


# Integrating spectral data and phylogeographic patterns to study plant genetic variation: a review

Jingxue Zhang<sup>1,2</sup>, Yuhong He<sup>2</sup>, Jiangui Liu<sup>3</sup>, Jibiao Fan<sup>1</sup>, Jiali Shang<sup>3\*</sup> and Xuebing Yan<sup>1\*</sup> 

<sup>1</sup> College of Animal Science and Technology, Yangzhou University, Yangzhou 225000, China

<sup>2</sup> Department of Geography, Geomatics and Environment, University of Toronto Mississauga, Mississauga L5L1C6, Canada

<sup>3</sup> Ottawa Research and Development Centre, Agriculture and Agri-Food Canada, Ottawa K1A0C6, Canada

\* Corresponding authors, E-mail: [jiali.shang@canada.ca](mailto:jiali.shang@canada.ca); [yxbbjzz@163.com](mailto:yxbbjzz@163.com)

## Abstract

The study of genetic variation is pivotal for understanding plant diversity and evolution. In recent years, remote sensing has played a significant role in phylogeography, facilitating the exploration of intricate relationships among genetics, spectral behavior, and evolution. This review article aims to present a comprehensive compilation of literature in two main areas: 1) investigating the potential of spectral data collected using remote sensing to study genetic diversity, and 2) using spectral characteristics to investigate functional dynamics associated with various phylogeographic patterns and identify genetic bases of important agronomic traits. Remote sensing has proven effective in detecting genetic variations across different geographical regions. Additionally, this review examines the limitations, challenges, and prospects associated with integrating remote sensing and phylogeography. In essence, phylogeographic studies offer theoretical insights into understanding the genetic mechanisms underlying functional variability observed in remotely sensed spectral data. Leveraging rapid technological advancements in remote sensing and data fusion approaches will lead to new understanding of plant genetic diversity and the functional significance of plant traits. This knowledge is invaluable for informing strategies for the management and conservation of natural resources.

**Citation:** Zhang J, He Y, Liu J, Fan J, Shang J, et al. 2024. Integrating spectral data and phylogeographic patterns to study plant genetic variation: a review. *Grass Research* 4: e011 <https://doi.org/10.48130/grares-0024-0009>

## Introduction

Phylogeographic methods provide valuable insights into how genetic patterns among populations relate to geographic environments. These studies highlight how the historical context of geographic adaptive radiation shapes genetic distribution patterns. Genetic variations across vast geographic regions serve as essential indicators for assessing the adaptability of various genotypes<sup>[1,2]</sup>. By studying the genetic variations and distributions across large geographic scales, scientists can gain a deeper understanding of how evolutionary processes influence population genetic patterns over time. Additionally, phylogeographic studies provide valuable approaches for understanding and interpreting genetic variations in adaptive phenotypic traits, shedding light on the evolutionary mechanisms behind functional adaptations to diverse environments. Viewing this through a phylogeographic lens, it becomes clear that genetic divergence plays a crucial role in regulating phenotypic traits to facilitate adaptation across various geographic sites and environmental conditions<sup>[3]</sup>.

To comprehensively understand the dynamics of phenotypic traits, it is imperative to gather genotypic information from large and diverse geographic locations. This involves conducting population genetics and phylogeographic analyses using gene sequences, genetic markers, and genome sequences, as exemplified in studies by Wiens et al.<sup>[4]</sup> & Wang et al.<sup>[5]</sup>. With the rapid advancements in genomic sequencing technology, various innovative analytical methods leveraging genomics have emerged. These include genome-wide

association studies (GWAS) and genomic selection (GS). These methods aim to correlate DNA polymorphism data with phenotypic traits, enabling the identification of the genetic basis underlying important phenotypic traits<sup>[6]</sup>.

Spectral variability in optical remote sensing data may reveal genetic diversity at distinct geographical sites characterized by environmental heterogeneity<sup>[7–9]</sup>. The optical properties of plants, influenced by phenotypic traits such as biochemical content, leaf and canopy structure, and physiological functions, contribute to measurable spectral variability. Spectral information proves to be instrumental in accurately estimating genotype-specific phenotypic features. Consequently, spectral variation has the potential to capture genetic variations, particularly related to plant biochemical, physiological, and structural divergence. Using a combination of tools, including visual inspection, DNA molecular markers, and spectrometry<sup>[10,11]</sup>, it is possible to improve the efficiency for identifying genetic variation in plant materials. As demonstrated by Bush et al.<sup>[12]</sup>, phylogeographic affinities, genetic loci of functional traits, and spectral properties can be leveraged to parameterize process-based neutral and adaptive landscape genetics for biodiversity research.

Manually collecting phenotypic information across large areas is time-consuming and susceptible to data quality issues due to various factors during the data collection stages. Remote sensing offers an efficient alternative for gathering phenotypic data on plant diversity across different genetic groups in a short time frame. This approach helps to reduce

both the cost and time required for phenotyping while maintaining a high level of data accuracy and consistency<sup>[13]</sup>.

By using remote sensing, researchers can swiftly collect data and identify key phenotypic traits associated with physiology and biochemistry. This enhances our understanding of plant characteristics and their genetic variations. Studies have demonstrated a notable correlation between phenotypic traits measured using remote sensing data obtained from unmanned aerial vehicles (UAV) and the data obtained *in situ*<sup>[14]</sup>. The correlation underscores the reliability and potential of remote sensing in phenotypic data collection and analysis, marking a significant advancement in the study of plant genetics and phenotypes. High-throughput phenotyping technology, including 3D modeling, has played a pivotal role in large-scale quantitative trait loci (QTL) analysis. This technology has been instrumental in uncovering the genetic architecture governing dynamic plant growth in maize<sup>[15]</sup>, the genetic control of leaf elongation in barley (*Hordeum vulgare*)<sup>[16]</sup>, and the genomic prediction for canopy height in wheat<sup>[17]</sup>. The integration of genomic information under specific environmental conditions with plant ontogenetical, physiological, and biochemical properties, assessed through image-analysis-based phenotypic information, has enabled the targeted selection of more suitable cultivars for breeding purposes<sup>[18]</sup>.

Remote sensing technology has the potential to bridge the gaps between genotype and phenotype study, by alleviating the challenges in large amounts of data collection across extensive geographic extent. By supplying consistent and reliable spectral and phenotypic measurements, remote sensing proves invaluable in the field of phylogeography, offering insights into the natural dynamics of essential agronomic or ecological traits across evolutionary history. In the review, we described applications of remote sensing technology on phylogeographic studies from two aspects. The initial aspect revolves around assessing genetic variation through analyses of spectral variability, while the second centers on investigating phenotypic dynamics for phylogeographic patterns to identify genetic basis linked to potential phenotypic traits through remote sensing technology.

To consolidate this information, we conducted a thorough review of relevant publications, encompassing diverse platforms, sensors, wavelength ranges, spatial resolutions, model optimization techniques, and analysis methods (Table 1)<sup>[19,20]</sup>. We also summarized the advantages and disadvantages of remote sensing technologies in phenotypic measurements (Table 2). Finally, the review provides the deficiency and opportunities for future improvements in the cross-cutting research using remote sensing in phylogeography. Existing literatures demonstrate a noteworthy surge in interest regarding the evaluation of genetic variation by leveraging spectral features and identification of the genetic basis of remotely sensed phenotypic data (Figs 1 & 2). These studies play crucial roles in developing an integrated approach to identify, define, and conserve genotypic divergence, thereby enhancing our understanding of adaptive evolution and biodiversity. It aims to identify the genetic foundations of these adaptive traits, thereby contributing to advancements in genetic breeding and germplasm conservation.

In this study, we introduced a flowchart designed to facilitate the application of remote sensing technology on phylogeographic patterns (Fig. 3). The flowchart elucidates the

connections between various facets of the dataset, fostering a comprehensive understanding of their relationships.

## Identifying genotypic divergence through analysis of spectral variability

Recent research has underscored the role of genetic variation in regulating plant phenotypic traits by influencing the responses of physiological hormones to environmental conditions across diverse geographic locations<sup>[33]</sup>. Research has been undertaken to explore genetic variations by analyzing spectral variability and deriving phenotypic traits. These publications, along with our recent study on remote sensing in phylogeography<sup>[8–9, 23–25, 34, 35]</sup>, primarily focus on studying genetic divergence. These findings establish a robust foundation for understanding genetic variation revealed by spectral variability and exploring the practical applications of remotely sensed phenotypic traits in molecular breeding.

## Remote sensing technology

Remote sensing data acquired from satellites, UAVs, and ground-based platforms play a pivotal role in achieving high-throughput field phenotyping<sup>[36]</sup>. This allows for the characterization of genotypes in phylogeographic patterns across diverse environmental gradients. UAVs are especially useful in collecting remote sensing data for a large size of population and field studies. Ground-based platforms provide increased data resolution due to shorter distances between sensors and plant targets<sup>[31]</sup>. Various types of remotely sensed data, including those obtained from RGB, near infrared (NIR), short-wave infrared (SWIR) camera<sup>[37]</sup>, multispectral or hyperspectral sensors, light detection and ranging (LiDAR)<sup>[38,39]</sup>, and thermal sensors have been instrumental in assessing additional phenotypic traits indicative of genetic differences. VIS imaging systems such as RGB camera with three color sensors (blue, green and red) have been developed to measure plant growth, development, and responses to the environment<sup>[35]</sup>. Identified pixels through image processing algorithms are used for measuring the morphological (shape, structure), geometric (length, area), and color properties of each plant. NIR/SWIR cameras have spectral sensitivity in the range of 800–2,500 nm, correlated to plant growth rate, leaf water content, and leaf thickness<sup>[40]</sup>. The wavelength range of 800–1,700 nm encompasses three out of five water absorption regions. These regions can be future exploited to estimate the leaf water content. Thermal infrared cameras can detect long-wave infrared (LWIR) radiation that is emitted by objects to estimate leaf temperature<sup>[41]</sup>. Hyperspectral cameras with specific wavelength information can measure hundreds of spectral bands between 350 and 2,500 nm. Hyperspectral imaging has a promising potential for detecting plant traits related to abiotic and biotic stresses. Chlorophyll fluorescence imaging presents the potential capability to measure the quantum efficiency of photosystem II (PSII) in plant stress research<sup>[29]</sup>. Three-dimensional (3D) traits measured using laser-scanning techniques and depth/time-of-flight sensors can be employed to measure architectural features such as the number of leaves or tillers for analyzing plant growth and movement<sup>[42]</sup>. Remote sensing technologies allow the collection of high-throughput phenotypic traits that have precision capable of discerning genetic differences and facilitating the identification of genetic basis for adaptive plant traits.

**Table 1.** Approaches adopted to integrating remote sensing and phylogeography.

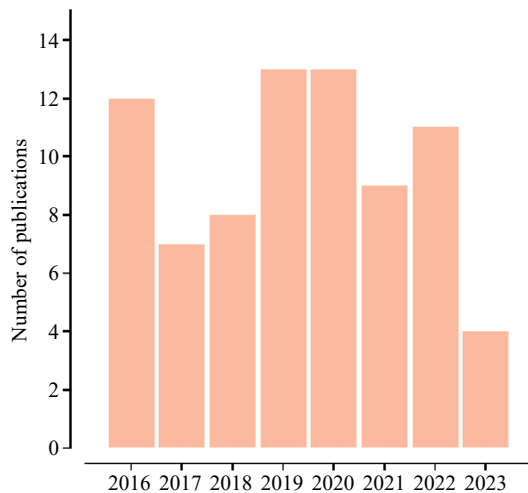
Spectral data	Applications related to phylogeographic relationship	Platforms	Sensors	Spatial resolution	Bands	Extracted features	Methods	Accuracy and references
Spectral variation related to genetic variation	Assessing genetic variation using variation in spectral patterns	Satellite	Landsat 5 TM USGS/ESPA	30 m	Blue (0.45–0.52 nm); Green (0.52–0.60 nm); Red (0.63–0.69 nm); Near Infrared (NIR) (0.76–0.90 nm); Shortwave Infrared (SWIR1) (1.55–1.75 nm); Thermal (10.40–12.50 nm); Shortwave Infrared (SWIR2) (2.08–2.3 nm)	Normalized difference vegetation index (NDVI); Normalized difference moisture index (NDMI)	Linear mixed effects model with the maximum-likelihood population effects (MLPE) parameterization	[21]
		ER-2 platform	NASA airborne visible/infrared imaging spectrometer (AVIRIS)	8 m	414–2,447 nm	NDVI; Water band index (WBI)	Partial least-squares discriminant analysis (PLSDA); Canonical correlation analyses (CCAs); Discriminant analyses; Linear-mixed models (GLMMs)	Kappa = 0.85–0.89[22]
		Automated tram	Portable field spectrometer (SVC HR-1024i; Spectra Vista); Imaging spectrometer (E Series; Headwall Photonics)	30 cm	340–2,500 nm	Leaf spectral reflectance	Partial least squares regression (PLSR); Mantel tests; D(TM) calculates; Linear regression models	[11]
		Airborne	Airborne imaging spectrometer (AIS); Field spectroradiometer (ASD)	2 m	372–2,540 nm	Spectral reflectance	Partial least squares (PLS) regression; PRESS statistic	RMSE = 0.2897[23]
		Ground	USB4000 portable equipment; Bruker FT-NIR MPA II (12,489–3,996 cm <sup>-1</sup> ); Portable thermo fisher MicroPHAZIR hyperspectral imaging system (HIS)	0.2 nm; 16 cm <sup>-1</sup> ; 16 cm <sup>-1</sup>	400–1,037 nm; 12,489–3,996 cm <sup>-1</sup> ; 1,595–2,396 nm	Spectral reflectance	Principal component analysis and soft independent modeling of class analogy (SIMCA)	Classification accuracy of 88%–91%[24]
				30 cm	400–1,000 nm	Leaf spectral reflectance	ParSketch-PLSDA method	Ranging from 81% to 96% for precision[25]
			ASD		350–2,500 nm	Leaf spectral reflectance	Partial least squares-discriminant analysis (PLS-DA); PCA	Kappa = 0.34 and 0.61[10]
	Classifying genotypic differentiation using spectrally derived traits	UAV	RGB or near-infrared, green and blue (NIR-GB) camera	40 m	Near-infrared, green and blue 350–2,500 nm	PHUAV for each plot from the DSM data	Genomic prediction modeling	r = 0.448 – 0.634[26]
Ground		ASD			Foliar traits	PLSR	RMSE = 9.1% – 19.4%[27]	
		LiDAR			Canopy height/structure	Fine-scale digital elevation models (DEMs)		

Table 1. (continued)

Spectral data	Applications related to phylogeographic relationship	Platforms	Sensors	Spatial resolution	Bands	Extracted features	Methods	Accuracy and references
Predicted phenotypes related to genetic pattern	Estimating the phenotypic dynamics under genetic pattern	UAV	12-megapixel DJI FC300X camera; 24-megapixel Sony a6000 RGB camera	Altitude of 25 and 120 m	Red-green-blue (RGB) bands	Canopy height metrics from the DSM data	Mixed linear models utilizing residual maximum likelihood (REML); Three-parameter Weibull sigmoid growth model	$R^2 > 99\%$ ; RMSE $< 4$ cm <sup>[28]</sup>
		The compact LIFT instrument	Blue light-emitting diode (LED); STS-VIS spectrometer (Ocean Optics, Florida, USA); RGB cameras	0.46 nm	445 nm; 400–800 nm	Fluorescence Data; NDVI; Alternative NDVI (NDVI_LII); Green normalized difference vegetation index (GNDVI); MERIS terrestrial chlorophyll index (MTCI); Photochemical reflectance index (PRI)	Least absolute shrinkage and selection operator (Lasso) regression; Linear modeling	$r = 0.7 - 0.92$ <sup>[29]</sup>
		UAV high-throughput phenotypic platforms (UAV-HTPPs) UAV and tractor	RGB or near-infrared, green, and blue (NIR-GB) camera  Multi-spectral camera; GreenSeeker spectral sensors	40–60 m  40–42 m for UAV; 1.32 m for tractor	  Green; Red; Red Edge; NIR; Blue	  Canopy cover; Canopy reflectance; Biomass	  Crop surface model; Ortho mosaics model	Accuracy for DSM was 2.31 cm/pixel <sup>[30]</sup>
UAVs	Large spatial scales for canopy measurement	UAV	RGB	18 m	RGB bands	Canopy cover; Canopy volume; and Excess greenness index.	Plant Growth Models; Variance inflation factor (VIF); GWAS	$R^2 = 0.02 - 0.11$ <sup>[31]</sup>
Satellites	Large spatial scales across landscapes and regions							$R^2 = 0.87 - 0.94$ <sup>[32]</sup>

Table 2. Advantages and disadvantages of remote sensing technologies in measuring plant traits.

Remote sensing platforms	Advantages	Disadvantages	Sensors	Measured traits
Ground-based platforms	High resolution; Not influenced by background for leaf measurement	Require long measurement time; Restricted by field conditions and challenging environment	Multispectral, hyperspectral sensors	Pigment concentrations; Quality traits (eg., oil, amylose, protein content, moisture); Total plant/canopy biomass; Yield
UAVs	Large spatial scales for canopy measurement	Low data resolution	Thermal infrared cameras	Leaf and canopy temperature
			VIS imaging systems	Morphological traits (eg., shape, structure, color properties); Geometric traits (eg., length, area, canopy cover, canopy volume); Photosynthetic behaviors; Pigment concentrations
			Near infrared (NIR), short-wave infrared (SWIR) camera Hyperspectral sensors	Leaf water content; Plant/canopy structure; Biomass
Satellites	Large spatial scales across landscapes and regions	Coarser spectral resolution; Lower spatial resolution; Susceptible to cloud interference	LIDAR	Pigment concentrations; Quality traits (eg., oil, amylose, protein content, moisture); Total plant/canopy biomass; Yield
			Thermal infrared cameras Multispectral, hyperspectral sensors	Plant structure (eg., crown density, structural complexity) Leaf and canopy temperature Vegetation structural properties (eg., species richness)

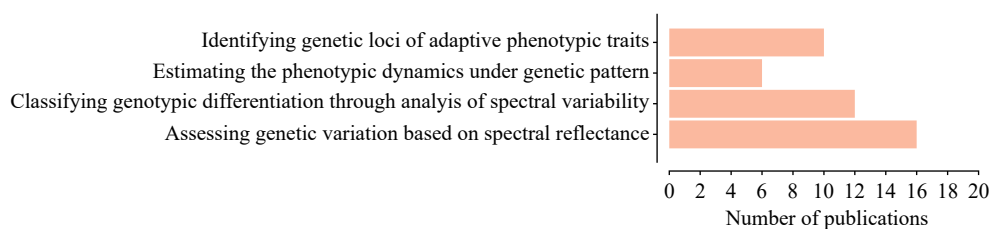


**Fig. 1** Annual number of publications related to the use of vegetation spectral data in phylogeography, from 2016 to 2023. These publications were searched using keywords 'vegetation spectral' and 'phylogeography', from the Web of Science.

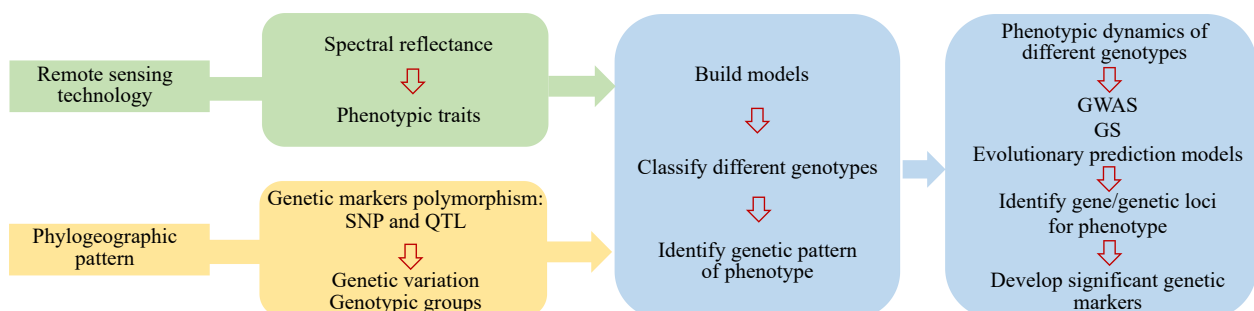
### Analysis of spectral variability for revealing genetic diversity

Establishing the correlation of remotely sensed data with empirically measured and biologically relevant traits is the greatest challenge in revealing genetic variation through analysis of spectral variability. Spectral wavelengths sensitive to genetic discrimination are closely linked to photosynthetic pigments, proteins, and plant fibers, suggesting that genetic variation in phylogeography is manifested in biochemical characteristics<sup>[24]</sup>. Infrared spectra in different regions of visible, near IR and mid-IR spectrum could allow capturing the phylogeographic signal in *Araucaria araucana* that separates Coastal and Andean origins<sup>[24]</sup>. This capability further reveals unique genetic patterns underlying diverse phenotypic traits. In these studies, biological, chemical, and structural traits were

extracted through remote sensing techniques, examining a range of species including crops such as rice, maize, and wheat<sup>[36]</sup>, as well as trees like Oaks and *Araucaria Araucana* and grass. Spectral variability related to physiological and highly heritable agronomic traits, such as plant height offers the potential to study genetic variation. RGB and near-infrared cameras mounted on UAV platforms facilitate the monitoring of plant height in maize and sorghum over extensive spatial regions and capturing equivalent genetic variation to manual measurements<sup>[28, 43]</sup>. The UAV imagery technology can be used for estimating variations in plant height across different genotypes and several genetic mapping populations. This has the potential to unveil genetic diversity. Plant height estimates of maize have previously offered evidence of equivalent or greater phenotypic variation partitioned to genetic factors. Genetic variation ( $s^2_G$ ) and repeatability (R) estimates presented the percentage of genetic variation explained by plant height using RGB imagery, which can be used to compare the accuracy of plant height estimates among different UAV platforms and different canopy structures<sup>[44]</sup>. In a study to understand drought-adaptive traits, Condorelli et al.<sup>[31]</sup> used multi-spectral cameras mounted on UAV and tractor-based platforms to gather NDVI measurements in a high-throughput manner. NDVI serves as an integrative measure to correlate with chlorophyll content and total plant/canopy biomass, showing sensitivity features to genetic variation and capacities to discriminate genotypes<sup>[45]</sup>. Important traits for forest structure, such as crown density and structural complexity could reveal genetic-based diversity within and between species through rapid phenotyping using ultra-dense UAV-LiDAR point clouds<sup>[36]</sup>. Certain spectral indices derived from optical remote sensing data demonstrate the potential to identify genetic variation in plants. The application of remotely sensed phenotypic traits in estimating genetic variation suggests that these traits may exhibit heritable genetic diversity in response to diverse environmental conditions.



**Fig. 2** The number of publications on different types of research relevant to integration of vegetation spectral data and phylogeography, from 2016 to 2023.



**Fig. 3** Flowchart illustrating the combination of remote sensing and phylogeography to study the genetic patterns.



Advances in computer vision-assisted analysis methods in high-throughput plant phenotyping data are needed to improve further. The soft independent modeling of class analogy (SIMCA) presented high prediction capability to discriminate phylogeographic origin with a classification accuracy of 88%–91%<sup>[24]</sup>. The mixed linear model, employing residual maximum likelihood (REML), in conjunction with the Weibull sigmoid growth model, has been used to estimate genetic variation based on predicted plant height and growth parameters<sup>[28]</sup>. Significant genetic variation was detected at the provenance level in *E. pauciflora* and *E. tenuiramis* by calculating genetic parameters including the quantitative inbreeding coefficient ( $Q_{ST}$ ) and narrow-sense heritability ( $h^2_{OP}$ ) estimates for remote sensing data derived from individual tree LiDAR point clouds<sup>[36]</sup>.

## Identifying genetic loci through analysis of phenotypic dynamics derived from remote sensing

In the realms of biodiversity and plant breeding, researchers have endeavored to elucidate the genetic mechanisms governing essential phenotypic traits, leading to the identification of numerous genetic loci associated with phenotyping across diverse genotypic groups<sup>[46]</sup>. A thorough exploration of phenotypic information is needed to ascertain its suitability for detecting genetic variations in phylogeographic patterns, spanning from the original geographic sites to extended regions. Phenotypic measurements of samples with phylogeographic patterns provide the dynamic nature of many important agronomical traits to adapt to different environments at large geographic scales. These biochemical traits derived from remote sensing technology have the potential for studying the association of genetic diversity and phenotypic traits. By analyzing spectral data that captures phenotypic changes across diverse genotypic groups and aligns with phylogeographic patterns, it becomes feasible to identify genes and genetic loci influencing the development of specific phenotypic traits. The capability facilitates the rapid discovery of new phenotypes<sup>[35]</sup>. The genetic architecture of plant traits has been determined to identify some large effect loci likely fixed during domestication and early selection<sup>[47]</sup>. The acquisition of high-throughput phenotypic data through remote sensing has proven to be pivotal in the development of molecular markers for spinach (*Spinacia oleracea* L.)<sup>[32]</sup>.

## Phenotypic dynamics in phylogeographic pattern detected through remote sensing technology

The high-throughput nature of phenotyping facilitates the examination of dynamic mechanisms in plant functional traits of genetically diverse groups. With the decreasing cost of genome-wide genotyping, large-scale phenotyping through remote sensing has become a crucial tool for exploring significant phenotypic variation attributed to genetic factors<sup>[43,44,48]</sup>. The recent progress in high-throughput phenotyping platforms (HTTPs) based primarily on the use of ground based platforms and/or UAVs provides unprecedented opportunities to accurately measure proxy traits in hundreds of plots<sup>[49–52]</sup>, as required in experiments to identify QTLs. An UAV remote sensing experiment, featuring repeated measurements every month with a flight duration of approximately 10 min, demonstrated a performance in measuring sorghum plant height

comparable to manual phenotypic measurements for genomics-assisted breeding<sup>[26]</sup>. Increased ability of aerial over ground-based platforms has been observed to detect quantitative trait loci (QTLs) for NDVI, particularly under terminal drought stress<sup>[28]</sup>. Various remote sensing techniques are commonly used to explore the phenotypic dynamics within phylogeographic relationships. NDVI was extracted from georeferenced ortho mosaic GeoTIFFs generated from imagery captured from multi-spectral cameras carried on UAVs and tractor-based platforms. The NDVI data has been employed to ascertain the genetic basis for grain yield using GWAS analysis<sup>[53]</sup>. Both airborne laser scanning and imaging spectroscopy have proven valuable for mapping phenotypic variation within species, particularly in tree functional traits<sup>[54]</sup>. Canopy cover, assessed through digital cameras, exhibits correlations with plant density, early vigor, leaf size attributed to genetic variation<sup>[55]</sup>. Thermal sensors emerge as effective tools for measuring canopy temperature, offering valuable insights into estimating water stress levels<sup>[56]</sup>. Furthermore, the integration of multispectral and hyperspectral imaging from different sensors is successful in discriminating and mapping long-range spatial phenotypic features<sup>[57,58]</sup>.

As we unravel the underlying genetic loci governing the variability of remotely sensed phenotypic traits, spectral-related proxies such as NDVI play increasingly vital roles in selecting valuable cultivars. These plant traits like plant height, dry biomass, leaf rolling, and phenology score obtained using remote sensing technology, chosen based on genetic information obtained from identified loci, facilitate the optimization of desirable traits in breeding programs<sup>[59–62]</sup>. Researchers have explored phenotypic dynamics in plants exhibiting phylogeographic patterns using the Normalized Difference Vegetation Index (NDVI)<sup>[51,63,64]</sup> as well as traits related to drought adaptation<sup>[31]</sup>. NDVI is proven to be useful in predicting canopy biomass and nitrogen status<sup>[55]</sup>. Using an optical sensor to detect NDVI values, researches identified a total of 44 genetic loci across different environments linked to traits such as stay-green (Stg) at physiological maturity, yield components, plant height, and yield-related physiological traits in wheat<sup>[52,65,66]</sup>. In the context of spinach, GWAS was employed to identify SNPs associated with growth parameters such as canopy cover, canopy volume, and excess green index (ExG) measured using RGB sensor mounted on UAVs<sup>[32]</sup>. The canopy height measurements and their respective growth rates obtained using UAV- RGB and NIR-GB camera identified unique QTLs and candidate genes controlling plant height in the GWAS study<sup>[67]</sup>. Furthermore, remote sensing techniques have been employed to investigate the adaptive significance of photosynthetic pathways in the grass family (Poaceae) under different light environments. These studies aimed to test ecological theories concerning adaptive evolution<sup>[34,68,69]</sup>. The integration of spectral data and phylogeography has proven instrumental in systematically examining the genetic architecture of agronomic traits<sup>[70,71]</sup> and gene-environment interactions<sup>[72]</sup>. These valuable insights, in turn, contributed significantly to the conservation and breeding of plant germplasm resources.

## Association analysis between genetic variation and phenotypic traits

To examine the dynamic interplay between phenotypic changes and phylogeographic patterns, it is crucial to employ a

variety of analytical methods. These methods are essential to elucidating the relationships between phylogeography and the expression of phenotypic traits derived from remote sensing data. The integration of spectroscopy-based functional traits with phylogeography can be achieved using Mantel tests, allowing for the assessment of correlations between their dissimilarity matrices. Additionally, Partial least squares (PLS) regression can be used to predict the likelihood of populations belonging to a particular genetic cluster based on their similarity in predicted remotely sensed phenotypic traits<sup>[9,23]</sup>. Notably, genomic prediction models have demonstrated a remarkable resemblance in sorghum between plant height measurements obtained *via* remote sensing and those obtained through conventional methods<sup>[26]</sup>.

Capitalizing on the rich genetic variation inherent in phylogeographic relationships and the phenotypic diversity extracted from remote sensing data across extensive geographic regions, GWAS presents a remarkable opportunity to pinpoint numerous significant genetic loci under natural phenotypic variation. The genomic study conducted by Huang et al.<sup>[73]</sup> pinpointed crucial components, including single-nucleotide polymorphisms (SNPs), QTLs, and sequencing-based genotypes, all exhibiting significant associations with phenotypic traits such as plant height and some growth parameters through GWAS<sup>[32,53]</sup>. This approach facilitates the optimization of marker development, particularly for complex agronomic traits<sup>[70,72]</sup>. Genomic selection model refers to an accurate prediction model using genomic selection (GS). UAVs-based plant height estimates in sorghum could perform similar results to manual measurements in the genomic prediction model<sup>[26]</sup>. Identification of QTL hotspots provided insight for marker-assisted and genomic selection analyses in sorghum, rice, and wheat breeding<sup>[6]</sup>. Advanced models seek to identify the genetic basis responsible for adaptive traits by leveraging extensive datasets of genomes and phenotypes. Uncovering specific genetic loci or candidate genes holds the potential to significantly enhance agriculturally important traits through selective breeding.

## Key factors influencing remote sensing applications in phylogeography

### Sources of origin of spectral variability

Remotely sensed spectral data can be obtained at various levels, including leaf, plant, canopy, and landscape, each exhibiting distinct sensitivity to phylogeographic patterns. Variability in biophysical parameters such as leaf area index (LAI), height, plant, and canopy structure, can impact spectral measurements. The biochemical components like pigments, biomass, water, and N contents, as well as functional components like photosynthesis, fluorescence, and stress response, may also influence spectral measurements, thus potentially confounding the study on relationships between remote sensing and phylogeography<sup>[8]</sup>. To gather diverse information and crucial phenotypic trait-related characteristics from field-grown plants, it is imperative to carefully select appropriate platforms and remote sensing sensors. High-throughput phenotypic measurements, facilitated by hyperspectral and multispectral sensors, often complimented with active LIDAR, have proven effective in measuring canopy reflectance, estimating plant height, and assessing biomass<sup>[49,74–76]</sup>. For instance, sensors equipped with a NIR band may outperform those limited to the

visible range alone in genomic prediction modeling<sup>[26]</sup>. Hyperspectral imaging has been instrumental in examining pigments, as well as quality traits such as oil, amylose, protein content, and moisture in whole rice grains and other crops. Additionally, it has been applied to assess N content and biomass in plants<sup>[77,78]</sup>.

### Spatial scales

Considerations for spatial scales are important when using remote sensing methods for targeted vegetation parameters or phenotypic traits. Factors such as plant and canopy structures, their physical and biochemical compositions, background soils and atmospheric conditions, and platforms, all influence spectral signals<sup>[79,80]</sup>. Therefore, spectral data acquired at different scales will be impacted by different interference factors. For instance, a measurement at the leaf scale may not be influenced by soil properties and canopy structures, so leaf-related traits would be measured with more confidence than plant or canopy-related traits. Canopy or landscape properties can be measured at a larger spatial scale, however, leaf level properties may only be estimated through empirical models or inversion of physical models with large uncertainty. Ground-based phenotyping platforms require a longer time to complete the measurements as compared to UAV-based remote sensing. The advantage of UAV-based phenotyping over a larger area in less time could minimize the effects of daily environmental fluctuations on spectral measurements<sup>[31]</sup>.

### Spectral scales

Spectral resolution and wavelength ranges play important roles in shaping the connections between spectral variability and phylogeographic patterns. Spectral signatures in visible ranges are sensitive to pigment concentrations and photosynthetic behaviors. Spectral reflectance in near-infrared to short-wave infrared ranges is sensitive to water content, biomass, and plant/canopy structures. Sensors with a high spectral resolution can capture narrow spectral signatures or targets. For instance, estimation of red-edge position and chlorophyll fluorescence requires high to super-high spectral resolution. Accurate estimation of plant water content may also benefit from narrow spectral resolution to eliminate influence from atmospheric water vapor content variation. Sensors with coarser spectral resolution are common and affordable, such as many multi-spectral sensors mounted on satellite platforms, but still be valuable for vegetation structural properties. Current ground- and UAVs-based remote sensing technologies primarily concentrate on NDVI-related bands, such as red and near-infrared, for dissecting genetic factors associated with drought adaptive traits in durum wheat<sup>[31]</sup>. The study by Hernandez-Stefanoni et al.<sup>[81]</sup> demonstrated that the Landsat shortwave infrared band had a robust correlation with species richness in tropical forest ecosystems. Therefore, the careful selection of appropriate spectral resolution and spectral ranges is imperative to enhance the integration of spectral variability and genetic diversity.

### Temporal scales

The spectral data collected from large-scale experiments are significantly influenced by both the stage of plant growth and daily fluctuations in environmental conditions<sup>[5]</sup>. Consequently, variations in leaf age and seasonal adjustments can introduce spectral variability within individual canopies, potentially complicating the relationship between spectral diversity and

phylogeography<sup>[82–84]</sup>. By examining the seasonal dynamics of phenotypic traits using multi-temporal remote sensing data, we gain valuable insights into how spectral reflectance, genotype, and the environment are interconnected. A comprehensive understanding of the temporal changes in spectral diversity improves our ability to accurately capture genetic variation through spectral variation and remotely sensed phenotypic traits across different growth stages<sup>[30,85]</sup>.

## Challenges and constraints in integrating remote sensing with phylogeography

In this paper, we present an in-depth investigation of research dedicated to the application of remote sensing in phylogeography. Despite significant advancements, there remain persistent overarching limitations and challenges in this interdisciplinary approach. These challenges stem from a lack of physical models linking spectral measurements with phylogeographic patterns and a deficiency in systematic and consistent data acquisition at large scales and quantities.

One significant constraint associated with remote sensing data is spatial resolution. Satellite imagery and other remote sensing technologies often have limited resolution, making it difficult to capture subtle genetic patterns at fine scales. As phylogeographic studies increasingly delve into population-level analyses, the growing gaps between genetic data and remote sensing data become more apparent.

The lack of temporal congruence between these data sources can introduce uncertainties in deciphering the link between spectral variability and genetic diversification. Remote sensing data offers snapshots of the landscape at specific time points, while phylogeographic processes unfold over evolutionary timescales. Achieving temporal synchronization is crucial but labor-intensive and complex.

The effective integration of remote sensing and phylogeography relies on the availability and accessibility of relevant datasets. Acquiring suitable remote sensing data and genetic information for study areas, especially in remote or politically sensitive regions, can be challenging. Additionally, disparities in data specifications and quality across diverse sources may hinder effective integration.

## Efforts to mitigate integration challenges

The integration of various data sources, UAVs, satellite imagery, and ground instrumentation, is essential for overcoming limitations in spatial coverage and data quality inherent in individual platforms<sup>[6]</sup>. UAVs offer high-resolution data but suffer from limited battery life and flight duration, while satellites provide broader coverage but lower spatial resolution and susceptibility to cloud cover. Ground-based instruments excel in accuracy but face challenges in certain environments like tall crops or muddy fields<sup>[86]</sup>.

To address these limitations, an integrated approach leveraging data from multiple platforms can be employed to extract both spectral and phenotypic information across geographical areas. However, ensuring consistency and high standards in imaging processing across different platforms, sensors, and timeframes is crucial for subsequent data analysis<sup>[87]</sup>.

Cloud-based image analysis platforms like Google Earth Engine (GEE)<sup>[88]</sup>, Amazon Web Services<sup>[89]</sup>, Landsat Cloud

(USGS), Copernicus Hub (ESA), and EOS Platform have become common for processing extensive datasets from various sensors<sup>[90]</sup>. These platforms facilitate the accurate remote sensing images processing and enable the selection of suitable imagery based on study parameters.

The adoption of cloud-based platforms, particularly GEE, has gained traction in biological studies<sup>[91]</sup>. Researchers have used these platforms for various applications, such as mapping forest changes and monitoring wildlife populations<sup>[92–94]</sup>. For instance, Millary<sup>[95]</sup> efficiently used the EOS Platform to analyze changes in vegetation greenness in the Cirebon Regency using the NDVI algorithm obtained from Landsat series images.

Furthermore, advancement in equipment accuracy and image processing techniques, along with rigorous plant measurements and outlier removal, are essential for enhancing the effectiveness of plant breeding initiatives. A deeper investigation into the connection between plant phylogeography, functional traits, and spectral attributes is needed for a comprehensive understanding of evolutionary dynamics and genetic diversity identification.

While remote sensing data provides valuable insights into environmental factors influencing phylogeographic processes, it captures only a fraction of these variables. Integrating analyses with additional environmental data sources and employing ecological modeling approaches become imperative to enrich the explanatory capacity of such studies<sup>[96]</sup>. However, careful interpretation and validation are required when connecting genetic patterns with remotely sensed environmental variables, considering potential confounding factors and the need for additional experimental validation.

## Conclusions

The use of remote sensing in phylogeography offers a valuable interdisciplinary approach to comprehending adaptive evolution processes and landscape genetics. This integration enables these researchers to pursue two primary objectives: 1) modeling and assessing temporal and spatial patterns of genetic variation through diverse spectral features obtained using remote sensing technology; and 2) identifying the genetic basis of specific phenotypic traits predicted from spectral data. We illustrate the estimation of genotypic divergence through the analysis of vegetation spectral variability. Additionally, we elucidate that the functional dynamics inherent in evolutionary processes can be studied by combining remote sensing technology with phylogeographic patterns. Furthermore, we demonstrate the identification of genetic loci associated with phenotypes derived from remote sensing data.

For the effective and meaningful application of remote sensing and phylogeography in biodiversity and plant breeding, amalgamating diverse data, methods, and theories is imperative. However, researchers must remain cognizant of inherent limitations and challenges. Overcoming spatial and temporal discrepancies, enhancing data availability and quality, addressing scale-related issues, and accounting for complex environmental interactions are pivotal steps in maximizing the potential of this integration. By overcoming these challenges, researchers can uncover profound insights into the intricate relationships between landscapes, environmental changes, and genetic variation.



## Author contributions

The authors confirm contribution to the paper as follows: study conception and design: Zhang J, Shang J, Yan X; draft manuscript preparation: Zhang J; manuscript editing and refining: He Y, Liu J, Shang J, Yan X, Fan J. All authors reviewed the results and approved the final version of the manuscript.

## Data availability

Data sharing not applicable to this article as no datasets were generated or analyzed during the current study.

## Acknowledgments

The authors would like to acknowledge National Natural Science Foundation of China (32171672) and Excellent Doctoral Thesis Project in Yangzhou University for partial funding of this effort.

## Conflict of interest

The authors declare that they have no conflict of interest.

## Dates

Received 2 February 2024; Revised 23 March 2024; Accepted 3 April 2024; Published online 6 May 2024

## References

- Mimura M, Yahara T, Faith DP, Vázquez-Domínguez E, Colautti RI, et al. 2017. Understanding and monitoring the consequences of human impacts on intraspecific variation. *Evolutionary Application* 10:121–39
- Urban MC, Bocedi G, Hendry AP, Mihoub JB, Pe'er G, et al. 2016. Improving the forecast for biodiversity under climate change. *Science* 353:aad8466
- Des Roches S, Post DM, Turley NE, Bailey JK, Hendry AP, et al. 2018. The ecological importance of intraspecific variation. *Nature Ecology & Evolution* 2:57–64
- Wiens JJ, Ackerly DD, Allen AP, Anacker BL, Buckley LB, et al. 2010. Niche conservatism as an emerging principle in ecology and conservation biology. *Ecology Letters* 13:1310–24
- Wang F, Chen B, Lin X, Zhang H. 2020. Solar-induced chlorophyll fluorescence as an indicator for determining the end date of the vegetation growing season. *Ecological Indicators* 109:105755
- Jung J, Maeda M, Chang A, Bhandari M, Ashapure A, Landivar-Bowles J. 2021. The potential of remote sensing and artificial intelligence as tools to improve the resilience of agriculture production systems. *Current Opinion in Biotechnology* 70:15–22
- Ustin SL, Gamon JA. 2010. Remote sensing of plant functional types. *New Phytologist* 186:795–816
- Cavender-Bares J, Meireles JE, Couture JJ, Kaproth MA, Kingdon CC, et al. 2016. Associations of leaf spectra with genetic and phylogenetic variation in oaks: prospects for remote detection of biodiversity. *Remote Sensing* 8:221
- Schweiger AK, Cavender-Bares J, Townsend PA, Hobbie SE, Madritch MD, et al. 2018. Plant spectral diversity integrates functional and phylogenetic components of biodiversity and predicts ecosystem function. *Nature Ecology and Evolution* 2:976–82
- Rohman A, Nugroho A, Lukitaningsih E, Sudjadi. 2014. Application of vibrational spectroscopy in combination with chemometrics techniques for authentication of herbal medicine. *Applied Spectroscopy Reviews* 49:603–13
- Rohman A, Windarsih A, Hossain MA, Johan MR, Ali ME, et al. 2019. Application of near- and mid-infrared spectroscopy combined with chemometrics for discrimination and authentication of herbal products: a review. *Journal of Applied Pharmaceut Science* 9:137–47
- Bush A, Sollmann R, Wilting A, Bohmann K, Cole B, et al. 2017. Connecting Earth observation to high-throughput biodiversity data. *Nature Ecology & Evolution* 1:0176
- Shi Y, Thomasson JA, Murray SC, Pugh NA, Rooney WL, et al. 2016. Unmanned aerial vehicles for high-throughput phenotyping and agronomic research. *PLoS One* 11:e0159781
- Bendig J, Bolten A, Bennertz S, Broscheit J, Eichfuss S, et al. 2014. Estimation biomass of barley using crop surface models (CMSs) derived from UAV-based RGB imaging. *Remote Sensing* 6:10395–412
- Zhang X, Huang C, Wu D, Qiao F, Li W, et al. 2017. High-throughput phenotyping and QTL mapping reveals the genetic architecture of maize plant growth. *Plant Physiology* 173:1554–64
- Ward B, Brien C, Oakey H, Pearson A, Negrão S, et al. 2019. High-throughput 3D modelling to dissect the genetic control of leaf elongation in barley (*Hordeum vulgare*). *The Plant Journal* 98:555–70
- Lyra DH, Virlet N, Sadeghi-Tehran P, Hassall KL, Wingen LU, et al. 2020. Functional QTL mapping and genomic prediction of canopy height in wheat measured using a robotic field phenotyping platform. *Journal of Experimental Botany* 71:1885–98
- Walter A, Liebisch F, Hund A. 2015. Plant phenotyping: from bean weighing to image analysis. *Plant Methods* 11:14
- Del Pozo S, Rodríguez-González P, Hernández-López D, Felipe-García B. 2014. Vicarious radiometric calibration of a multispectral camera on board an unmanned aerial system. *Remote Sensing* 6:1918–37
- Virlet N, Sabermanesh K, Sadeghi-Tehran P, Hawkesford MJ. 2017. Field Scanalyzer: an automated robotic field phenotyping platform for detailed crop monitoring. *Functional Plant Biology* 44:143–53
- Gutiérrez-Rodríguez J, Gonçalves J, Civantos E, Martínez-Solano I. 2017. Comparative landscape genetics of pond-breeding amphibians in mediterranean temporal wetlands: the positive role of structural heterogeneity in promoting gene flow. *Molecular Ecology* 26:5407–20
- Madritch MD, Kingdon CC, Singh A, Mock KE, Lindroth RL, et al. 2014. Imaging spectroscopy links aspen genotype with below-ground processes at landscape scales. *Philosophical Transactions of the Royal Society B* 369:20130194
- Czyż EA, Guillén Escribà C, Wulf H, Tedder A, Schuman MC, et al. 2020. Intraspecific genetic variation of a *Fagus sylvatica* population in a temperate forest derived from airborne imaging spectroscopy time series. *Ecology and Evolution* 10:7419–30
- Rojas-Rioseco M, del Castillo RP, González-Campos J, Ipinza R, Sanhueza MI, et al. 2023. Phylogeographic origin authentication of *Araucaria araucana* (Mol.) K Koch seedlings through the application of spectroscopy techniques in different infrared ranges and chemometric methods. *New Forests* 54:467–89
- Ryckewaert M, Metz M, Héran D, George P, Grèzes-Besset B, et al. 2021. Massive spectral data analysis for plant breeding using parSketch-PLSDA method: discrimination of sunflower genotypes. *Biosystems Engineering* 210:69–77
- Watanabe K, Guo W, Arai K, Takanashi H, Kajiya-Kanegae H, et al. 2017. High-throughput phenotyping of sorghum plant height using an unmanned aerial vehicle and its application to genomic prediction modeling. *Frontiers in Plant Science* 8:421
- Wang Z, Chlus A, Geygan R, Ye Z, Zheng T, et al. 2020. Foliar functional traits from imaging spectroscopy across biomes in the eastern North America. *New Phytologist* 228:494–511
- Anderson SL, Murray SC, Chen Y, Malambo L, Chang A, et al. 2020. Unoccupied aerial system enabled functional modeling of maize height reveals dynamic expression of loci. *Plant Direct* 4:e00223

29. Keller B, Matsubara S, Rascher U, Pieruschka R, Steier A, et al. 2019. Genotype specific photosynthesis x environment interactions captured by automated fluorescence canopy scans over two fluctuating growing seasons. *Frontiers in Plant Science* 10:1482
30. Wang R, Gamon JA. 2019. Remote sensing of terrestrial plant biodiversity. *Remote Sensing of Environment* 231:111218
31. Condorelli GE, Maccaferri M, Newcomb M, Andrade-Sanchez P, White JW, et al. 2019. Comparative aerial and ground based high throughput phenotyping for the genetic dissection of NDVI as a proxy for drought adaptive traits in durum wheat. *Front Plant Science* 9:893
32. Awika HO, Bedre R, Yeom J, Marconi TG, Enciso J, et al. 2019. Developing growth-associated molecular markers via high-throughput phenotyping in Spinach. *The Plant Genome* 12:190027
33. Wang Y, Zhao J, Lu W, Deng D. 2017. Gibberellin in plant height control: old player, new story. *Plant Cell Reports* 36:391–98
34. Zhang J, Han M, Wang L, Chen M, Chen C, et al. 2023. Study of genetic variation in bermuda grass along longitudinal and latitudinal gradients using spectral reflectance. *Remote Sensing* 15:896
35. Yang W, Guo Z, Huang C, Duan L, Chen G, et al. 2014. Combining high-throughput phenotyping and genome-wide association studies to reveal natural genetic variation in rice. *Nature Communication* 5:5087
36. Camarretta NA, Harrison P, Lucieer A, Potts BM, Davidson N, et al. 2020. From drones to phenotype: using UAV-LiDAR to detect species and provenance variation in tree productivity and structure. *Remote Sensing* 12:3184
37. Malambo L, Popescu SC, Murray SC, Putman E, Pugh NA, et al. 2018. Multitemporal field-based plant height estimation using 3D point clouds generated from small unmanned aerial systems high-resolution imagery. *International Journal of Applied Earth Observation and Geoinformation* 64:31–42
38. Torabzadeh H, Leiterer R, Hueni A, Schaepman ME, Morsdorf F. 2019. Tree species classification in a temperate mixed forest using a combination of imaging spectroscopy and airborne laser scanning. *Agricultural and Forest Meteorology* 279:107744
39. Valbuena R, O'Connor B, Zellweger F, Simonson W, Vihervaara P, et al. 2020. Standardizing ecosystem morphological traits from 3D information sources. *Trends in Ecology & Evolution* 35:656–67
40. Fahlgren N, Gehan MA, Baxter I. 2015. Lights, camera, action: high-throughput plant phenotyping is ready for a close-up. *Current Opinion in Plant Biology* 24:93–99
41. Sirault XRR, James RA, Furbank RT. 2009. A new screening method for osmotic component of salinity tolerance in cereals using infrared thermography. *Function Plant Biology* 36:970–77
42. Dornbusch T, Michaud O, Xenarios I, Fankhauser C. 2014. Differentially phased leaf growth and movements in *Arabidopsis* depend on coordinated circadian and light regulation. *The Plant Cell* 26:3911–21
43. Pugh NA, Horne DW, Murray SC, Carvalho G Jr, Malambo L, et al. 2018. Temporal estimates of crop growth in sorghum and maize breeding enabled by unmanned aerial systems. *The Plant Phenome Journal* 1:1–10
44. Anderson SL, Murray SC, Malambo L, Ratcliff C, Popescu S, et al. 2019. Prediction of maize grain yield before maturity using improved temporal height estimates of unmanned aerial systems. *The Plant Phenome Journal* 2:1–15
45. Christopher JT, Christopher MJ, Borrell AK, Fletcher S, Chenu K. 2016. Stay-green traits to improve wheat adaptation in well-watered and water-limited environments. *Journal of Experimental Botany* 67:5159–72
46. Pan Q, Xu Y, Li K, Peng Y, Zhan W, Li W, et al. 2017. The genetic basis of plant architecture in 10 maize recombinant inbred line populations. *Plant Physiology* 175:858–73
47. Peiffer JA, Romay MC, Gore MA, Flint-Garcia SA, Zhang Z, et al. 2014. The genetic architecture of maize height. *Genetics* 196:1337–56
48. Araus JL, Cairns JE. 2014. Field high-throughput phenotyping: the new crop breeding frontier. *Trends in Plant Science* 19:52–61
49. Duan T, Zheng B, Guo W, Ninomiya S, Guo Y, et al. 2016. Comparison of ground cover estimates from experiment plots in cotton, sorghum and sugarcane based on images and ortho-mosaics captured by UAV. *Functional Plant Biology* 44:169–83
50. Shakoor N, Lee S, Mockler TC. 2017. High throughput phenotyping to accelerate crop breeding and monitoring of diseases in the field. *Current Opinion in Plant Biology* 38:184–92
51. Trapp JJ, Urrea CA, Zhou J, Khot LR, Sankaran S, et al. 2016. Selective phenotyping traits related to multiple stress and drought response in dry bean. *Crop Science* 56:1460–72
52. Shi S, Azam FI, Li H, Chang X, Li B, et al. 2017. Mapping QTL for stay-green and agronomic traits in wheat under diverse water regimes. *Euphytica* 213:246
53. Sukumaran S, Dreisigacker S, Lopes M, Chavez P, Reynolds MP. 2015. Genome-wide association study for grain yield and related traits in an elite spring wheat population grown in temperate irrigated environments. *Theoretical and Applied Genetics* 128:353–63
54. Schneider FD, Morsdorf F, Schmid B, Petchey OL, Hueni A, et al. 2017. Mapping functional diversity from remotely sensed morphological and physiological forest traits. *Nature Communication* 8:1441
55. Liebisch F, Kirchgessner N, Schneider D, Walter A, Hund A. 2015. Remote, aerial phenotyping of maize traits with a mobile multi-sensor approach. *Plant Methods* 11:9
56. Zhang L, Niu Y, Zhang H, Han W, Li G, et al. 2019. Maize canopy temperature extracted from UAV thermal and RGB imagery and its application in water stress monitoring. *Frontiers in Plant Science* 10:1270
57. Beć KB, Grabska J, Siesler HW, Huck CW. 2020. Handheld near-infrared spectrometers: where are we heading? *NIR News* 31:28–35
58. Mishra P, Lohumi S, Ahmad Khan H, Nordon A. 2020. Close-range hyperspectral imaging of whole plants for digital phenotyping: recent applications and illumination correction approaches. *Computers and Electronics in Agriculture* 178:105780
59. Langridge P, Reynolds MP. 2015. Genomic tools to assist breeding for drought tolerance. *Current Opinion in Biotechnology* 32:130–35
60. Maccaferri M, El-Feki W, Nazemi G, Salvi S, Canè MA, et al. 2016. Prioritizing quantitative trait loci for root system architecture in tetraploid wheat. *Journal of Experimental Botany* 67:1161–78
61. Mason RE, Addison CK, Babar A, Acuna A, Lozada D, et al. 2018. Diagnostic markers for vernalization and photoperiod loci improve genomic selection for grain yield and spectral reflectance in wheat. *Crop Science* 58:242–52
62. Baret F, Madec S, Irfan K, Lopez J, Comar A, Hemmerlé M, et al. 2018. Leaf-rolling in maize crops: from leaf scoring to canopy-level measurements for phenotyping. *Journal of Experimental Botany* 69:2705–16
63. Ramya P, Singh GP, Jain N, Singh PK, Pandey MK, et al. 2016. Effect of recurrent selection on drought tolerance and related morphophysiological traits in bread wheat. *PLoS ONE* 11:e0156869
64. Shokat S, Sehgal D, Liu F, Singh S. 2020. GWAS analysis of wheat prebreeding germplasm for terminal drought stress using next generation sequencing technology. *Preprints* 2020020272
65. Pinto RS, Lopes MS, Collins NC, Reynolds MP. 2016. Modelling and genetic dissection of staygreen under heat stress. *Theoretical and Applied Genetics* 129:2055–74
66. Gao F, Wen W, Liu J, Rasheed A, Yin G, et al. 2015. Genome-wide linkage mapping of QTL for yield components, plant height and yield-related physiological traits in the Chinese Wheat Cross Zhou 8425B/Chinese Spring. *Frontiers in Plant Science* 6:1099
67. Wang X, Zhang R, Song W, Han L, Liu X, et al. 2019. Dynamic plant height QTL revealed in maize through remote sensing phenotyping using a high throughput unmanned aerial vehicle (UAV). *Scientific Reports* 9:3458

68. Edwards EJ, Still CJ. 2008. Climate, phylogeny and the ecological distribution of C4 grasses. *Ecology Letters* 11:266–76
69. Givnish TJ, Montgomery RA. 2014. Common-garden studies on adaptive radiation of photosynthetic physiology among Hawaiian lobeliads. *Proceedings of the Royal Society B* 281:e20132944
70. Huang X, Wei X, Sang T, Zhao Q, Feng Q, et al. 2010. Genome-wide association studies of 14 agronomic traits in rice landraces. *Nature Genetics* 42:961–67
71. Huang X, Zhao Y, Wei X, Li C, Wang A, et al. 2012. Genome-wide association study of flowering time and grain yield traits in a worldwide collection of rice germplasm. *Nature Genetics* 44:32–39
72. Zhao K, Tung CW, Eizenga GC, Wright MH, Ali ML, et al. 2011. Genome-wide association mapping reveals a rich genetic architecture of complex traits in *Oryza sativa*. *Nature Communications* 2:467
73. Huang H, Deng J, Lan Y, Yang A, Deng X, et al. 2018. A fully convolutional network for weed mapping of unmanned aerial vehicle (UAV) imagery. *PLoS One* 13:e0196302
74. Holman FH, Riche AB, Michalski A, Castle M, Wooster MJ, et al. 2016. High throughput field phenotyping of wheat plant height and growth rate in field plot trials using UAV based remote sensing. *Remote Sensing* 8:1031
75. Liang Z, Pandey P, Stoerger V, Xu Y, Qiu Y, et al. 2018. Conventional and hyperspectral time-series imaging of maize lines widely used in field trials. *GigaScience* 7:gix117
76. Tripodi P, Massa D, Venezia A, Cardi T. 2018. Sensing technologies for precision phenotyping in vegetable crops: current status and future challenges. *Agronomy* 8:57
77. Barnaby JY, Huggins TD, Lee H, McClung AM, Pinson SRM, et al. 2020. Vis/NIR hyperspectral imaging distinguishes sub-population, production environment, and physicochemical grain properties in rice. *Scientific Reports* 10:9284
78. Sun D, Cen H, Weng H, Wan L, Abdalla A, et al. 2019. Using hyperspectral analysis as a potential high throughput phenotyping tool in GWAS for protein content of rice quality. *Plant Methods* 15:54
79. Anderson CB. 2018. Biodiversity monitoring, earth observations and the ecology of scale. *Ecology Letters* 21:1572–85
80. Gamon JA, Wang R, Gholizadeh H, Zutta B, Townsend PA, et al. 2020. Consideration of scale in remote sensing of biodiversity. In *Remote Sensing of Plant Biodiversity*, eds. Cavender-Bares J, Gamon JA, Townsend PA. Cham: Springer. pp. 425–47. [https://doi.org/10.1007/978-3-030-33157-3\\_16](https://doi.org/10.1007/978-3-030-33157-3_16)
81. Hernández-Stefanoni JL, Gallardo-Cruz JA, Meave JA, Rocchini D, Bello-Pineda J, et al. 2012. Modeling  $\alpha$ - and  $\beta$ -diversity in a tropical forest from remotely sensed and spatial data. *International Journal of Applied Earth Observation and Geoinformation* 19:359–68
82. Wong CYS, Gamon JA. 2015. The photochemical reflectance index provides an optical indicator of spring photosynthetic activation in evergreen conifers. *New Phytologist* 206:196–208
83. Chavana-Bryant C, Malhi Y, Wu J, Asner GP, Anastasiou A, et al. 2017. Leaf aging of Amazonian canopy trees as revealed by spectral and physicochemical measurements. *New Phytologist* 214:1049–63
84. Thornley R, Gerard FF, White K, Verhoef A. 2022. Intra-annual taxonomic and phenological drivers of spectral variance in grasslands. *Remote Sensing of Environment* 271:112908
85. Anderegg J, Yu K, Aasen H, Walter A, Liebisch F, et al. 2019. Spectral vegetation indices to track senescence dynamics in diverse wheat germplasm. *Frontiers in Plant Science* 10:1749
86. Pandey NK, Diwakar M. 2020. A review on cloud-based image processing services. In *2020 7th International Conference on Computing for Sustainable Global Development (INDIACom)*, New Delhi, India. pp. 108–12. <https://doi.org/10.23919/INDIACom49435.2020.9083718>
87. Corti M, Marino Gallina P, Cavalli D, Cabassi G. 2017. Hyperspectral imaging of spinach canopy under combined water and nitrogen stress to estimate biomass, water, and nitrogen content. *Biosystems Engineering* 158:38–50
88. Gorelick N, Hancher M, Dixon M, Ilyushchenko S, Thau D, et al. 2017. Google Earth Engine: planetary-scale geospatial analysis for everyone. *Remote Sensing of Environment* 202:18–27
89. Bermudez I, Traverso S, Mellia M, Munafò M. 2013. Exploring the cloud from passive measurements: the Amazon AWS case. In *2013 Proceedings IEEE INFOCOM*. Turin, Italy: IEEE. pp. 230–34. <https://doi.org/10.1109/INFOCOM.2013.6566769>
90. Dede M, Widiawaty MA, Pramulatsih GP, Ismail A, Ati A, et al. 2019. Integration of participatory mapping, crowdsourcing and geographic information system in flood disaster management (case study Ciledug Lor, Cirebon). *Journal of Information Technology and Its Utilization* 2:44–47
91. Kumar L, Mutanga O. 2018. Google Earth Engine applications since inception: usage, trends, and potential. *Remote Sensing* 10:1509
92. Hansen MC, Potapov PV, Moore R, Hancher M, Turubanova SA, et al. 2013. High-resolution global maps of 21st-century forest cover change. *Science* 342:850–53
93. Joshi AR, Dinerstein E, Wikramanayake E, Anderson ML, Olson D, et al. 2016. Tracking changes and preventing loss in critical tiger habitat. *Science Advances* 2:e1501675
94. Moxley JH, Bogomolni A, Hammill MO, Moore KMT, Polito MJ, et al. 2017. Google Haul Out: earth observation imagery and digital aerial surveys in coastal wildlife management and abundance estimation. *BioScience* 67:760–68
95. Dede M, Widiawaty MA. 2020. Utilization EOS Platform as cloud-based GIS to analyze vegetation greenness in Cirebon Regency, Indonesia. *Journal of Information Technology and Its Utilization* 3:1–4
96. Stahl AT, Fremier AK, Heinse L. 2021. Cloud-based environmental monitoring to streamline remote sensing analysis for biologists. *BioScience* 71:1249–60



Copyright: © 2024 by the author(s). Published by Maximum Academic Press, Fayetteville, GA. This article is an open access article distributed under Creative Commons Attribution License (CC BY 4.0), visit <https://creativecommons.org/licenses/by/4.0/>.