

Original Research

Open Access

Multigenerational developmental and skeletal toxicity from benzo[a]pyrene exposure in F0 and F2 medaka: metabolic trade-offs and survival costs

Yinhua Chen¹, Huiju Lin², Jiangang Wang¹, Feilong Li³, Rim EL Amouri¹, Jack Chi-Ho Ip⁴, Wenhua Liu¹ and Jiezhong Mo^{1*}

Received: 11 November 2025

Revised: 16 December 2025

Accepted: 26 December 2025

Published online: 19 January 2026

Abstract

Benzo[a]pyrene is a ubiquitous emerging contaminant; however, whether and how it disrupts fish development and skeletal health across multiple generations, particularly at the metabolomic level, remains unclear. The latent effects of embryonic benzo[a]pyrene exposure at environmentally relevant levels (2.5–80 µg/L) on developmental toxicity and skeletal integrity were assessed over three generations (F0–F2) of medaka. The F1 and F2 offspring were reared in clean water to isolate the inherited impacts. Untargeted metabolomic analysis (Variable Importance in Projection > 1.0, $p < 0.05$ in Student's *t*-test, and ratio ≥ 1.5 or ≤ 0.67) of F0 and F2 embryos identified 233 (86) and 254 (752) differentially abundant metabolites in the F0 (F2) embryos exposed ancestrally to benzo[a]pyrene at 2.5 and 20 µg/L. Benzo[a]pyrene exposure induced multigenerational impairments, including reduced survivorship and hatching success in F0 embryos, altered morphometrics (e.g., reduced body length, smaller eye pit, and larger pericardial cavity) in F0–F1 larvae, and elevated skeletal malformation frequency (e.g., craniofacial and spinal deformities) in F0–F2 larvae. Metabolomic perturbations were linked to four major biological processes: (1) metabolic activation and oxidative stress; (2) disruption of energy metabolism and redox balance, evidenced by a triad of elevated 3-hydroxybutyric acid, oxoglutaric acid, and depleted 2-hydroxyglutarate; (3) dysregulation of cell signaling and developmental programming, including energy crisis (elevated AMP), impaired vascular development (reduced prostaglandin I2), and disrupted efferocytosis; and (4) impairment of neurological and sensory functions. These findings demonstrate that F0 embryonic benzo[a]pyrene exposure induces lasting multigenerational toxicity by disrupting metabolic, signaling, and sensory pathways, ultimately leading to developmental impairment and osteotoxicity. While F2 larvae showed partial morphological recovery, this may represent a survival trade-off, where metabolic resources were redirected from skeletal development to support the formation of essential organs. This study identifies the metabolomic basis for the multigenerational developmental and skeletal toxicity induced by embryonic benzo[a]pyrene exposure.

Keywords: Emerging contaminant, Fish, Multigenerational impact, Skeletal malformation, Metabolome, Biomarker

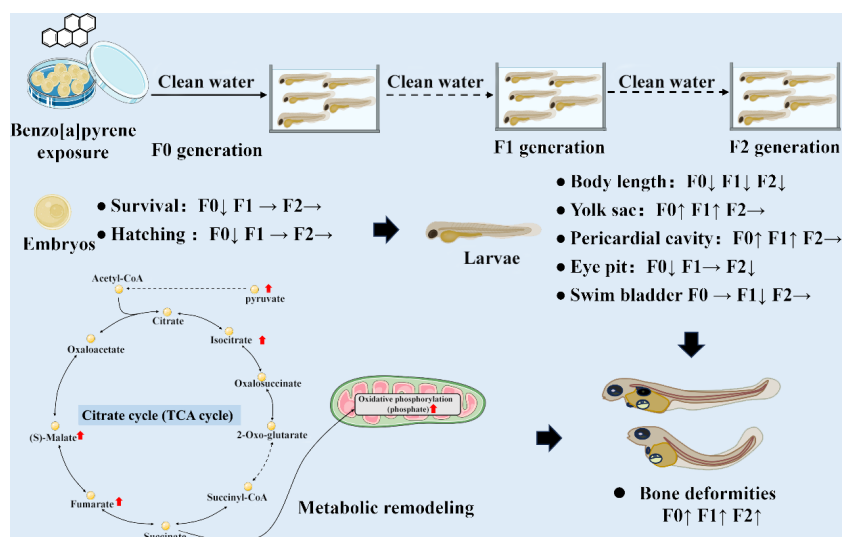
Highlights

- Benzo[a]pyrene exposure induced multigenerational developmental toxicity in medaka.
- Reduced body length and elevated bone deformities were observed in F0–F2 larvae.
- Novel metabolomic data were linked to developmental and skeletal impairment.
- Benzo[a]pyrene exposure disrupted energy metabolism and oxidative balance.
- Energy remodeling and compensation contributed to the developmental recovery.

* Correspondence: Jiezhong Mo (jzhmo@stu.edu.cn)

Full list of author information is available at the end of the article.

Graphical abstract



Introduction

Polycyclic aromatic hydrocarbons are persistent environmental contaminants primarily from anthropogenic activities such as fossil fuel combustion and industrial processes^[1]. Among them, benzo[a]pyrene is a prototypical compound, detected in aquatic environments at concentrations typically ranging from ng/L to sub- μ g/L, and up to mg/L at extreme pollution scenarios^[2,3]. Benzo[a]pyrene exerts its toxicity mainly through metabolic activation. As a potent aryl hydrocarbon receptor agonist, it is converted by cytochrome P450 enzymes into reactive intermediates. This process can lead to DNA adduct formation, mutagenesis, and oxidative stress, magnified as diverse adverse outcomes^[4,5].

The toxicological profile of benzo[a]pyrene is extensive, encompassing carcinogenicity, immunotoxicity, neurotoxicity, and notably, profound developmental and reproductive toxicity^[3,6,7]. In fish models, waterborne or dietary benzo[a]pyrene exposure causes severe developmental defects, including increased mortality, premature or delayed hatching, growth reduction, cardiac dysfunction, pericardial and yolk sac edema, and craniofacial and spinal deformities^[8–14]. Crucially, these effects are not confined to the directly exposed generation. It has been demonstrated that a parental dietary benzo[a]pyrene exposure (10, 114, and 1,012 μ g/g food for 21 d) in zebrafish induced prominent body morphology deformities and premature hatching in the F1 generation, with craniofacial defects emerging prominently in the F2 generation, despite these offspring never being directly exposed^[10]. Ancestral benzo[a]pyrene exposure (1 μ g/L for 21 d) caused bone impairments in larvae and adult male medaka of F1–F3 generations^[11–13].

A key target of benzo[a]pyrene toxicity is the skeletal system. Osteotoxicity, manifested as spinal curvature and craniofacial defects, is a well-documented phenotype in fish exposed to benzo[a]pyrene^[5]. In medaka (*Oryzias latipes*), embryonic exposure to benzo[a]pyrene (0.01, 0.1, and 1 mg/L for 2 h with a nanosecond-pulsed electric field) induces teratogenic effects, including cardiovascular abnormalities and a characteristic curvature of the backbone^[14]. The molecular underpinnings of this osteotoxicity may involve the dysregulation of genes critical for bone formation. For instance, in marbled rockfish (*Sebastes marmoratus*), embryonic

benzo[a]pyrene exposure (0.05, 0.5, and 5 nmol/L) for 7 d caused skeletal deformities and altered the expression of genes involved in osteogenesis^[15]. Transcriptomic analyses in zebrafish further reveal that benzo[a]pyrene exposure disrupts pathways related to extracellular matrix organization and collagen synthesis, essential for skeletal integrity^[16]. Furthermore, embryonic benzo[a]pyrene exposure (0.25, 0.5, 0.6, and 0.8 μ mol/L for 72 h) induces severe morphological and skeletal deformities in zebrafish embryos, delays hatching by structurally altering the zebrafish hatching enzyme, and accumulates primarily in the yolk sac^[17]. At the molecular level, benzo[a]pyrene exposure triggers oxidative stress, leads to the downregulation of critical bone-related genes (e.g., *sox9a*, *spp1/opn*, and *col1a1*), and induces apoptosis in the head, backbone, and tail regions^[17].

While the phenotypic consequences of benzo[a]pyrene exposure are well-characterized, the fundamental metabolic disruptions that drive these multigenerational developmental and osteotoxic effects remain unclear. Transcriptomic studies have provided invaluable insights, identifying dysregulated pathways such as xenobiotic metabolism signaling, NRF2-mediated oxidative stress response, glutathione-mediated detoxification, and arachidonic acid metabolism^[13,14,16,18]. These pathways are intrinsically linked to cellular metabolism, which suggests that benzo[a]pyrene induces a profound metabolic reprogramming. However, a comprehensive metabolomic profile that connects these molecular initiations to the phenotypic endpoints across generations is lacking.

Metabolomics, the systematic study of small-molecule metabolites, offers a powerful tool to decipher the functional outcome of genomic, transcriptomic, and proteomic changes. It can reveal metabolic pathway disruptions—such as in energy metabolism, amino acid synthesis, lipid peroxidation, and antioxidant defense—that are the direct cause of observed toxicity^[19,20].

In this study, it is hypothesized that embryonic exposure to benzo[a]pyrene induces persistent metabolomic perturbations that underlie the observed multigenerational developmental toxicity and osteotoxicity in medaka fish. These perturbations likely involve pathways related to oxidative stress, energy metabolism, inflammation, and bone matrix formation. This metabolomic perspective is critical as it moves beyond the transcriptional potential revealed by

prior studies to directly identify the functional biochemical dysregulations—the altered metabolic fluxes and effector molecules—that are the proximate cause of phenotypic damage. Using medaka as a model, this study aims to: (1) characterize the multigenerational impacts of embryonic benzo[a]pyrene exposure on developmental malformations and skeletal integrity; and (2) employ an untargeted metabolomics approach to identify the key metabolic pathways disrupted in directly exposed (F0) and subsequent (F2) generations. By integrating phenotypic data with metabolomic profiles, this study elucidates the mechanistic metabolic links between early-life benzo[a]pyrene exposure and adverse health outcomes that persist across generations, providing a deeper understanding of its long-term ecological risks.

Materials and methods

Fish husbandry conditions

Sexually mature medaka (*Oryzias latipes*, 4 months old) were maintained under conditions approved by the Animal Ethics Committee of Shantou University (Shantou, China). Embryos were collected within 30 min of the light cycle beginning. Each of 15 pairs of fish was randomly allocated to 10 cube tanks (30 cm × 30 cm × 30 cm; 20-L volume) containing filtered tap water. Stock fish were maintained at optimal conditions (26 ± 0.5 °C, pH 7.3 ± 0.1, 7.2 ± 0.2 mg O₂ L⁻¹, and a light : dark cycle of 14:10), and fed *Artemia nauplii* twice per day^[7,21].

Chemical treatment and multigenerational experimental design

Collected embryos were rinsed, cleaned, and sorted (stages 8–9) before being distributed into glass Petri dishes (diameter = 7 cm). Each dish contained benzo[a]pyrene exposure solutions (CAS No. 57–63–6; purity ≥ 98%; dissolved in DMSO) at nominal concentrations ranging from 2.5 to 80 µg/L, with 30 embryos per replicate and four replicates per treatment. The solvent control (0.4% DMSO) contained no detectable benzo[a]pyrene, while the measured exposure concentrations were 2.2 ± 0.4, 17.0 ± 5.0, and 73.0 ± 10.0 µg/L for the respective treatment groups^[7]. Embryos were incubated at 26 ± 0.5 °C, and the exposure solution was renewed daily. Owing to the limited volume of the exposure solution, benzo[a]pyrene concentrations were not ascertained during the 24-h interval before medium renewal.

The embryonic exposure period concluded near the time of hatching at 8 d post fertilization. From day 9 post-fertilization onwards, the benzo[a]pyrene solution was replaced with a clean embryonic culture medium. Upon hatching, the F0 generation larvae were transferred to clean water, and raised to adulthood to produce the F1 and F2 generations^[7,12], and 15 pairs of adult medaka were randomly assigned and reared in each replicate tank. For F0–F2 embryos, mortality and hatching rates were recorded twice daily over a 20-d period. However, mortality rates were not recorded over the course of the multigenerational studies. Newly hatched larvae from each generation were observed and imaged using an inverted stereo microscope (Zeiss Stemi 305, Germany). Morphometric analysis was performed on these images using ImageJ software to measure total body length (mm), eye pit area (mm²), yolk sac area (mm²), pericardial cavity area (mm²), and swim bladder area (mm²). A larva was scored positive for a craniofacial deformity if the head morphology was abnormal or the head and trunk were not aligned, for a spinal deformity if the trunk exhibited clear curvature (e.g., lordosis, kyphosis, or scoliosis), and for a fin/tail deformity if the fin structures showed a distinct bend or malformation. To minimize bias, all morphometric and deformity

assessments were conducted by the same observer who was blinded to the treatment groups.

Untargeted metabolomics analysis

Sample preparation and metabolite extraction

To ensure that the observed metabolic disruptions specifically reflect mechanisms underlying developmental defects such as spinal curvature, rather than acute, nonspecific stress responses or lethality, metabolomic profiling was only conducted on F0 and F2 embryos ancestrally exposed to environmentally relevant, sublethal concentrations of benzo[a]pyrene (2.5 and 20 µg/L). F0 medaka embryos, with or without benzo[a]pyrene exposure, were sampled on day 8 and stored at –20 °C for metabolomics analysis. Medaka embryos (100 embryos × four replicates) were homogenized in 500 µL of 80% methanol (prechilled to –20 °C) using a grinder with steel beads. A small aliquot (20 µL) of the homogenate was retained, dried under vacuum, and redissolved. The protein content of this aliquot was then quantified using a BCA assay, which was used for subsequent normalization of metabolite abundances. The homogenate was incubated at –20 °C for 30 min to precipitate proteins and subsequently centrifuged at 20,000 × g for 10 min at 4 °C. The supernatant was then recentrifuged for 5 min under the same conditions. The final supernatant was collected for analysis. A quality control (QC) sample was generated by pooling equal aliquots of all sample supernatants.

Liquid chromatography-high resolution mass spectrometry

Metabolite separation was performed using UPLC, followed by detection on a TripleTOF 6600 mass spectrometer equipped with an electrospray ionization source operating in both positive and negative ionization modes. Key source parameters included temperature, 500 °C; curtain gas, 30 psi; sheath and auxiliary gas, 60 psi each; and ion spray voltage, ±5,000 V/–4,500 V. Data were acquired in an information-dependent acquisition mode. Each cycle consisted of a high-resolution TOF-MS survey scan (m/z 60–1,200, 150 ms) followed by MS/MS scans on the top 12 most intense ions (m/z 25–1,200, 30 ms/scan), with a 4-s dynamic exclusion.

Data processing and metabolite annotation

Raw data files were converted to mzML format and processed using XCMS and MetaboAnnotation toolbox in R for peak picking, alignment, and annotation of adducts and isotopes. Metabolites were annotated by matching accurate mass (with a tolerance of < 10 ppm) against the KEGG and HMDB databases. Putative identifications were further validated by isotopic distribution patterns and comparison to an in-house MS/MS spectral library. A feature intensity matrix was generated for statistical analysis.

Quality control and statistical analysis

Data quality was ensured by rigorous preprocessing: metabolic features (with confirmed MS/MS spectra) not detected in > 50% of QC samples or > 80% of biological samples were removed. Missing values were imputed using the k-nearest neighbor algorithm. Signal intensity drift was corrected using the MetNormalizer algorithm on the QC samples. Features with a relative standard deviation > 30% in the QC samples were excluded. Differential analysis was performed using Student's *t*-tests with FDR adjustment for multiple comparisons, and supervised Partial Least Squares-Discriminant Analysis (PLS-DA) was employed for group separation. A permutation test was executed 200 times to assess the risk of overfitting for the applied model (Supplementary Figs S1–S10 & Supplementary Table S1). Features with a Variable Importance in Projection (VIP) score > 1.0, *p* < 0.05 in the Student's *t*-test, and ratio ≥ 1.5 or ≤ 0.67 were considered as differentially abundant metabolites (DAMs). The DAMs identified in

both modes were combined and subjected to KEGG pathway enrichment analysis.

Statistical analysis

Statistical analyses were performed with GraphPad Prism 10.4. Data are expressed as mean $\pm$ SEM from four independent replicates ($n = 4$). Differences between the solvent control and benzo[a]pyrene treatments across F0–F2 generations were assessed by two-way ANOVA followed by Tukey's post hoc test (for morphometric data) or a χ^2 test (for survival, hatching, and deformity data). Results were considered statistically significant at $*p < 0.05$.

Results

Survivorship and hatching success of F0–F2 medaka embryos

In the F0 generation, benzo[a]pyrene exposure significantly suppressed embryonic development, delayed hatching, and reduced hatching success in medaka. A profound, concentration-dependent increase in mortality was observed, becoming significant from day 10 (80 $\mu\text{g/L}$) and day 13 (40 $\mu\text{g/L}$) onward ($p < 0.05$, χ^2 test; Fig. 1a). At 20 dpf, the cumulative survival rate dropped from 92.5% $\pm$ 1.8% for the F0 control to 90.0% $\pm$ 1.2%, 89.3% $\pm$ 1.4%, and 13.3% $\pm$ 4.9% for the 2.5, 20, and 80 $\mu\text{g/L}$ treatments, respectively. In contrast, descending F1 and F2 generations from exposed ancestors showed non-significant reduction in survivorship at 2.5, 20, and 80 $\mu\text{g/L}$ concentrations compared to the control (Fig. 1b, c).

In the F0 generation, benzo[a]pyrene exposure induced a delay in the onset of hatching, with a 2-d delay at concentrations ≤ 10 $\mu\text{g/L}$,

and a 3-d delay at higher concentrations compared to the solvent control (Fig. 2a). At 20 dpf, the cumulative hatching rate decreased from 92.5% $\pm$ 2.1% for the F0 control to 90.0% $\pm$ 1.2%, 87.5% $\pm$ 2.0%, and 12.5% $\pm$ 5.5% for the 2.5, 20, and 80 $\mu\text{g/L}$ treatments, respectively. Significant reductions ($p < 0.05$, χ^2 test) in cumulative hatching rates were observed at 40 and 80 $\mu\text{g/L}$. While a 1-d hatching delay persisted in the F1 generation at 20 and 80 $\mu\text{g/L}$, the reductions in hatching success were largely recovered (Fig. 2b). In the F2 generation, both hatching timing and success were comparable to the control (Fig. 2c).

Developmental toxicity and osteotoxicity in F0–F2 medaka larvae

Ancestral benzo[a]pyrene exposure induced multigenerational developmental and osteotoxic effects in medaka F0–F2 larvae. Significant, concentration-specific alterations were observed: body length was reduced in the 20 $\mu\text{g/L}$ treatment across all three generations (4.608 $\pm$ 0.061 mm compared to 4.751 $\pm$ 0.039 mm in F0 control; 4.360 $\pm$ 0.043 mm compared to 4.627 $\pm$ 0.026 mm in F1 control; 4.260 $\pm$ 0.032 mm compared to 4.352 $\pm$ 0.021 mm in F2 control). Eye pit size decreased in F0 (0.0883 $\pm$ 0.0010, 0.0833 $\pm$ 0.0011, and 0.0832 $\pm$ 0.0010 mm² for 2.5, 20, and 80 $\mu\text{g/L}$ treatment compared to 0.0919 $\pm$ 0.0011 mm² for the control, respectively) and F2 (0.0918 $\pm$ 0.0010 mm² for the 20 $\mu\text{g/L}$ treatment compared to 0.0952 $\pm$ 0.0009 mm² for the control) larvae. Swim bladder size was reduced only in F1 larvae (0.0718 $\pm$ 0.0082 and 0.0715 $\pm$ 0.0074 mm² for the 20 and 80 $\mu\text{g/L}$ treatment compared to 0.0995 $\pm$ 0.0083 mm² for the control, respectively). In contrast, yolk sac edema was induced in all F0 treatments and in F1 larvae (0.417 $\pm$ 0.010 and 0.436 $\pm$ 0.011 mm² for the 20 and 80 $\mu\text{g/L}$

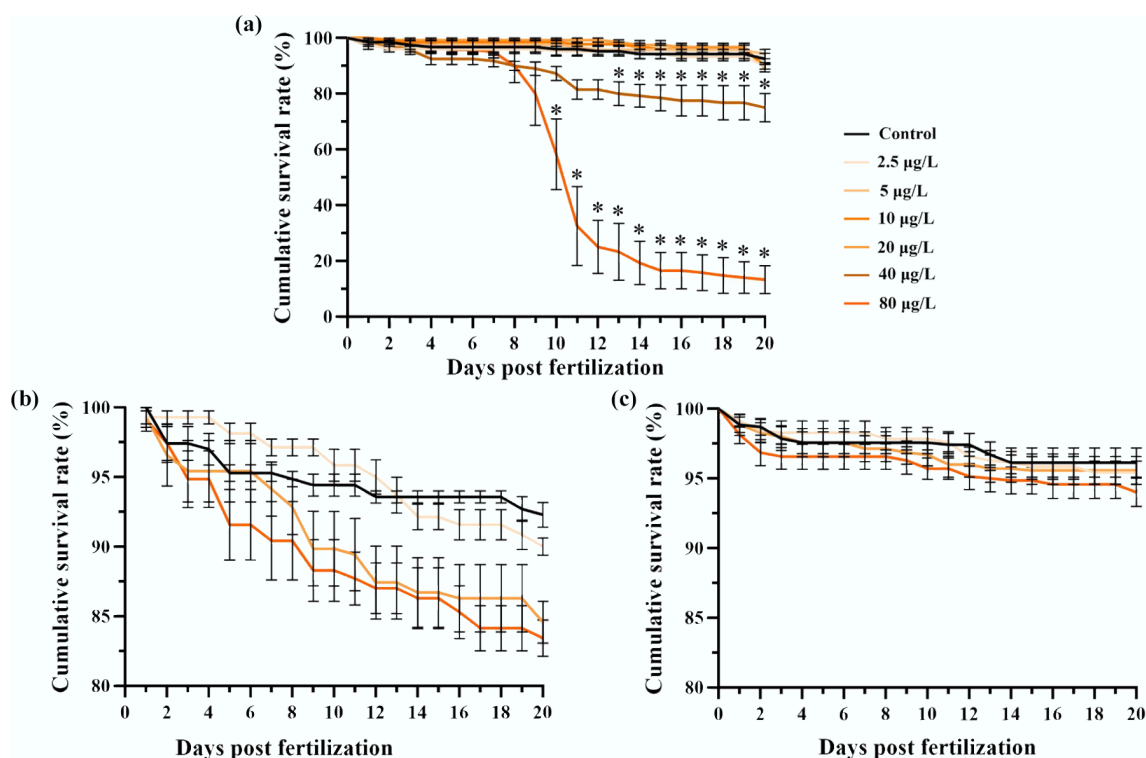


Fig. 1 Multigenerational survivorship of medaka embryos affected by F0 embryonic benzo[a]pyrene exposure. The exposure lasted for 8 d, after which all individuals of F0–F2 generations were reared in clean water. Cumulative survival rates are shown for the (a) F0, (b) F1, and (c) F2 embryos with or without ancestral benzo[a]pyrene exposure ($n = 4$; data are presented as mean $\pm$ SEM). A comparison of the mean values is provided, and * indicates $p < 0.05$, according to a χ^2 test.

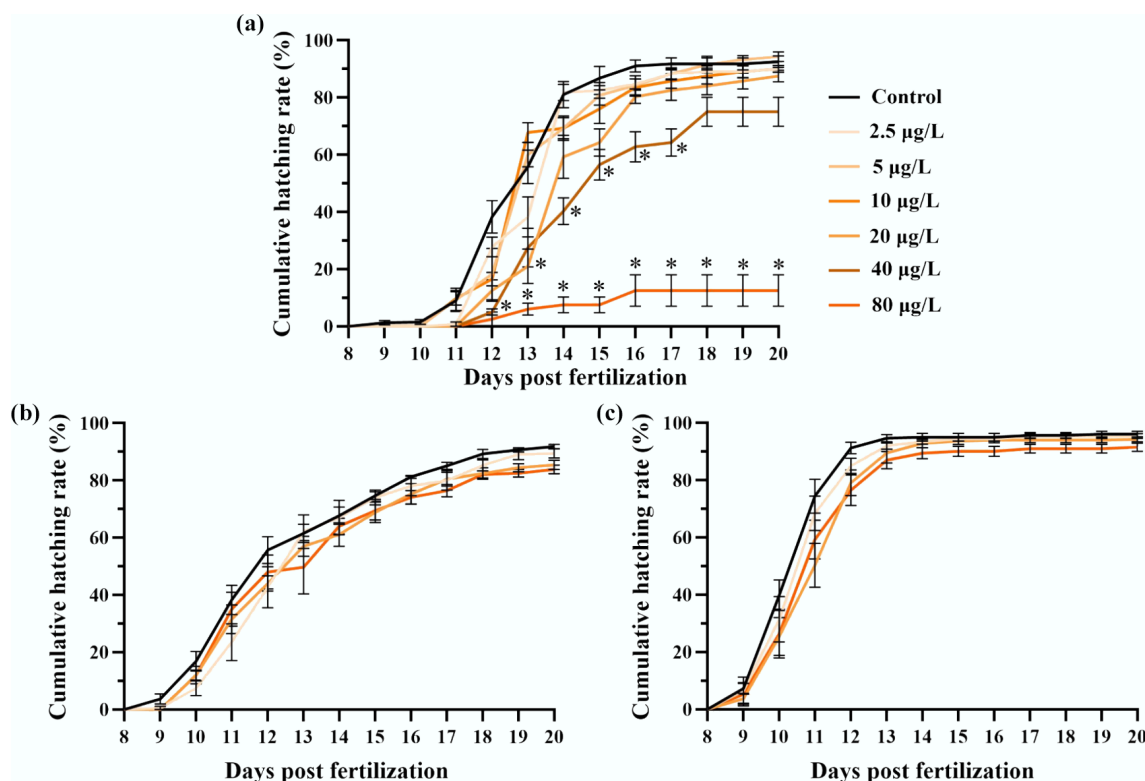


Fig. 2 Multigenerational hatching success of medaka embryos affected by F0 embryonic benzo[a]pyrene exposure. The exposure lasted for 8 d, after which all individuals of F0–F2 generations were reared in clean water. Cumulative hatching rates are presented for the (a) F0, (b) F1, and (c) F2 medaka embryos with or without ancestral benzo[a]pyrene exposure ($n = 4$; data are presented as mean $\pm$ SEM). A comparison of the mean values is provided, * indicates $p < 0.05$, according to a χ^2 test.

treatment compared to $0.385 \pm 0.009 \text{ mm}^2$ for control, respectively), while pericardial cavity size was significantly larger in F0 (0.0514 ± 0.0011 and $0.0527 \pm 0.0012 \text{ mm}^2$ for the 20 and $80 \mu\text{g/L}$ treatment compared to $0.0451 \pm 0.0008 \text{ mm}^2$ for the control, respectively) and F1 (0.0462 ± 0.0012 , 0.0551 ± 0.0014 , and $0.0531 \pm 0.0013 \text{ mm}^2$ for the 2.5, 20, and $80 \mu\text{g/L}$ compared to $0.0428 \pm 0.0009 \text{ mm}^2$ for the control, respectively) treatments ($p < 0.05$, two-way ANOVA; Fig. 3a–e).

Ancestral benzo[a]pyrene exposure induced body deformities—including craniofacial, spinal, and fin deformities—in F0–F2 larvae (Fig. 4). In the F0 generation, craniofacial deformity (spinal deformity) elevated significantly ($p < 0.05$, χ^2 test) from $39.9\% \pm 5.1\%$ ($22.1\% \pm 4.0\%$) for the control to $51.1\% \pm 5.1\%$ ($36.9\% \pm 6.1\%$), $68.1\% \pm 5.9\%$ ($48.2\% \pm 8.1\%$), and $69.2\% \pm 7.8\%$ ($53.3\% \pm 4.9\%$) for the 2.5, 20, and $80 \mu\text{g/L}$ treatments, respectively. These effects persisted transgenerationally, and significantly higher incidences of spinal curvature were observed in the F1 and F2 generations at 20 and $80 \mu\text{g/L}$ ($43.6\% \pm 5.8\%$ and $50.1\% \pm 5.0\%$ compared to $24.5\% \pm 5.9\%$ for F1 control; $23.6\% \pm 3.3\%$ and $23.0\% \pm 3.6\%$ compared to $10.7\% \pm 3.1\%$ for F2 control, respectively), while craniofacial deformity persisted at $20 \mu\text{g/L}$ ($68.2\% \pm 3.8\%$ compared to $52.7\% \pm 1.9\%$ for F1 control; $36.9\% \pm 2.3\%$ compared to $20.2\% \pm 2.2\%$ for F2 control). In contrast, fin deformity was only significantly increased at $40 \mu\text{g/L}$ in the F0 generation ($21.1\% \pm 4.8\%$ compared to $11.0\% \pm 2.8\%$ for F0 control).

Metabolomic alterations in F0 and F2 embryos

Metabolomic analysis of F0 and F2 embryos demonstrated good stability, as shown by the quality control total ion chromatograms (Supplementary Figs S1, S2). We confidently identified 812 (positive

mode) and 736 (negative mode) metabolites in F0 embryos, and 1,051 (positive) and 1,041 (negative) in F2 embryos. In principal component analysis (PCA), PC1 and PC2 explain 32% and 20.9% of the total variance in metabolites detected for the F0 embryos, respectively. By contrast, PC1 and PC2 explain 50.1% and 10.2% of the total variance in metabolites detected for the F2 embryos, respectively (Supplementary Figs S3, S4).

The high-quality metabolomic data are evidenced by good repeatability and strong correlation of QC samples (Supplementary Figs S3–S6). In PCA score plots, replicates were clustered by treatment in both F0 and F2 generations. A clear separation was observed between the control and the 2.5 and $20 \mu\text{g/L}$ treatments in F0 embryos. In contrast, F2 profiles showed proximity between the control and 2.5 $\mu\text{g/L}$ group, while the $20 \mu\text{g/L}$ treatment remained distinct. OPLS-DA models for F0 and F2 generations showed a clear separation in metabolomic profiles between control and benzo[a]pyrene treatment groups (Supplementary Figs S7–S10), with high model reliability (R^2 : 0.994–0.999, Q^2 : 0.644–0.98; Supplementary Table S1). Based on a threshold of $\text{VIP} > 1.0$, $p < 0.05$, and a fold change ≥ 1.5 or ≤ 0.67 , 233 and 254 DAMs were identified in F0 embryos exposed to 2.5 and $20 \mu\text{g/L}$ benzo[a]pyrene, respectively (Supplementary Fig. S11). In the F2 generation, ancestral exposure resulted in 86 and 752 DAMs at the same concentrations (Supplementary Fig. S12). Notably, 42 DAMs were shared between the $20 \mu\text{g/L}$ treatments across both generations (Fig. 5).

For each generation, all DAMs from positive and negative ionization modes were pooled for KEGG pathway analysis. This revealed distinct multigenerational disruption: in the F0 generation, 20 and 26 pathways were significantly ($p < 0.05$) altered at 2.5 and $20 \mu\text{g/L}$

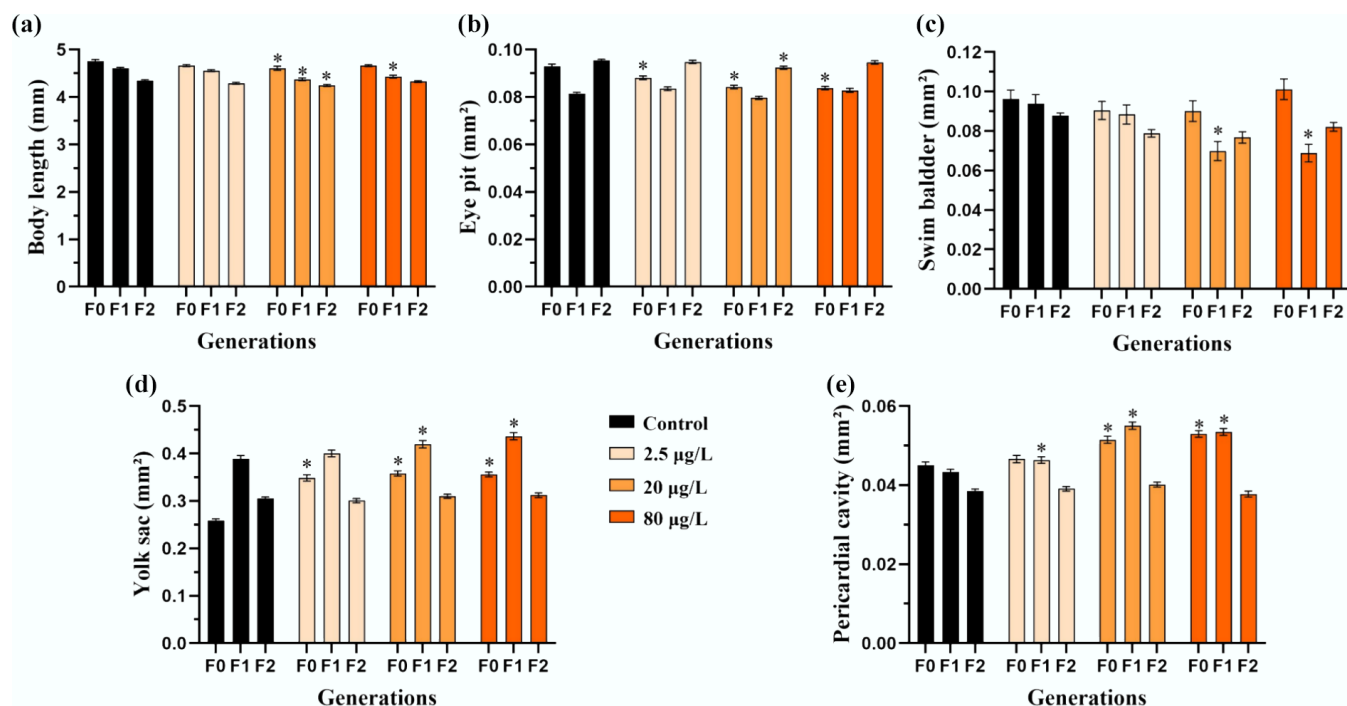


Fig. 3 Multigenerational developmental toxicity of freshly hatched medaka larvae induced by F0 embryonic benzo[a]pyrene exposure. Morphometric parameters including (a) total body length, (b) eye pit, (c) swim bladder, (d) yolk sac, and (e) pericardial cavity are presented for freshly hatched larvae from the F0–F2 generations ($n = 4$; data are presented as mean $\pm$ SEM). A comparison between each benzo[a]pyrene treatment and the corresponding control is provided, and * indicates $p < 0.05$, according to a two-way ANOVA followed by Tukey's *post hoc* test.

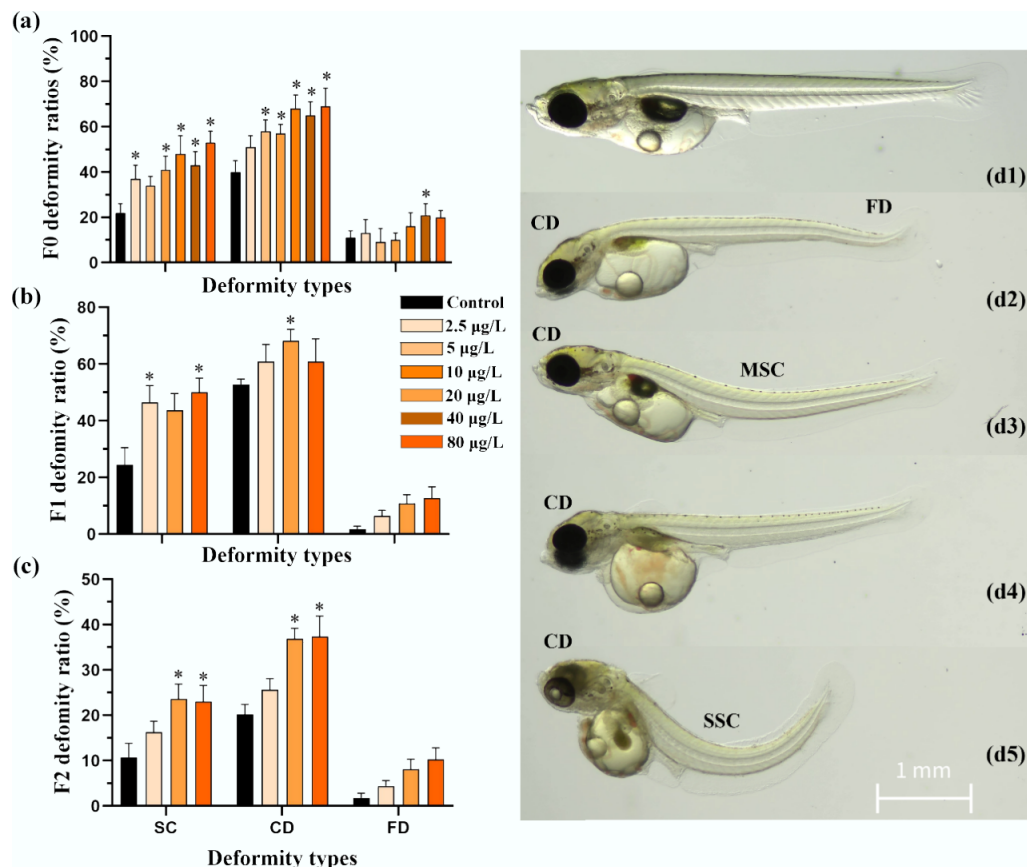


Fig. 4 Multigenerational osteotoxicity of medaka freshly hatched larvae induced by F0 embryonic benzo[a]pyrene exposure. Skeletal deformities in craniofacial, trunk, and tail fin are presented for medaka freshly hatched larvae from the (a) F0, (b) F1, and (c) F2 generations with and without ancestral benzo[a]pyrene exposure ($n = 4$; data are presented as mean $\pm$ SEM). A comparison of the mean values is provided, and * indicates $p < 0.05$, according to a χ^2 test. Representative images of control fish with (d1) normal body shape, and (d2–d5) larvae with craniofacial deformity (CD), spinal curvature (mild and severe spinal curvature are labeled using MSC and SSC, respectively), and fin deformity (FD) in the benzo[a]pyrene treatment groups are shown.



Fig. 5 A heat map showing alterations in metabolomic profiles in F0 and F2 medaka embryos following F0 embryonic benzo[a]pyrene exposure. A total of 42 differentially abundant metabolites are shared by the 20 µg/L benzo[a]pyrene treatment of F0 and F2 generations, alongside their corresponding profiles in the 2.5 µg/L benzo[a]pyrene treatment of F0 and F2 embryos. The column from left to right represents F0 2.5 µg/L treatment, F0 20 µg/L treatment, F2 2.5 µg/L treatment, and F2 20 µg/L treatment, respectively. 42DPMM: 42. 2-(4,4-Difluoro-1-piperidinyl)-6-methoxy-N-[1-(1-methylethyl)-4-piperidinyl]-7-[3-(1-pyrrolidinyl)propoxy]-4-quinazolinamine.

benzo[a]pyrene, respectively (Supplementary Tables S2, S3). These pathways spanned four key functional categories: (1) detoxification (e.g., ABC transporters); (2) metabolic homeostasis (e.g., taurine and hypotaurine, purine, and pyrimidine metabolism); (3) cell signaling and development (e.g., VEGF, PI3K-Akt, mTOR, FoxO, and cAMP signaling pathways); and (4) neurological and sensory functions

(e.g., neuroactive ligand-receptor interaction, GABAergic and serotonergic synapses). Conversely, the F2 generation showed a more pronounced concentration-response, with only three pathways altered at 2.5 µg/L compared to 71 at 20 µg/L (Table 1; Supplementary Tables S4–S6). A notable feature in F2 was the significant enrichment of pathways governing oxidative balance, including the

Table 1 Significantly enriched KEGG metabolic pathways in F2 medaka embryos ancestrally exposed to benzo[a]pyrene at 20 µg/L

Pathway	Level 2	Pathway ID	p-value	Up-DAMs	Down-DAMs
Central carbon metabolism in cancer	Cancer: overview	ko05230	1.16E-15	Pyruvic acid; L-Alanine; Serine; Fumaric acid; Leucine; Isocitric acid; Isoleucine; Methionine; Malic acid; L-Arginine; L-Tyrosine; L-Tryptophan; Lactic acid	Glutamine
Glycine, serine and threonine metabolism	Amino acid metabolism	ko00260	7.26E-10	Pyruvaldehyde; Glyoxylic acid; Pyruvic acid; Serine; Phosphoserine; Sarcosine; Choline; Guanidoacetic acid; Ectoine; L-Tryptophan	Glyceric acid
D-Amino acid metabolism	Metabolism of other amino acids	ko00470	4.32E-06	Pyruvic acid; L-Alanine; Serine; 5-Aminopentanoic acid; Ornithine; Lysine; Methionine; L-Arginine	Glutamine
Efferocytosis	Transport and catabolism	ko04148	3.25E-05	Ornithine; Methionine; L-Arginine; LysoPC (P-18:1(9Z)/0:0); LysoPC(P-18:0/0:0); LysoPC(18:3(6Z,9Z,12Z)/0:0); LysoPC(22:6(4Z,7Z,10Z,13Z,16Z,19Z)/0:0); Lactic acid	–
Arginine and proline metabolism	Amino acid metabolism	ko00330	4.47E-05	Glyoxylic acid; Pyruvic acid; 5-Aminopentanoic acid; Sarcosine; Creatinine; Guanidoacetic acid; Ornithine; L-Arginine	–
Tyrosine metabolism	Amino acid metabolism	ko00350	8.84E-05	Pyruvic acid; Fumaric acid; Beta-Tyrosine; 4-Hydroxycinnamic acid; 5,6-Dihydroxyindole; L-Tyrosine; DOPA	Homovanillic acid
Glyoxylate and dicarboxylate metabolism	Carbohydrate metabolism	ko00630	0.000165	Glyoxylic acid; Pyruvic acid; Serine; Isocitric acid; Malic acid	Glyceric acid; Glutamine
Cysteine and methionine metabolism	Amino acid metabolism	ko00270	0.000242	Pyruvic acid; L-Alanine; Serine; Phosphoserine; 5-Methylthioadenosine; Methionine; S-Adenosylhomocysteine	–
Pyruvate metabolism	Carbohydrate metabolism	ko00620	0.000273	Pyruvaldehyde; Pyruvic acid; Lactic acid; Fumaric acid; Malic acid	–
Citrate cycle (TCA cycle)	Carbohydrate metabolism	ko00020	0.000434	Pyruvic acid; Fumaric acid; Isocitric acid; Malic acid	–
Arginine biosynthesis	Amino acid metabolism	ko00220	0.000759	Fumaric acid; Ornithine; L-Arginine	Glutamine
Valine, leucine and isoleucine biosynthesis	Amino acid metabolism	ko00290	0.000759	Pyruvic acid; alpha-Ketoisovaleric acid; Leucine; Isoleucine	–
Alanine, aspartate and glutamate metabolism	Amino acid metabolism	ko00250	0.001635	Pyruvic acid; L-Alanine; Fumaric acid	Glutamine
Lysine degradation	Amino acid metabolism	ko00310	0.003631	5-Aminopentanoic acid; Glutaric acid; L-Pipecolic acid; Lysine; N6-Acetyl-L-lysine	–
Valine, leucine and isoleucine degradation	Amino acid metabolism	ko00280	0.007321	alpha-Ketoisovaleric acid; 3-Hydroxyisovaleric acid; Leucine; Isoleucine	–
Oxidative phosphorylation	Energy metabolism	ko00190	0.019938	Phosphate; Fumaric acid	–

sulfur relay system, thiamine metabolism, and sulfur metabolism. Multiple metabolomic pathways related to energy metabolism were identified in the 20 µg/L treatment of the F2 generation. Cross-generational analysis identified a core signature of 10 pathways consistently disrupted by the 20 µg/L treatment in both F0 and F2 generations, including mTOR signaling, purine metabolism, ABC transporters, and neuroactive ligand-receptor interaction, underscoring their role as persistent targets of toxicity (Supplementary Tables S3, S5).

Discussion

F0 embryonic exposure to environmental benzo[a]pyrene induced multigenerational developmental toxicity and transgenerational osteotoxicity in medaka. The effects include reduced survivorship and hatching success in the F0 generation, alterations in morphometric parameters (e.g., body length, eye pit, swim bladder, yolk sac, and pericardial cavity) in the F0–F1 generations, and an increased frequency of skeletal malformations (e.g., craniofacial and spinal deformities) in the F0–F2 generations. Novel metabolomic data from F0 and F2 embryos linked these phenotypic changes to disruptions in four major biological processes: (1) metabolic activation and initial stress; (2) energy metabolism and oxidative balance; (3) cell signaling and developmental programming; and (4) neurological and sensory functions.

Metabolic activation and initial stress

The overall toxicity of the procarcinogen benzo[a]pyrene is determined by the critical balance between its metabolic activation and detoxification^[1,4]. Upon cellular entry, benzo[a]pyrene is primarily acti-

vated via the drug metabolism–other enzymes pathway by enzymes like cytochrome P450 (e.g., CYP1A1) and epoxide hydrolase. This metabolism generates the highly reactive, ultimate carcinogen BPDE and ROS, leading to genetic damage, oxidative stress, and epigenetic modifications^[5]. The developmental toxicity observed in F0 embryos and larvae following embryonic benzo[a]pyrene exposure was attributable to this molecular and cellular damage, which is supported by benzo[a]pyrene-induced cellular dysfunction and apoptosis in skeletal tissues of fish embryos and larvae reported previously^[14,17,22,23]. Concurrently, ABC transporters function as cellular efflux pumps to expel benzo[a]pyrene and its metabolites, serving as a key defense mechanism, and their efficiency may modulate the intracellular concentration of these compounds^[24–26]. An overload or inhibition of this system consequently exacerbates all downstream toxic effects. The resulting DNA adducts and oxidative damage can induce genetic mutations, aberrant epigenetic modifications, and cell death in embryos^[5,14,17], forming the probable molecular basis for the observed embryonic lethality, delayed development, reduced hatching success, and organ dysmorphogenesis—such as craniofacial deformity, spinal curvature, and fin deformity—in the F0 generation.

Supporting this mechanism, metabolomic analysis revealed enrichment of the drug metabolism-other enzymes and ABC transporters pathways in F0 embryos, indicating active benzo[a]pyrene metabolism and detoxification after exposure. Notably, in the unexposed F2 generation, ABC transporters remained significantly enriched, suggesting their potential role in transporting substances across pericellular membranes in a transgenerational context. While survivorship and development in the F1–F2 generations were comparable to the control levels, alterations in morphometric parameters were largely present in F1 larvae and mostly alleviated

in F2 larvae. These findings partially align with previous work^[10], where dietary benzo[a]pyrene administration to F0 adult zebrafish induced multigenerational toxicity, albeit with effects manifesting differently across generations. A key distinction in the present study is the persistence of shortened body length, craniofacial deformity, and spinal curvature into the F1–F2 generations. This finding on transgenerational osteotoxicity in freshly hatched medaka larvae is novel and distinct from the transgenerational vertebral deformity reported in older (17 d post-hatch) F1–F3 medaka larvae descended from benzo[a]pyrene-exposed ancestors^[11,12].

Energy metabolism and oxidative balance

Energy metabolism and oxidative balance are fundamental to successful embryo development, as they fuel and protect the complex process of morphogenesis^[27]. The embryo, dependent on a finite yolk supply, requires efficient energy production through glycolysis and beta-oxidation to generate the necessary ATP for cell division, tissue differentiation, and morphogenesis^[28,29]. This high metabolic rate inherently produces ROS, which in controlled amounts act as vital developmental signals but in excess cause oxidative stress, damaging DNA, proteins, and lipids^[27,28]. Such disruption can lead to failed organ formation, skeletal deformities, and improper fin development, making the harmonious interplay between energy production and antioxidant defense essential for accurate execution of the genetic blueprint^[29,30].

In F0 embryos, benzo[a]pyrene exposure induced widespread metabolic dysregulation, directly impairing core pathways including C5-branched dibasic acid metabolism, butanoate metabolism, and purine and pyrimidine metabolism. A characteristic triad of metabolite changes—elevated 3-hydroxybutyric acid, increased oxoglutaric acid (α -KG), and decreased 2-hydroxyglutarate (2-HG)—was identified as a core mechanism of toxicity. This triad may collectively disrupt energy metabolism, redox balance, and epigenetic regulation^[20,31]. The rise in 3-hydroxybutyric acid possibly suggests a compensatory shift toward ketogenesis, while the high α -KG/2-HG

ratio possibly activates α -KG-dependent dioxygenases^[32,33]. This may lead to the suppression of HIF-1 α , disrupting hypoxic signaling^[34], and cause epigenetic dysregulation, corrupting the genetic blueprint for morphogenesis and manifesting as severe body malformations^[20,35,36]. Indeed, benzo[a]pyrene exposure has been shown to induce both global and gene-specific DNA hypomethylation in developing zebrafish embryos^[37]. Most recently, it was found to alter histone methylation and acetylation patterns in *twist*-positive and *col10a1*-positive osteoblasts of medaka^[38]. Building on these epigenetic findings, a hypothetical model is proposed that links metabolomic dysregulation (e.g., an altered α -KG/2-HG ratio) to the observed spinal curvature and reduced body length (Supplementary Fig. S13). Further experimental validation is required to confirm the mechanistic role of this pathway.

Concurrently, the significant dysregulation of purine and pyrimidine metabolism, evidenced by an imbalance in key nucleotides, directly impedes embryonic development^[39]. Benzo[a]pyrene-induced DNA damage activates repair systems that massively consume the nucleotide pool, while oxidative stress likely disrupts both *de novo* and salvage synthesis pathways^[40,41]. During rapid embryonic cell division, this insufficient nucleotide supply may impede genome replication and transcription, leading to arrested cell division, which manifests as developmental delay, reduced body size, and organ hypoplasia^[42]. Furthermore, reduced L-arginine content may disrupt the vital nitric oxide signaling pathway. The resulting nitric oxide deficiency may impair cardiovascular development, neural communication, and angiogenesis, contributing to the observed axial and skeletal malformations^[36,43,44].

In the unexposed F2 generation, broader metabolic remodeling and compensatory responses were observed (Table 1 & Fig. 6). Pathways including the citrate cycle, oxidative phosphorylation, and multiple amino acid metabolisms were dysregulated, with key metabolites like pyruvic acid, isocitric acid, α -KG, fumaric acid, malic acid, lactic acid, and various amino acids (L-alanine, serine, leucine, isoleucine, methionine, lysine, D-valine, D-phenylalanine, L-arginine,

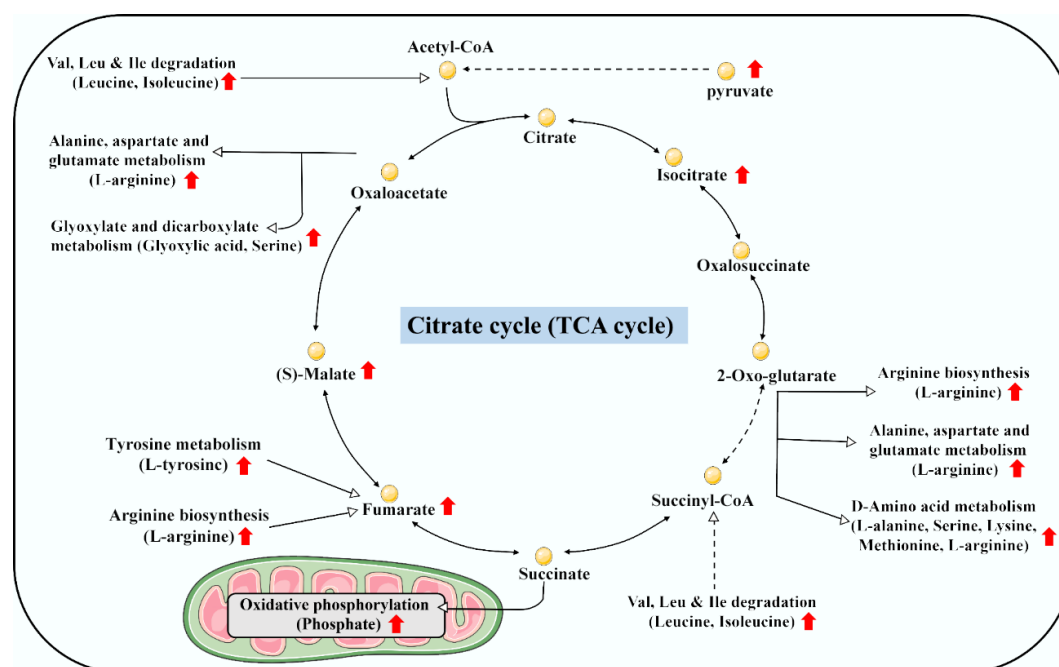


Fig. 6 Schematic diagram showing an overwhelming elevation of energy metabolism and supply in F2 medaka embryos descended from F0 ancestors with embryonic exposure to benzo[a]pyrene at 20 μ g/L.

L-tyrosine, and L-tryptophan) being exclusively elevated. This pattern suggests an overwhelming compensatory regulation aimed at boosting energy production and biosynthetic precursor supply. The accumulation of taurine further supports this response, possibly by enhancing antioxidant defense and cellular osmotic balance^[45,46]. This metabolic compensation underpins the observed recovery in phenotypic endpoints such as survivorship, hatching success, and most morphometric parameters.

Despite this general recovery, skeletal deformities persisted transgenerationally. Based on the F0 and F2 embryo metabolomic data, the massive metabolic shift in F2 larvae might not represent a successful compensatory regulation but rather a maladaptive establishment of a pathological steady state, aligning with the 'energy debt' hypothesis^[47,48]. This reprogramming might fail to rescue skeletal development when resources are misallocated, where the reversal of acylcarnitine profiles from suppression in F0 to sharp elevation in F2 indicates a high-cost shift toward catabolic energy mobilization, thereby continuously depriving skeletogenesis of necessary anabolic substrates. Importantly, these metabolic changes possibly operated within an immutable epigenetic landscape: the normalization of 2-hydroxyglutarate in F2 alongside elevated 5-methylcytosine seemingly supports a 'hit-and-run' mechanism^[49], where the initial exposure durably locked in erroneous epigenetic programs (e.g., dysregulation of osteogenic genes) during a sensitive early developmental window^[5,50]. The F2 metabolic profile possibly reflects a costly survival trade-off: basic homeostasis was prioritized and likely redirects metabolic resources toward essential organogenesis (e.g., brain, heart, and eyes) at the direct and sustained expense of skeletal development. Nonetheless, these hypotheses represent critical avenues, and future research is required to fully elucidate the underlying genetic and epigenetic mechanisms.

Cell signaling and developmental programming

Cell signaling and developmental programming are the fundamental architects of fish embryo development, directly responsible for establishing and maintaining a normal body shape^[51]. Any disruption to this intricate signaling can hijack these essential pathways, leading to organogenesis dysfunction and malformations such as skeletal deformities^[52]. Benzo[a]pyrene exposure may induce such disruptions through multiple interconnected signaling pathways.

A primary mechanism involves an energy crisis triggered by benzo[a]pyrene metabolism. The observed increase in adenosine monophosphate (AMP) in F0 embryos, a critical biomarker of metabolic stress, signals ATP depletion^[53]. This depletion results from benzo[a]pyrene-induced oxidative stress, cellular damage repair, and mitochondrial dysfunction^[6,54]. Elevated AMP directly inhibits the mechanistic target of rapamycin complex 1 (mTORC1), a central coordinator of anabolic processes and a known energy sensor^[55]. The consequent suppression of mTORC1 activity might dysregulate its downstream effectors, including the PI3K-Akt and FoxO signaling pathways, which are essential for cell growth, proliferation, and survival during morphogenesis^[56]. This cascade of energy-sensing failure likely underlies critical developmental defects such as failed neural tube closure, impaired organ development, and skeletal malformations.

Additionally, benzo[a]pyrene toxicity impairs cardiovascular and tissue morphogenesis, possibly through the suppression of prostaglandin I₂ (PGI₂) within the VEGF signaling pathway^[57,58]. As a potent vasodilator and inhibitor of platelet aggregation, diminished PGI₂ levels may compromise the establishment of the embryonic circulatory system, leading to impaired vessel formation,

inadequate blood flow, and localized hypoxia^[57,59]. This is evidenced by underdeveloped cardiovascular systems and enlarged pericardial cavities in benzo[a]pyrene-exposed embryos^[60,61]. Furthermore, the role of PGI₂ as a guide for cell differentiation and migration means its deficiency disrupts precise morphogenetic processes, resulting in failed organogenesis and body shape deformities. The recovery of embryonic development in the F2 generation alongside PGI₂ accumulation may further underscore its critical role; however, function validation in a subsequent study is required to ascertain this hypothesis.

The multigenerational dimension of this toxicity and its recovery is elucidated by contrasting metabolic profiles during efferocytosis. In F0 embryos, benzo[a]pyrene may induce excessive apoptosis, overloading the efferocytic machinery and depleting key signaling metabolites^[62]. Reductions in pro-resolving LysoPCs (e.g., LysoPC(22:6)) may impair phagocyte recruitment and inflammation resolution, while a concurrent drop in L-arginine cripples the production of nitric oxide and polyamines, which are essential for vascularization and osteochondral development^[62,63]. This failure possibly leads to accumulated apoptotic debris, secondary necrosis, and a persistent pro-inflammatory state that disrupts skeletal patterning, manifesting as severe malformations^[14,17,18].

On the contrary, the F2 generation exhibits a powerful, coordinated shift towards metabolic compensation and tissue repair. Elevated levels of ornithine, methionine, L-arginine, specific pro-resolving LysoPCs (including LysoPC[P-18:0]), and lactic acid indicate a restored capacity for efficient efferocytosis and anabolic support. Restored methionine levels may facilitate crucial epigenetic reprogramming, the rise in LysoPCs promotes inflammation resolution, and increased lactic acid may fuel aerobic glycolysis to generate ATP and biosynthetic precursors. Collectively, these changes create a favorable anabolic and anti-inflammatory milieu that rectifies the developmental errors of the F0 fish, enabling the substantial phenotypic recovery observed in the F2 offspring.

Neurological and sensory functions

Fish possess a sophisticated suite of neurological and sensory functions that are exquisitely adapted to their aquatic environment. Their brains feature highly developed regions, such as the optic tectum for visual processing and a large cerebellum for coordinated movement, which efficiently interpret a constant stream of sensory data^[64]. The proper formation of the head, with sufficient brain space and correctly sized eye pits, is a structural prerequisite for these normal neurological and sensory functions, which are in turn critical for survival^[65].

In F0 embryos, benzo[a]pyrene exposure disrupts this crucial development by dysregulating key neuro-specific metabolomic pathways, including neuroactive ligand-receptor interaction, GABAergic synapse, serotonergic synapse, and pathways of neurodegeneration. This molecular disruption is phenotypically manifested as a smaller eye pit and a malformed head. The toxicity is driven by a confluence of metabolic disturbances: elevated AMP signals a severe cellular energy deficit (ATP depletion) that may halt energy-intensive morphogenetic processes, while increased 5-hydroxy-L-tryptophan suggests aberrant serotonin synthesis that can lead to defective neural and vascular patterning^[53]. Simultaneously, decreased L-arginine impairs nitric oxide production, possibly compromising vasodilation and angiogenic signaling, and a reduction in PGI₂ disrupts pathways essential for vascular integrity and tissue remodeling^[43,59], warranting functional validation in a future study.

This multifaceted failure in energy supply, neurotransmitter

balance, and circulatory signaling synergistically might have caused widespread errors in organogenesis and axial patterning. The structural deformities incurred during embryonic development can also lead to neurobehavioral toxicity in post-hatch larvae, manifesting as abnormal motor behaviors and learning deficits^[5]. Conversely, the observed accumulation of L-arginine and PGI₂ in the F2 generation is likely a key factor to contribute to the alleviation of these neurological and sensory dysfunctions, facilitating a partial developmental recovery.

Limitations and implications

This study provides metabolomic insights into the multigenerational toxicity of benzo[a]pyrene in fish, yet several limitations should be acknowledged. First, while metabolomic perturbations suggest epigenetic involvement (e.g., altered α -KG and 2-HG levels), direct epigenetic measurements, such as DNA methylation or histone modifications, were not performed. Future work integrating multi-omics approaches will be essential to validate epigenetic inheritance as a mechanistic pathway. Second, the exposure was conducted under controlled laboratory conditions, which may not fully capture the complexity of natural aquatic environments where mixtures of pollutants and environmental stressors coexist. Finally, the observed phenotypic recovery in F2 larvae, interpreted here as a survival trade-off, warrants further investigation into long-term fitness consequences, such as reproductive success and disease susceptibility in later life stages or generations.

Despite these limitations, the findings carry clear environmental implications. Benzo[a]pyrene is a widespread persistent pollutant, and the demonstration of its multigenerational effects—even after exposure ceases—highlights a latent ecological risk that could compromise population resilience over time. The metabolomic shifts identified here, particularly in energy and redox metabolism, may serve as early warning biomarkers for assessing multigenerational toxicity in wild fish populations. From a regulatory perspective, these results underscore the need to consider long-term and multigenerational impacts in ecological risk assessments, rather than relying solely on acute or single-generation toxicity data. Ultimately, this work reinforces the importance of mitigating benzo[a]pyrene emissions to protect aquatic ecosystems and the biodiversity they support.

Conclusions

This study demonstrates that embryonic benzo[a]pyrene exposure at environmentally relevant concentrations induces multigenerational developmental and skeletal toxicity in medaka. The observed impairments, including elevated skeletal malformations and altered morphometrics across the F0–F2 generations, are mechanistically linked to widespread metabolomic dysregulation. Key perturbations in energy metabolism, redox balance, and cellular signaling pathways (e.g., the triad of 3-hydroxybutyric acid, α -KG, and 2-HG) provide a functional basis for the observed transgenerational phenotypes. The partial morphological recovery in the F2 generation suggests a potential survival trade-off, wherein metabolic resources are prioritized for essential functions at the expense of skeletal development. These findings underscore the value of metabolomics in revealing the latent and inherited physiological costs of early-life toxicant exposure.

Supporting information

It accompanies this paper at: <https://doi.org/10.48130/newcontam-0025-0022>.

Ethical statements

All procedures were reviewed and preapproved by the Animal Ethics Committee of Shantou University, identification number: 202210002, approval date: 10 November 2022.

Author contributions

The authors confirm their contributions to the paper as follows: Yinhua Chen: investigation, methodology, data curation, visualization, writing – original draft; Huiju Lin, Jiangang Wang, Feilong Li, Rim EL Amouri, Jack Chi-Ho Ip, Wenhua Liu: visualization, writing – review and editing; Jiezhong Mo: conceptualization, methodology, data curation, writing – original draft, funding acquisition, supervision. All authors reviewed the results and approved the final version of the manuscript.

Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Funding

This work was supported by the National Natural Science Foundation of China (Grant No. 42207318), and Shantou University STU Scientific Research Initiation Grant (Grant No. NTF23010).

Declarations

Competing interests

The authors declare that they have no known competing financial interests.

Author details

¹Guangdong Provincial Key Laboratory of Marine Disaster Prediction and Prevention, Shantou University, Shantou 515063, China; ²Department of Applied Science, School of Science and Technology, Hong Kong Metropolitan University, Hong Kong SAR, China; ³Guangdong Provincial Key Laboratory of Water Quality Improvement and Ecological Restoration for Watersheds, School of Ecology, Environment and Resources, Guangdong University of Technology, Guangzhou 510006, China; ⁴Science Unit, Lingnan University, Hong Kong SAR, China

References

- [1] Bukowska B, Mokra K, Michałowicz J. 2022. Benzo[a]pyrene—environmental occurrence, human exposure, and mechanisms of toxicity. *International Journal of Molecular Sciences* 23(11):6348
- [2] Das DN, Bhutia SK. 2018. Inevitable dietary exposure of Benzo[a]pyrene: carcinogenic risk assessment an emerging issues and concerns. *Current Opinion in Food Science* 24:16–25
- [3] Negi K, Chaudhary P. 2024. Benzo[a]pyrene: a carcinogen, its sources, adverse effects, and preventions. *The Applied Biology & Chemistry Journal* 5(3):44–56
- [4] Chepelev NL, Moffat ID, Labib S, Bourdon-Lacombe J, Kuo B, et al. 2015. Integrating toxicogenomics into human health risk assessment: lessons learned from the benzo[a]pyrene case study. *Critical Reviews in Toxicology* 45(1):44–52
- [5] Mo J, Chen Y, Lai KP, Seemann F, Liu W. 2025. Benzo[a]pyrene osteotoxicity, neurotoxicity, and epigenetic effects in fishes and mammals: a review. *Environmental Chemistry Letters* 23(6):1837–1862

- [6] Zeb R, Yin X, Chen F, Wang KJ. 2024. Chronic exposure to environmental concentrations of benzo[a]pyrene causes multifaceted toxic effects of developmental compromise, redox imbalance, and modulated transcriptional profiles in the early life stages of marine medaka (*Oryzias melastigma*). *Aquatic Toxicology* 273:107016
- [7] Chen Y, Yang Y, Qin X, Wang J, Tang G, et al. 2025. Embryonic benzo[a]pyrene exposure induces multigenerational reproductive effects on adult male medaka: phenotypic and transcriptomic insights. *Toxics* 13(10):886
- [8] Huang L, Wang C, Zhang Y, Li J, Zhong Y, et al. 2012. Benzo[a]pyrene exposure influences the cardiac development and the expression of cardiovascular relative genes in zebrafish (*Danio rerio*) embryos. *Chemosphere* 87(4):369–375
- [9] Huang L, Zuo Z, Zhang Y, Wu M, Lin JJ, et al. 2014. Use of toxicogenomics to predict the potential toxic effect of Benzo(a)pyrene on zebrafish embryos: ocular developmental toxicity. *Chemosphere* 108:55–61
- [10] Corrales J, Thornton C, White M, Willett KL. 2014. Multigenerational effects of benzo[a]pyrene exposure on survival and developmental deformities in zebrafish larvae. *Aquatic Toxicology* 148:16–26
- [11] Seemann F, Peterson DR, Witten PE, Guo BS, Shanthanagouda AH, et al. 2015. Insight into the transgenerational effect of benzo[a]pyrene on bone formation in a teleost fish (*Oryzias latipes*). *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology* 178:60–67
- [12] Mo J, Au DWT, Wan MT, Shi J, Zhang G, et al. 2020. Multigenerational impacts of benzo[a]pyrene on bone modeling and remodeling in medaka (*Oryzias latipes*). *Environmental Science & Technology* 54(19):12271–12284
- [13] Mo J, Wan MT, Au DWT, Shi J, Tam N, et al. 2023. Transgenerational bone toxicity in F3 medaka (*Oryzias latipes*) induced by ancestral benzo[a]pyrene exposure: cellular and transcriptomic insights. *Journal of Environmental Sciences* 127:336–348
- [14] Yamaguchi A, Uchida M, Ishibashi H, Hirano M, Ichikawa N, et al. 2020. Potential mechanisms underlying embryonic developmental toxicity caused by benzo[a]pyrene in Japanese medaka (*Oryzias latipes*). *Chemosphere* 242:125243
- [15] He C, Zuo Z, Shi X, Li R, Chen D, et al. 2011. Effects of benzo(a)pyrene on the skeletal development of *Sebastiscus marmoratus* embryos and the molecular mechanism involved. *Aquatic Toxicology* 101(2):335–341
- [16] Fang X, Corrales J, Thornton C, Clerk T, Scheffler BE, et al. 2015. Transcriptomic changes in zebrafish embryos and larvae following benzo[a]pyrene exposure. *Toxicological Sciences* 146(2):395–411
- [17] Elfawy HA, Anupriya S, Mohanty S, Patel P, Ghosal S, et al. 2021. Molecular toxicity of Benzo(a)pyrene mediated by elicited oxidative stress infer skeletal deformities and apoptosis in embryonic zebrafish. *Science of The Total Environment* 789:147989
- [18] Tarasco M, Gavaia PJ, Bensimon-Brito A, Cardeira-da-Silva J, Ramkumar S, et al. 2021. New insights into benzo[a]pyrene osteotoxicity in zebrafish. *Ecotoxicology and Environmental Safety* 226:112838
- [19] Bracewell-Milnes T, Saso S, Abdalla H, Nikolau D, Norman-Taylor J, et al. 2017. Metabolomics as a tool to identify biomarkers to predict and improve outcomes in reproductive medicine: a systematic review. *Human Reproduction Update* 23(6):723–736
- [20] Zhao J, Yao K, Yu H, Zhang L, Xu Y, et al. 2021. Metabolic remodelling during early mouse embryo development. *Nature Metabolism* 3(10):1372–1384
- [21] Cheung NKM, Hinton DE, Au DWT. 2012. A high-throughput histoarray for quantitative molecular profiling of multiple, uniformly oriented medaka (*Oryzias latipes*) embryos. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology* 155(1):18–25
- [22] da Silva Moreira S, de Lima Inocêncio LC, Jorge BC, Reis ACC, Hisano H, et al. 2021. Effects of benzo(a)pyrene at environmentally relevant doses on embryo-fetal development in rats. *Environmental Toxicology* 36(5):831–839
- [23] Wang D, Rietdijk MH, Kamelia L, Boogaard PJ, Rietjens IMCM. 2021. Predicting the *in vivo* developmental toxicity of benzo[a]pyrene (BaP) in rats by an *in vitro*-*in silico* approach. *Archives of Toxicology* 95(10):3323–3340
- [24] Costa J, Reis-Henriques MA, Castro LFC, Ferreira M. 2012. Gene expression analysis of ABC efflux transporters, CYP1A and GST α in Nile tilapia after exposure to benzo(a)pyrene. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology* 155(3):469–482
- [25] Kranz J, Hessel S, Aretz J, Seidel A, Petzinger E, et al. 2014. The role of the efflux carriers Abcg2 and Abcc2 for the hepatobiliary elimination of benzo[a]pyrene and its metabolites in mice. *Chemico-Biological Interactions* 224:36–41
- [26] Yuan L, Lv B, Zha J, Wang W, Wang Z. 2014. Basal and benzo[a]pyrene-induced expression profile of phase I and II enzymes and ABC transporter mRNA in the early life stage of Chinese rare minnows (*Gobio-cypris rarus*). *Ecotoxicology and Environmental Safety* 106:86–94
- [27] Takahashi M. 2012. Oxidative stress and redox regulation on *in vitro* development of mammalian embryos. *The Journal of Reproduction and Development* 58(1):1–9
- [28] Hou Z, Fuiman LA. 2020. Nutritional programming in fishes: insights from mammalian studies. *Reviews in Fish Biology and Fisheries* 30(1):67–92
- [29] Miyazawa H, Aulehla A. 2018. Revisiting the role of metabolism during development. *Development* 145:dev131110
- [30] Riddle MR, Hu CK. 2021. Fish models for investigating nutritional regulation of embryonic development. *Developmental Biology* 476:101–111
- [31] Mierziak J, Burgberger M, Wojtasik W. 2021. 3-Hydroxybutyrate as a metabolite and a signal molecule regulating processes of living organisms. *Biomolecules* 11(3):402
- [32] Lin AP, Abbas S, Kim SW, Ortega M, Bouamar H, et al. 2015. D2HGDH regulates alpha-ketoglutarate levels and dioxygenase function by modulating IDH2. *Nature Communications* 6(1):7768
- [33] Baksh SC, Finley LWS. 2021. Metabolic coordination of cell fate by α -ketoglutarate-dependent dioxygenases. *Trends in Cell Biology* 31(1):24–36
- [34] Lin YC, Wu CY, Hu CH, Pai TW, Chen YR, et al. 2020. Integrated hypoxia signaling and oxidative stress in developmental neurotoxicity of benzo[a]pyrene in zebrafish embryos. *Antioxidants* 9(8):731
- [35] Du X, Hu H. 2021. The roles of 2-hydroxyglutarate. *Frontiers in Cell and Developmental Biology* 9:651317
- [36] Zhan CL, Lu QY, Lee SH, Li XH, Kim JD, et al. 2024. IDH2 and GLUD1 depletion arrests embryonic development through an H4K20me3 epigenetic barrier in porcine parthenogenetic embryos. *Zoological Research* 45(6):1175–1187
- [37] Fang X, Thornton C, Scheffler BE, Willett KL. 2013. Benzo[a]pyrene decreases global and gene specific DNA methylation during zebrafish development. *Environmental Toxicology and Pharmacology* 36(1):40–50
- [38] Trujillo AS, Labeille RO, Jayarajan R, Mack D, Seemann F. 2025. Parental benzo[a]pyrene exposure impacts histone modifications in osteoblast subpopulations. *Environmental Epigenetics* 11(1):dvaf032
- [39] Kundu D, Dubey VK. 2021. Purines and pyrimidines: metabolism, function and potential as therapeutic options in neurodegenerative diseases. *Current Protein & Peptide Science* 22(2):170–189
- [40] Li W, Hu J, Adebali O, Adar S, Yang Y, et al. 2017. Human genome-wide repair map of DNA damage caused by the cigarette smoke carcinogen benzo[a]pyrene. *Proceedings of the National Academy of Sciences of the United States of America* 114(26):6752–6757
- [41] Godschalk RWL, Verhofstad N, Verheijen M, Yauk CL, Linschooten JO, et al. 2015. Effects of benzo[a]pyrene on mouse germ cells: heritable DNA mutation, testicular cell hypomethylation and their interaction with nucleotide excision repair. *Toxicology Research* 4(3):718–724
- [42] Jurecka A, Tyliki-Szymanska A. 2022. Inborn errors of purine and pyrimidine metabolism: a guide to diagnosis. *Molecular Genetics and Metabolism* 136(3):164–176
- [43] Wu G, Meininger CJ, McNeal CJ, Bazer FW, Rhoads JM. 2021. Role of L-arginine in nitric oxide synthesis and health in humans. In *Amino Acids in Nutrition and Health*, ed. Wu G. Volume 1332. Cham: Springer. pp. 167–187 doi: 10.1007/978-3-030-74180-8_10
- [44] Pallisco R, Lazzarino G, Bilotta G, Marroni F, Mangione R, et al. 2023. Metabolic signature of energy metabolism alterations and excess nitric oxide production in culture media correlate with low human embryo

- quality and unsuccessful pregnancy. *International Journal of Molecular Sciences* 24(1):890
- [45] Baliou S, Adamaki M, Ioannou P, Pappa A, Panayiotidis MI, et al. 2021. Protective role of taurine against oxidative stress. *Molecular Medicine Reports* 24(2):1–19
- [46] Huang M, Yang X, Zhou Y, Ge J, Davis DA, et al. 2021. Growth, serum biochemical parameters, salinity tolerance and antioxidant enzyme activity of rainbow trout (*Oncorhynchus mykiss*) in response to dietary taurine levels. *Marine Life Science & Technology* 3(4):449–462
- [47] Casagrande S, Hau M. 2019. Telomere attrition: metabolic regulation and signalling function? *Biology Letters* 15(3):20180885
- [48] Tobler M, Gómez-Blanco D, Hegemann A, Lapa, M., Neto JM, et al. 2022. Telomeres in ecology and evolution: a review and classification of hypotheses. *Molecular Ecology* 31(23):5946–5965
- [49] Amabile A, Migliara A, Capasso P, Biffi M, Cittaro D, et al. 2016. Inheritable silencing of endogenous genes by hit-and-run targeted epigenetic editing. *Cell* 167(1):219–232.e14
- [50] Jeremias G, Gonçalves FJM, Pereira JL, Asselman J. 2020. Prospects for incorporation of epigenetic biomarkers in human health and environmental risk assessment of chemicals. *Biological Reviews of the Cambridge Philosophical Society* 95(3):822–846
- [51] Li P, Elowitz MB. 2019. Communication codes in developmental signalling pathways. *Development* 146(12):dev170977
- [52] Dahlen CR, Borowicz PP, Ward AK, Caton JS, Czernik M, et al. 2021. Programming of embryonic development. *International Journal of Molecular Sciences* 22(21):11668
- [53] Marín-Aguilar F, Pavillard LE, Giampieri F, Bullón P, Cordero MD. 2017. Adenosine monophosphate (AMP)-activated protein kinase: a new target for nutraceutical compounds. *International Journal of Molecular Sciences* 18(2):288
- [54] Das DN, Naik PP, Mukhopadhyay S, Panda PK, Sinha N, et al. 2017. Elimination of dysfunctional mitochondria through mitophagy suppresses benzo[a]pyrene-induced apoptosis. *Free Radical Biology and Medicine* 112:452–463
- [55] Dunlop EA, Tee AR. 2009. Mammalian target of rapamycin complex 1: signalling inputs, substrates and feedback mechanisms. *Cellular Signalling* 21(6):827–835
- [56] Yu JSL, Cui W. 2016. Proliferation, survival and metabolism: the role of PI3K/AKT/mTOR signalling in pluripotency and cell fate determination. *Development* 143(17):3050–3060
- [57] Andrade GM, Martínez GV, Mayoral LPC, Hernández-Huerta MT, Zenteno E, et al. 2020. Molecules and prostaglandins related to embryo tolerance. *Frontiers in Immunology* 11:555414
- [58] Fu C, Li Y, Xi H, Niu Z, Chen N, et al. 2022. Benzo[a]pyrene and cardiovascular diseases: an overview of pre-clinical studies focused on the underlying molecular mechanism. *Frontiers in Nutrition* 9:978475
- [59] Sugimoto Y, Aikawa S, Inazumi T, Hirota Y. 2025. Roles of prostaglandin signaling in implantation and decidualization. *Progress in Lipid Research* 100:101357
- [60] Gerger CJ, Weber LP. 2015. Comparison of the acute effects of benzo[a]pyrene on adult zebrafish (*Danio rerio*) cardiorespiratory function following intraperitoneal injection versus aqueous exposure. *Aquatic Toxicology* 165:19–30
- [61] Cherr GN, Fairbairn E, Whitehead A. 2017. Impacts of petroleum-derived pollutants on fish development. *Annual Review of Animal Biosciences* 5(1):185–203
- [62] Henson PM. 2017. Cell removal: efferocytosis. *Annual Review of Cell and Developmental Biology* 33(1):127–144
- [63] Mehrotra P, Ravichandran KS. 2022. Drugging the efferocytosis process: concepts and opportunities. *Nature Reviews Drug Discovery* 21(8):601–620
- [64] Engel-Yeger B. 2021. The involvement of altered sensory modulation in neurological conditions and its relevance to neuro-rehabilitation: a narrative literature review. *Disability and Rehabilitation* 43:2511–2520
- [65] Laberge F, Hara TJ. 2001. Neurobiology of fish olfaction: a review. *Brain Research Reviews* 36(1):46–59



Copyright: © 2026 by the author(s). Published by Maximum Academic Press, Fayetteville, GA. This article is an open access article distributed under Creative Commons Attribution License (CC BY 4.0), visit <https://creativecommons.org/licenses/by/4.0>.