles. Shoots and roots were dried to constant weight at 65 °C and respectively weighed to measure the dry biomass of stems, leaves, and roots. Scan and calculate the number of alfalfa nodules using Epson Perfection V700 Photo (flat panel color image scanner) and WinRHIZO software. To measure TC and TN values of alfalfa components concluding roots, stems, and leaves by using Vario MACRO Cube Elemental.

We used the chloroform fumigation extraction method to determine MBC and MBN^[32]. Five grams of two soil subsamples and incubate the non-fumigated group in a sealed cardboard box for 24 h (with water and NaOH solution inside). The fumigated group is vacuum filtered for 24 h (with water, CCl₃, and NaOH solution inside) in a suction filter. After removal, add 20 ml of K₂SO₄ solution (0.5 M), shake and filter. Take 10 mL of the leaching solution and measure the data using an Analytik Jena Multi N/C3100 instrument to calculate the DOC, MBC, and MBN values.

Soil NH₄⁺-N and NO₃⁻-N were extracted from 10 g subsamples, added 50 mL KCl solution (1 M), shaken for 30 min, and then analyzed the filtrate with a continuous flow analyzer (AutoAnalyser 3, Analytical, Norderstedt, Germany)^[33]. 1:2.5 soil/water suspension was measured substrates pH using Sartorius PB-10.

We used the molybdate-ascorbic acid method for the colorimetric measurement of available phosphorus^[30]. Specifically, a 2.5 g air-dried substrate sample was added to 25 mL 0.5 mol/L NaHCO₃ solution and about 1 g dephosphorized activated carbon, shaken at 20–25 °C for 30 min, and the suspension was filtered by double filter paper. Add 2.5 mL of molybdenumantimony anti-mixed color reagent to 5 mL soil suspension, constant volume to 25 mL, shake and place at room temperature for 30 min. Available phosphorus concentration was determined using a multimode microplate reader (Varioskan LUX; Thermo Scientific, USA) at 880 nm wavelength.

Data analysis

Alfalfa growth data and soil indicator data were analyzed using one-way analysis of variance (ANOVA) by Duncan's multiple rang test and SPSS 20.0 software for processing; the significance level was p < 0.05. Before analysis, the Shapiro-Wilk test

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was used to evaluate the normality of the data. Levene test was used for analysis of variance to determine homogeneity of variance. Where necessary, the parameters were square root transformed to achieve normality and homogeneity. To explore the relationship between variables, the redundancy analysis (RDA) and Pearson correlation analysis were used for evaluation. All the figures were performed in R 4.2.1.

Results

Plant biomass and nutrient uptake

The shoot and root biomass of alfalfa cultured with 20% SMS, 30% SMS and CF treatments were significantly higher than those of CK treatment (Fig. 1, p < 0.05), but there was no significant difference between 10% SMS and CK treatments (p > 0.05). And there was no significant difference between the shoot and root biomass of 20% SMS and 30% SMS treatments compared with CF treatment (p < 0.05). CF treatment had the largest number of root nodules number, followed by 20% SMS, 30% SMS, and 10% SMS treatments, and CK treatment had the least number (Fig. 2). The number of nodules with 20% SMS and 30% SMS treatments was higher than CK treatment (Fig. 2, p < 0.05).

TN and TC content of forage grass reflects its excellent quality. Our data showed that 20% SMS treatment had the highest TN and TC contents of alfalfa leaf (Table 1). The TN and TC contents of alfalfa root and stem were the highest with CF treatment, and there was no significant difference compared with 20% SMS and 30% SMS treatments (p > 0.05).

Substrate physicochemical properties

The concentrations of DOC and available P significantly increased with the increase of SMS addition in the substrate (Fig. 3a & b, p < 0.05), and the concentrations were the lowest in the CK treatment. There was no significant difference between the 10%SMS and CF treatments (Fig. 3b, p > 0.05).

The MBN and MBC concentrations of 10% SMS, 20% SMS, and 30% SMS treatments were significantly higher than those of CF and CK treatments (Fig. 3c, p < 0.05). There was no difference between CF and CK treatments (p > 0.05).

 NO_3^- concentrations of 10% SMS, 20% SMS, and 30% SMS treatments were significantly lower than that of CK and CF treatments (Fig. 3e, p < 0.05). NH_4^+ concentration was the lowest in 20% SMS treatment, while other treatments had no significant difference with CK treatment (Fig. 3d, p < 0.05). There was no significant difference in soil pH among all treatments (Table 2, p > 0.05).

Correlation analysis

RDA was used to assess how the physicochemical properties of the substrates influenced alfalfa growth indicators. It can be seen that there are significant differences in the substrate

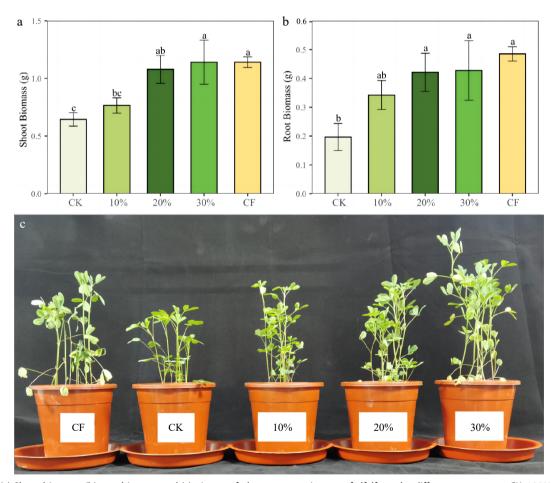


Fig. 1 (a) Shoot biomass, (b) root biomass and (c) picture of plant pot experiments of alfalfa under different treatments. CK: 100% soil; 10%: 10% SMS; 20%: 20% SMS; 30%: 30% SMS; CF: 100% soil with chemical fertilizers added. Different lowercase letters above the columns represent significant differences among these treatments according to Duncan tests.

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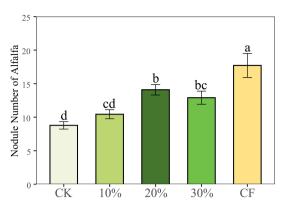


Fig. 2 Number of alfalfa nodules under different treatments. CK: 100% soil; 10%: 10% SMS; 20%: 20% SMS; 30%: 30% SMS; CF: 100% soil with chemical fertilizers added. Different lowercase letters above the columns represent significant differences among these treatments according to Duncan tests.

environment among different treatments, which have a significant impact on plant physiological indicators (Fig. 4). Plant physiological indicators respond strongly to the physicochemical characteristics of the substrate. The results show that the variance contribution rate of principal component 1 (PC1) and principal component 2 (PC2) respectively are 42.72% and 26.10%, and the cumulative variance contribution rate of PC1 and PC2 is 68.82%. Obtained through RDA diagram and calculation of Pearson correlation coefficient (PCC), available P had the strongest correlation with the PC1 and treatments distribution ($r^2 = 0.4035$, p = 0.008), followed by root nodule ($r^2 = 0.2829$, p = 0.047).

The Pearson correlation analysis showed that the shoot biomass value of alfalfa was significantly correlated with the available P in the substrate (Fig. 5, p < 0.05). MBC and MBN were strongly correlated with DOC and available P (p < 0.05). Meanwhile, there is a strong correlation between DOC and P (p < 0.001).

Discussion

SMS has been increasingly used as soil amendment either freshly or after composting processes in recent years^[34]. The current study found that adding appropriate amounts of SMS to degraded grassland soil can promote the growth and quality of alfalfa. The shoot biomass of alfalfa in 20% SMS and 30% SMS treatment significantly increased respectively by 1.67 and 1.77 times compared to CK treatment (Fig. 1). 20% SMS treatment had the highest TN contents of alfalfa stem and TC contents of alfalfa leaf (Table 1). This result supports previous studies, which illustrated that sometimes organic manure

Table 1. Total carbon, nitrogen content, and carbon and nitrogen ratio (TC, TN, C/N) of alfalfa harvested in different treatments.

Treatment	Root-TN (g)	Root-TC (g)	Root-C/N (%)	Stem-TN (g)	Stem-TC (g)	Stem-C/N (%)	Leaf-TN (g)	Leaf-TC (g)	Leaf-C/N (%)
СК	0.53 ± 0.26b	8.05 ± 3.69b	15.44 ± 0.86b	$0.27 \pm 0.11 b$	7.27 ± 2.73c	27.17 ± 1.27a	0.73 ± 0.49a	12.51 ± 8.20b	23.32 ± 12.55ab
10% SMS	$0.83 \pm 0.28 ab$	$14.13 \pm 4.18ab$	17.13 ± 1.09a	$0.28\pm0.09b$	8.49 ± 2.96bc	29.99 ± 3.39a	$0.98 \pm 0.47a$	$23.49 \pm 9.58ab$	24.8 ± 3.63a
20% SMS	$1.00 \pm 0.30 ab$	17.71 ± 5.83a	17.56 ± 0.79a	$0.52 \pm 0.16a$	14.58 ± 4.21ab	28.23 ± 2.71a	2.50 ± 2.16a	29.74 ± 7.79a	15.7 ± 6.43ab
30% SMS	$1.04 \pm 0.52 ab$	17.92 ± 8.80a	17.28 ± 0.57a	0.44 ± 0.19ab	13.16 ± 5.88abc	29.82 ± 2.14a	1.60 ± 1.09a	25.07 ± 17.19ab	11.76 ± 7.87b
CF	1.16 ± 0.15a	19.77 ± 2.30a	17.13 ± 0.45a	0.58 ± 0.13a	15.53 ± 2.61a	27.78 ± 6.58a	1.93 ± 0.21a	29.06 ± 1.85ab	15.14 ± 1.13ab

Different lowercase letters in the table represent significant differences among these treatments according to Duncan tests.

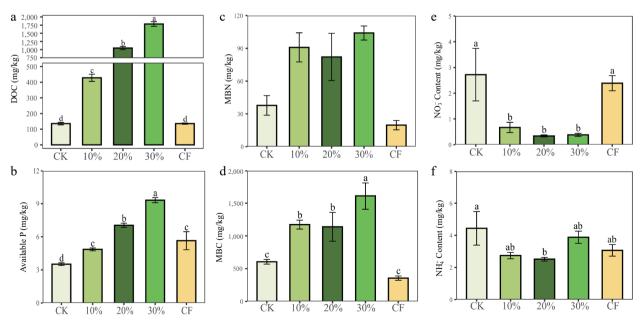


Fig. 3 Effects of different treatments on (a) soil dissolved organic carbon (DOC), (b) available P, (c) microbial biomass nitrogen (MBN), (d) microbial biomass carbon (MBC), (e) nitrate nitrogen (NO_3^-) and (f) ammonium nitrogen (NH_4^+) . Means ± S.E. CK: 100% soil; 10%: 10% SMS; 20%: 20% SMS; 30%: 30% SMS; CF: 100% soil with chemical fertilizers added. Different lowercase letters above the columns represent significant differences among these treatments according to Duncan tests.

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Table 2. The pH of substrate with different treatments after harvesting.

Experimental treatment	рН
СК	7.12 ± 0.18a
10% SMS	$7.08 \pm 0.09a$
20% SMS	$7.06 \pm 0.08a$
30% SMS	7.06 ± 0.15a
CF	7.10 ± 0.06a

Different lowercase letters in the table represent significant differences among these treatments according to Duncan tests.

amendment promoted plant growth more efficiently byproviding a higher nutrient value compared with inorganic fertilizer^[35].

We found that our SMS materials contain far more nutrients needed by plants than soil, which means that adding SMS to degraded grasslands with poor soil is beneficial for plant growth. As an important factor affecting soil quality, soil nutrients typically can maintain soil quality and promote plant productivity^[36]. During the growth process of *Pleurotus ostreatus*, due to the need for autotrophic nutrients, enzymes are secreted to mineralize the nutrients in the substrate, such as laccases, cellulases, hemicellulases, and xylanases^[37]. Some of these nutrients are absorbed by the mushrooms themselves, while others are left in the substrate, such as carbohydrates, protein, and fat^[38]. So, SMS contains different levels of carbon and nitrogen higher than soil, which can improve the storage capacity of soil nutrients when applied as substitutes for fertilizers^[39]. The results of our research concluded that the soil application of SMS improved soil fertility since the DOC and available P were increased significantly (Fig. 3). When applied as an alternative fertilizer, the nitrogen in SMS can be slowly released into the soil, which is beneficial for plants to assimilate nitrogen^[40]. And SMS is rich in phosphorus, which can be used as a phosphorus additive in soil to increase soil organic matter and nutrient content^[41,42]. Besides, the rich organic matter in SMS improves soil structure, soil aeration, and water retention, and even increases soil microbial activity^[43]. The increased organic C and available N in soils amended with organic manure would provide more organic resources that benefit the microbial activity in soils^[44]. Based on the previous research, four months after the addition of Agaricus bisporus SMS to the soil, organic N and available P content of the soil increased, while the impact on pH is not significant^[7], which is consistent with our experimental results. The effectiveness of SMS as an organic soil amendment has also been positively evaluated by Courtney & Mullen^[20]. The effect of applying SMS was equivalent to fertilization, where the C, N and P contents in the soil respectively increased by 40%, 28%, and 230%, meanwhile, the calcium, potassium, and magnesium content increased by three times.

In addition, SMS increased the beneficial microbial biomass in the substrate, which may be one of the reasons why SMS

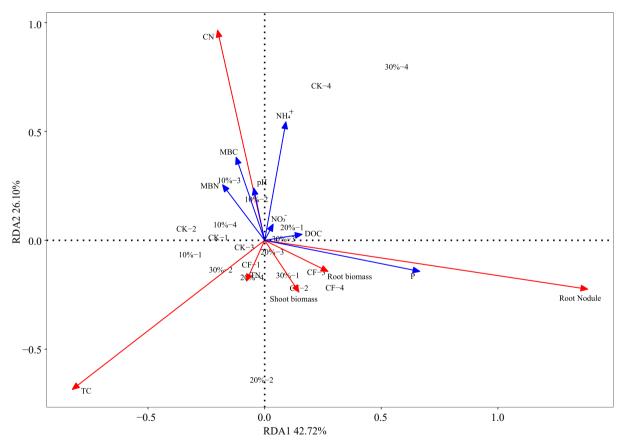


Fig. 4 The Redundancy Analysis (RDA) shows the effects of five experimental treatments on physical and chemical properties in substrate including pH, dissolved organic carbon (DOC), microbial biomass nitrogen (MBN), microbial biomass carbon (MBC), NH_4^+ and NO_3^- , available P (P) and plant indicators including shoot biomass, root biomass, plant total nitrogen (TN), total carbon (TC), and carbon nitrogen ratio (CN). The red arrow in the figure represents plant growth indicators, and the blue arrow in the figure represents physical and chemical properties in substrate.

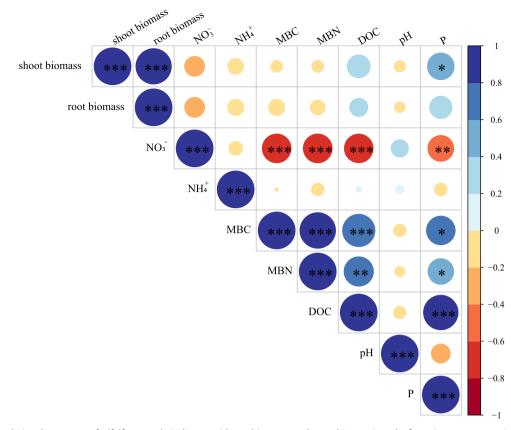


Fig. 5 Correlation heatmaps of alfalfa growth indicators (shoot biomass and root biomass) and of nutrient concentration indicators in substrates (NO_3^- , NH_4^+ , MBC, MBN, DOC, pH and P). NO_3^- : nitrate nitrogen; NH_4^+ : ammonium nitrogen; MBC: microbial biomass carbon; MBN: microbial biomass nitrogen; DOC: dissolved organic carbon; pH: acidity and alkalinity of substrates; P: available P. The color intensity in each panel indicates the relative correlation between read numbers of two groups. Blue represents positive correlation, and red represents negative correlation. Statistically significant correlations are indicated with *p < 0.05, **p < 0.01, and ***p < 0.001.

promotes plant growth. A previous study has found that Rhizobia is beneficial to leguminous plants and can effectively fix nitrogen in the air for plant use^[45]. Our data showed that the number of nodules increased with the increase of SMS addition, which can be further inferred to increase the effect of nitrogen fixation (Fig. 2). The mutually beneficial relationship between symbionts and host plants is established of based on an 'investment-return' strategy balance, which means that alfalfa uses rhizobia to obtain nutrients while correspondingly spending energy to maintain microbial life^[46]. Although the number of nodules in CF treatment was significantly higher than in other treatments (Fig. 2), there was no significant increase in biomass compared with 20% SMS and 30% SMS. It is possible because higher levels of nitrogen or phosphorus would break potential nutrient limitations, allowing host plants to easily obtain essential nutrients for growth through their roots, and plants would invest fewer resources in rhizobia at this point^[47]. The MBN and MBC concentrations of adding SMS treatments were significantly higher than no SMS treatments (CK and CF), which indicated that SMS addition promotes microbial biomass, while inorganic fertilizer addition does not have such effects (Fig. 3c & d). Research has shown that the different microbial populations of bacteria and fungi carried by SMS also can degrade organic foreign compounds in soil^[48], which may promote plant absorption of chemical fertilizers. Additional research instructed that microorganisms play an important role in soil fertility and nutrient cycling^[49]. Therefore,

SMS can promote the formation of root nodules, and increase the N content and the biomass of alfalfa.

Correlation analysis indicated that the soil available P concentration and the number of alfalfa nodules significantly affect the growth of alfalfa (Fig. 4). The shoot biomass of alfalfa was significantly correlated with the soil available P concentration, and MBC and MBN have a strong correlation with DOC content, and DOC is also related to P content (Fig. 5). Alfalfa has high requirements for phosphorus, sulfur, potassium, and some micronutrients, such as boron, and if one nutrient is deficient in soil, crop growth will be poor even if other nutrients are abundant^[50]. Therefore, it is inferred that the rich organic carbon and high microbial activity in the SMS improve nutrient cycling within the substrate, and the high P content in the substrate promotes an increase in alfalfa biomass.

Conclusions

The experimental results showed that the shoot and root biomass, DOC, available P, MBC, and MBN concentrations of alfalfa enhanced with the increase of SMS addition in degraded grassland soil. Among them, 20% SMS and 30% SMS treatments had no significant difference in alfalfa biomass compared with CF treatment. 20% SMS treatment had the second highest number of alfalfa nodules only after the CF treatment. Total carbon content in leaves and total nitrogen content in stems of 20% SMS treatment were significantly higher than

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other treatments. The available P was significantly correlated with shoot biomass, MBC, MBN, and DOC. In conclusion, the effects of 20% SMS treatment on alfalfa growth and soil improvement were superior to other treatments. Therefore, adding 20% SMS to the degraded grassland soil to cultivate alfalfa has a good economic and application prospect, which can improve the yield and quality of alfalfa, promote the recycling of waste and reduce the waste of resources. This study provides a theoretical basis and support for the large-scale application and popularization of this technology in the field.

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Conflict of interest

The authors declare that they have no conflicts of interest. Yingjun Zhang is the Editorial Board member of *Grass Research* who was blinded from reviewing or making decisions on the manuscript. The article was subject to the journal's standard procedures, with peer review handled independently of this Editorial Board member and his research groups.

Dates

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References

- 1. China Edible Fungi Association. 2022. Analysis of the results of the 2020 national edible fungi statistical survey. *Edible Fungi of China* 41:85–91
- 2. Jing Q. 2020. The current situation and development trend of mechanized production of edible mushrooms in China. *China Agricultural Machinery Safety Supervision* 224:23–24
- Kakon A, Choudhury BK, Saha S. 2012. Mushroom is an ideal food supplement. *Journal of Dhaka National Medical College & Hospital* 18:58–62
- 4. Ahmed M, Abdullah N, Nuruddin MM. 2016. Yield and nutritional composition of oyster mushrooms: an alternative nutritional source for rural people. *Sains Malaysiana* 45:1609–15
- 5. Zakia B, Srivastava H. 1962. Studies on cultivation of *Pleurotus* spp. on paddy straw. *Food Science* 11:363–65
- Singh MP, Singh VK. 2011. Yield performance and nutritional analysis of *Pleurotus citrinopileatus* on different agrowastes and vegetable wastes. *Proc. 7th International Conference on Mushroom Biology and Mushroom Products, Arcachon, France, 2011*, 1: 385–92. Villenave d'Ornon Cedex, France: Institut National de la Recherche Agronomique (INRA).
- Medina E, Paredes C, Bustamante MA, Moral R, Moreno-Caselles J. 2012. Relationships between soil physico-chemical, chemical and biological properties in a soil amended with spent mushroom substrate. *Geoderma* 173–174:152–61
- 8. Priadi D, Arfani A, Saskiawan I, Mulyaningsih ES. 2016. Use of grass and spent mushroom compost as a growing medium of local

tomato (Lycopersicon esculentum Miller) seedling in the nursery. Journal of Agricultural Science 38:242–50

- Jiang H, Zhang M, Chen J, Li S, Shao Y, et al. 2017. Characteristics of bio-oil produced by the pyrolysis of mixed oil shale semi-coke and spent mushroom substrate. *Fuel* 200:218–24
- Estrada AER, del Mar Jimenez-Gasco M, Royse DJ. 2009. Improvement of yield of *Pleurotus eryngii* var. *eryngii* by substrate supplementation and use of a casing overlay. *Bioresource Technology* 100:5270–76
- Mohd Hanafi FH, Rezania S, Mat Taib S, Md Din MF, Yamauchi M, et al. 2018. Environmentally sustainable applications of agro-based spent mushroom substrate (SMS): an overview. *Journal of Material Cycles and Waste Management* 20:1383–96
- Jordan SN, Mullen GJ, Murphy MC. 2008. Composition variability of spent mushroom compost in Ireland. *Bioresource Technology* 99:411–18
- Najafi B, Faizollahzadeh Ardabili S, Shamshirband S, Chau KW. 2019. Spent mushroom compost (SMC) as a source for biogas production in Iran. *Engineering Applications of Computational Fluid Mechanics* 13:967–82
- Moon YH, Shin PG, Cho SJ. 2012. Feeding value of spent mushroom (*Pleurotus eryngii*) substrate. *Journal of Mushroom Science* and Production 10:236–43
- Gobbi V, Nicoletto C, Zanin G, Sambo P. 2018. Specific humus systems from mushrooms culture. *Applied Soil Ecology* 123:709–13
- Lou Z, Sun Y, Bian S, Baig SA, Hu B, et al. 2017. Nutrient conservation during spent mushroom compost application using spent mushroom substrate derived biochar. *Chemosphere* 169:23–31
- Rinker DL. 2002. Handling and using 'spent' mushroom substrate around the world. Proc. 4th International Conference on Mushroom Biology and Mushroom Products, Cuernavaca, 2002, 43–60.
- Lopes RX, Zied DC, Martos ET, de Souza RJ, da Silva R, et al. 2015. Application of spent *Agaricus subrufescens* compost in integrated production of seedlings and plants of tomato. *International Journal of Recycling of Organic Waste in Agriculture* 4:211–18
- Adedokun O, Orluchukwu J. 2013. Pineapple: organic production on soil amended with spent mushroom substrate. *Agriculture and Biology Journal of North America* 4:590–93
- Courtney RG, Mullen GJ. 2008. Soil quality and barley growth as influenced by the land application of two compost types. *Bioresource Technology* 99:2913–18
- Vahid Afagh H, Saadatmand S, Riahi H, Khavari-Nejad RA. 2019. Influence of spent mushroom compost (SMC) as an organic fertilizer on nutrient, growth, yield, and essential oil composition of German chamomile (*Matricaria recutita* L.). *Communications in Soil Science and Plant Analysis* 50:538–48
- 22. Landschoot P, McNitt A. 1995. Using spent mushroom substrate (mushroom soil) as a soil amendment to improve turf. Thesis. USA: Pennsylvania State University.
- 23. Leong YK, Ma TW, Chang JS, Yang FC. 2022. Recent advances and future directions on the valorization of spent mushroom substrate (SMS): a review. *Bioresource Technology* 344:126157
- Hawkins C, Yu LX. 2018. Recent progress in alfalfa (Medicago sativa L.) genomics and genomic selection. The Crop Journal 6:565–75
- Biazzi E, Nazzicari N, Pecetti L, Annicchiarico P. 2019. GBS-based genome-wide association and genomic selection for alfalfa (*Medicago sativa*) forage quality improvement. *The Model Legume Medicago truncatula*923–27
- 26. Tussipkan D, Manabayeva SA. 2022. Alfalfa (*Medicago sativa* L.): genotypic diversity and transgenic alfalfa for phytoremediation. *Frontiers in Environmental Science* 10:828257
- 27. Ren H, Wan Y, Zhao Y. 2018. Phytoremediation of polychlorinated biphenyl-contaminated soil by transgenic alfalfa associated bioemulsifier AlnA. In *Twenty Years of Research and Development* on Soil Pollution and Remediation in China, eds Luo Y, Tu C. Singapore: Springer Singapore. pp. 645–53. https://doi.org/10.1007/978-981-10-6029-8_39

- 28. Yang G, Liu N, Lu W, Wang S, Kan H, et al. 2014. The interaction between arbuscular mycorrhizal fungi and soil phosphorus availability influences plant community productivity and ecosystem stability. Journal of Ecology 102:1072-82
- 29. Xiao Y, Liu M, Chen L, Ji L, Zhao Z, et al. 2020. Growth and elemental uptake of Trifolium repens in response to biochar addition, arbuscular mycorrhizal fungi and phosphorus fertilizer applications in low-Cd-polluted soils. *Environmental Pollution* 260:113761
- 30. Murphy J, Riley JP. 1962. A modified single solution method for the determination of phosphate in natural waters. Analytica Chimica Acta 27:31-36
- 31. Liu L, Bao A, Li H, Bai W, Liu H, et al. 2023. Overexpression of ZxABCG11 from Zygophyllum xanthoxylum enhances tolerance to drought and heat in alfalfa by increasing cuticular wax deposition. The Crop Journal 11:1140-51
- 32. Collins CG, Carey CJ, Aronson EL, Kopp CW, Diez JM. 2016. Direct and indirect effects of native range expansion on soil microbial community structure and function. Journal of Ecology 104:1271-83
- 33. Yang C, Zhang F, Liu N, Hu J, Zhang Y. 2018. Changes in soil bacterial communities in response to the fairy ring fungus Agaricus gennadii in the temperate steppes of China. Pedobiologia 69:34-40
- 34. Marín-Benito JM, Sánchez-Martín MJ, Rodríguez-Cruz MS. 2016. Impact of spent mushroom substrates on the fate of pesticides in soil, and their use for preventing and/or controlling soil and water contamination: a review. Toxics 4:17
- 35. Álvarez-López V, Prieto-Fernández Á, Cabello-Conejo MI, Kidd PS. 2016. Organic amendments for improving biomass production and metal yield of Ni-hyperaccumulating plants. Science of The Total Environment 548–549:370–79
- 36. Oladele SO. 2019. Changes in physicochemical properties and quality index of an Alfisol after three years of rice husk biochar amendment in rainfed rice - Maize cropping sequence. Geoderma 353.359-71
- 37. Cohen R, Persky L, Hadar Y. 2002. Biotechnological applications and potential of wood-degrading mushrooms of the genus Pleurotus. Applied Microbiology and Biotechnology 58:582-94
- 38. Luo W, Yuan J, Luo Y, Li G, Nghiem LD, et al. 2014. Effects of mixing and covering with mature compost on gaseous emissions during composting. Chemosphere 117:14-19
- 39. Akhtar K, Wang W, Ren G, Khan A, Feng Y, et al. 2018. Changes in soil enzymes, soil properties, and maize crop productivity under wheat straw mulching in Guanzhong, China. Soil and Tillage Research 182:94-102

- 40. Uzun I. 2004. Use of spent mushroom compost in sustainable fruit production. Journal of Fruit and Ornamental Plant Research 12:157-65
- 41. Lou Z, Zhu J, Wang Z, Baig SA, Fang L, et al. 2015. Release characteristics and control of nitrogen, phosphate, organic matter from spent mushroom compost amended soil in a column experiment. Process Safety and Environmental Protection 98:417-23
- 42. Zhu H, Sun L, Zhang Y, Zhang X, Qiao J. 2012. Conversion of spent mushroom substrate to biofertilizer using a stress-tolerant phosphate-solubilizing Pichia farinose FL7. Bioresource Technology 111:410-16
- 43. Oo AN, Iwai CB, Saenjan P. 2015. Soil properties and maize growth in saline and nonsaline soils using cassava-industrial waste compost and vermicompost with or without earthworms. Land Degradation & Development 26:300-10
- 44. Liu M, Li Y, Cher Y, Deng S, Xiao Y. 2017. Effects of different fertilizers on growth and nutrient uptake of Lolium multiflorum grown in Cd-contaminated soils. Environmental Science and Pollution Research 24:23363-70
- 45. Masson-Boivin C, Sachs JL. 2018. Symbiotic nitrogen fixation by rhizobia-the roots of a success story. Current Opinion in Plant Biology 44:7-15
- 46. Guo C, Zhou J, Zhang Y. 2023. Host and nutrient mediated the synergistic effect of Arbuscular Mycorrhizal Fungi (AMF) and Rhizobiumon plant growth. Acta Agrestia Sinica 31:1931–38
- 47. Shantz AA, Lemoine NP, Burkepile DE. 2016. Nutrient loading alters the performance of key nutrient exchange mutualisms. Ecology | etters 19:20-28
- 48. Mohammadi-Sichani MM, Assadi MM, Farazmand A, Kianirad M, Ahadi AM, Ghahderijani HH. 2017. Bioremediation of soil contaminated crude oil by Agaricomycetes. Journal of Environmental Health Science and Engineering 15:8
- 49. Du Toit A. 2018. Attracting bacteria in the soil. Nature Reviews Microbiology 16:122
- 50. Malhi SS, Goerzen DW. 2010. Improving yield in alfalfa seed stands with balanced fertilization. Journal of Plant Nutrition 33:2157-66

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