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https://doi.org/10.48130/tihort-0025-0008

Technology in Horticulture 2025, 5: e013

# Augmenting thermo-hydro-chemical seed priming and germination dynamics on genotype/cultivar-Arka Shubra: a spineless green manure legume cover crop *Mucuna pruriens* L. DC. var. utilis

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#### **Abstract**

The exploration of seed priming techniques represents a critical avenue for enhancing agricultural productivity, particularly in leguminous cover crops like *Mucuna pruriens* L. DC. var. utilis is commonly known as the velvet bean. This spineless green manure legume has significant potential for sustainable agricultural systems, offering multiple ecosystem services including soil fertility improvement, erosion control, and nitrogen fixation. Over 2022–2023, an experiment was conducted to evaluate the impact of diverse seed treatments on the germination dynamics and seedling growth of trichomeless (spineless) *Mucuna pruriens*. The experiment was designed in an RCBD with three replications, occurred under shaded conditions. Various treatments were applied, including scarification, soaking in hot and normal water,  $H_2SO_4$ ,  $KNO_3 + HNO_3$ ,  $GA_3$ , and thiourea. The results indicated that  $H_2SO_4$  treatment significantly enhanced the germination rate (57.14%  $\pm$  2.49%) and reduced the mortality rate (42.86%  $\pm$  2.49%) compared with the other treatments. Additionally, the  $H_2SO_4$  treatment exhibited higher values for VI I (2.91  $\pm$  0.21) and VI II (51.18  $\pm$  4.41) than the other treatments. Furthermore, the application of  $H_2SO_4$  resulted in a greater seedling drying weight (0.90  $\pm$  0.04 g) in comparison to other treatments, with  $GA_3$  showing similar trend at  $0.82 \pm 0.02$  g. The observed PCA plot indicates that PC-1 and PC-2 collectively explained 96.17% of the total variation. These findings highlight the effectiveness of  $H_2SO_4$  in promoting favorable germination and seedling growth characteristics in spineless *Mucuna pruriens*.

Citation: Jnanesha AC, Ranjith Kumar S, Bharath Kumar S, Venu Gopal S, Sravya K, et al. 2025. Augmenting thermo-hydro-chemical seed priming and germination dynamics on genotype/cultivar-Arka Shubra: a spineless green manure legume cover crop *Mucuna pruriens* L. DC. var. utilis. *Technology in Horticulture* 5: e013 https://doi.org/10.48130/tihort-0025-0008

# Introduction

The velvet bean is a member of the *Fabaceae* family and commonly referred to as '*M. pruriens* L. DC. var. utilis', is an annual climbing leguminous, green manure, legume, and land cover crop plant<sup>[1-4]</sup>. *M. pruriens* (L.) DC. grows wild in India<sup>[5-7]</sup> and thrives in tropical and subtropical regions of America, Africa, Asia, the Pacific Islands, and India<sup>[8-11]</sup>. It has 150 species worldwide, including 15 documented in India. *M. pruriens* is a medicinally valuable crop<sup>[12,13]</sup>. In India, two main species of *M. pruriens* (L.) DC. are commonly found: *M. pruriens* var. pruriens and *M. pruriens* var. utilis. *M. pruriens* var. pruriens, a wild variation with a black seed coat, has reddish-brown irritating trichomes on the pod that produce severe itching when in contact with the skin<sup>[14]</sup>.

Human contact with *Mucuna pruriens* (L.) DC. leads to itchy dermatitis attributed to *mucuna* production<sup>[15]</sup>. Due to this, farmers were hesitating mainly in terms of cultivation and toughness in harvesting this crop. The other varieties, like CIM-Ajar, CIM-Nirom (released by CSIR-CIMAP, Lucknow), and Arka Shubra<sup>[13,14,16,17]</sup>, possess non-irritating trichomes, and their distinctive velvety appearance is due to the dense silky trichomes, earning them the common name 'velvet beans' [12,18–20].

This study focused on *M. pruriens* (L.) DC., var. utilis, a commercially significant plant renowned for treating central nervous system disorders such as dementia, Parkinson's, and Alzheimer's. Almost, all plant parts contain L-DOPA, with seeds having the highest quantity, followed by roots, stems, and leaves. *Mucuna pruriens* (L.) DC., var.

utilis (variety: Arka Shubra, developed by ICAR-IIHR, Hessaraghatta Lake Post, Bengaluru) seeds exhibited notably high L-DOPA content (51.9 mg/g) and proline (1.74 mg/g), along with strong antioxidant activity (86.5%)<sup>[21]</sup>. This medicinally valuable crop is used for culinary purposes, with pods consumed as vegetables and leaves utilized as animal feed<sup>[22–24]</sup>. The characteristic pods bear fruits, and this plant has been studied in various studies<sup>[23,25,26]</sup>.

L-DOPA, a valuable compound found in *M. pruriens* is in high demand globally, with the world market reaching 250 tons/year, costing USD\$101 billion/year<sup>[27–29]</sup>. *M. pruriens* is a versatile plant with historical, cultural, and medicinal significance, offering a range of pharmacological properties and facing challenges in meeting the high demand for its valuable compound, L-DOPA<sup>[30–32]</sup>. The velvet beans exhibit intense itching and dermatitis upon contact<sup>[30,32]</sup>. Despite L-DOPA being present in all plant parts, its extraction from wild populations faces challenges due to limited availability, posing difficulties in meeting the growing demand<sup>[33,34]</sup>.

Spineless *M. pruriens* is a tropical leguminous plant that is celebrated for its versatility, serving as a cover crop, forage, traditional medicine source, and high-protein food<sup>[13,35–37]</sup>. Similar to other legumes like soybeans, common beans, and mung beans, velvet beans thrive in environments with ample moisture and warmth, both in cultivated and wild varieties<sup>[38–42]</sup>. To maximize its potential, understanding and optimizing the germination process are crucial. Seed treatments, including scarification and soaking, offer potential methods to enhance seed germination and, subsequently, crop establishment.

Mortality (%) =  $\left[\frac{\text{No. of seeds ungerminated} + \text{Dead seedlings}}{\text{Total no. of seeds}}\right] \times 100$ 

The survival (%) was calculated using the following formula:

Survival (%) = 
$$\left[\frac{\text{No. of healthy seedlings}}{\text{Total no. of seeds}}\right] \times 100$$

For survival (%) we use visual observations for healthy seedlings that typically exhibit vibrant green leaves, strong stems, and a well-developed root system. Signs of distress include yellowing leaves, wilting, stunted growth, or discoloration.

Days to germination were calculated as:

Days to germination = Days to initial emergence – Days to final emergence

Days to Initial Emergence refers to the number of days required for the first seedling to break through the soil surface after planting. In contrast, Days to Final Emergence indicates the total number of days for all seeds within a given treatment to emerge. Additionally, 'Days to Germination' is defined as the time from planting to the first emergence of a seedling, rather than the difference between initial and final emergence.

The speed of germination was calculated using the following formula<sup>[31]</sup>:

$$GS = \frac{\sum ni}{\sum di}$$

Here, 'ni' is the number of germinated seeds, and 'di' is the total number of days.

The Vigor index was calculated by the following formula<sup>[32]</sup>:

$$VI = \frac{GP \times SL}{100}$$

Here, 'GP' is the germination (%), and 'SL' is the seedling length.

# Observations

The following seven attributes were recorded: ASL = average seedling length (cm); DFG = days for germination; Germ (%) = germination (%); Mort (%) = mortality (%); SDW = seedlings dry weight (g); SP = survival (%), and VI = Vigour Index.

#### Statistical analysis

The mean data were analyzed using IBM SPSS Statistics Ver. 19 software<sup>[44]</sup> for DMRT (Duncan's Multiple Range Test). Multivariate PCA was conducted using PAST Ver. 4.3 software to assess the impact of various treatments on the germination of *M. Pruriens*, and correlation analysis was employed to further investigate the relationships between the treatment variables and germination outcomes.

# **Results and discussion**

#### Germination and mortality (%) (GP and MP)

The germination (%) of spineless M. Pruriens exhibited notable variations under different treatments.  $\text{H}_2\text{SO}_4$  treatment yielded the highest germination rate at  $57.14\% \pm 2.49\%$ , surpassing other treatments. Hot water treatment followed closely, with a germination rate of  $44.25\% \pm 1.35\%$ , comparable to the  $42.86\% \pm 1.87\%$  observed with normal water treatment. In contrast, the control group exhibited a significantly lower germination rate at  $22.0\% \pm 0.96\%$  compared to the rest of the treatments. This outcome suggests that applying  $\text{H}_2\text{SO}_4$  positively influenced germination, outperforming both hot and normal water treatments. The efficacy of  $\text{H}_2\text{SO}_4$  in enhancing germination may be attributed to its specific effects on seed coat permeability and the release of dormancy mechanisms  $^{[43,45-47]}$ . The relatively high germination rates observed with hot water and normal water treatments also indicate their potential to promote seed germination.

Thermo-hydro-priming, a method of soaking seeds in water to initiate germination without sprouting, has effectively enhanced germination rates and seedling vigor in various crops. This technique activates metabolic processes, improving germination rates and seedling growth. Chemical priming has also been shown to have positive effects on root development and seedling vigor<sup>[2]</sup>. It can significantly improve germination rates, with some studies indicating up to 88% germination when combined with mechanical treatments<sup>[40]</sup>. Seed priming enhances physiological traits, such as root and shoot length, and biochemical responses, contributing to improved crop establishment and yield. This study aimed to evaluate the effects of different seed priming treatments on germination rate and seedling establishment and determine the optimal seed priming treatment for improving germination dynamics and seedling establishment in Mucuna pruriens L. DC. var. utilis (Arka Shubra).

# **Materials and methods**

# **Experimental site**

During 2022–2023, the experiment was conducted under Shade net Nursery conditions, at the CSIR-CIMAP RC, Experimental Farm in Hyderabad (17.25° N latitude and 78.33° E longitude). Spineless *M. pruriens* (L.) DC. Var. utilis, genotype/variety (Arka Shubra) seeds were used in this experiment, and it was designed in a randomized complete block design (RCBD) with seven treatment combinations with three repetitions.

# **Seed collection**

Seeds of spineless *M. pruriens* (L.) DC. Var., genotype/variety: Arka Shubra, were obtained from ICAR-IIHR, Hessaraghatta Lake Post, Bengaluru farm, ensuring genetic purity and viability. The study examined the influence of different seed treatments on seed germination, using the following methods:

#### Scarification

The Seeds were scarified by chemicals to break the seed coat.

#### Soakina

The Seeds were soaked in normal water (24 h) and hot water (80  $^{\circ}$ C) for 5 min.

# **Treatments**

The following seed treatments were used in the experiment:  $T_1$  (control),  $T_2$  (hot water at 80 °C for 5 min),  $T_3$  (normal water for 24 h),  $T_4$  ( $H_2SO_4$  1% for 5 min),  $T_5$  (KNO $_3$ 1% + HNO $_3$  1% for 24 h),  $T_6$  (GA $_3$  500 ppm for 24 h), and  $T_7$  (thiourea for 24 h). A total of 30 seeds for each treatment were subjected to their respective treatments and were sown in separate trays under controlled environmental conditions on June 10<sup>th</sup>, 2022, and June 22<sup>nd</sup>, 2023. In this experiment, we utilized sand, vermiculite, and paper towels as substrate. The seeds were exposed to normal daylight conditions, and the temperature was maintained within the range of 20–30 °C to facilitate optimal germination. Germination was monitored daily, and the percentage of germinated seeds was recorded.

# Sampling and measurement

To obtain a complete picture of germination behavior, the following parameters were calculated using formulas and methodologies:

Germination (%) was calculated using the following formula<sup>[43]</sup>:

Germination (%) = 
$$\left[\frac{\text{No. of seeds germinated}}{\text{Total no. of seeds}}\right] \times 100$$

The mortality (%) was calculated using the following formula:

The markedly lower germination rate in the control group underscores the importance of the applied treatments in optimizing germination conditions for spineless *M. Pruriens* (L.) DC. The obtained results are corroborated with the findings of Wanjekeche et al.<sup>[48]</sup> who found that *Mucuna* seeds treated in hot water recorded higher germination compared to the control. Similarly, the mortality percentages in spineless *M. Pruriens* varied significantly across treatments, with the control group recording a markedly higher mortality rate of 78% compared to the other groups.

In contrast, the  $H_2SO_4$  treatment exhibited the lowest mortality rate at 42.86%  $\pm$  2.49%, followed by hot water treatment (55.75%  $\pm$  1.35%) and normal water treatment (57.14%  $\pm$  1.87%) (Table 1). This divergence in mortality rates highlights the potential impact of different treatments on the survival of *Mucuna pruriens*. The significantly higher mortality in the control group suggests that natural conditions or a lack of specific treatments may adversely affect seedling survival. Conversely, the lower mortality rates observed in the  $H_2SO_4$ , hot water, and normal water treated plants indicate their potential to enhance seedling survival.

Spineless *Mucuna* seeds commonly display both physical and physiological dormancy, hindering germination in a timely and uniform manner<sup>[49,50]</sup>. This dormancy is attributed to the impermeability of the seed coat and the presence of inhibitory substances within the seed. Sulfuric acid is recognized for its capacity to break seed dormancy via scarification, which entails weakening or thinning the seed coat <sup>[51]</sup>. The application of H<sub>2</sub>SO<sub>4</sub> can effectively target the robust seed coat of Spineless *Mucuna*, promoting water absorption and facilitating the initiation of germination<sup>[34,51]</sup>.

Enhanced germination observed in spineless *Mucuna* seeds following acid scarification with H<sub>2</sub>SO<sub>4</sub> can be attributed to the alleviation of physical and physiological barriers that impede gaseous exchange and water uptake. This treatment likely facilitated improved imbibitions and respiration, which are critical for initiating metabolic processes and fostering early germination. Similar findings have been reported by Bhuse et al.<sup>[52]</sup> in Senna species, where acid scarification significantly improved germination rates by overcoming seed coat dormancy. The hard seed coat in *Mucuna pruriens* acts as a physical barrier to water and oxygen diffusion. H<sub>2</sub>SO<sub>4</sub>, a strong acid, chemically erodes the seed coat, creating micro-pores and cracks that enhance permeability. This mechanical breakdown allows for faster and more efficient imbibitions, which is essential for rehydrating seed tissues and activating metabolic processes.

#### Average seedling length (ASL) cm

The average seedling length of Spineless *Mucuna* exhibited notable variations under different treatments, with the application of  $\rm H_2SO_4$  resulting in a significantly higher seedling length of 19.67  $\pm$  0.83 cm. This length was comparable to that observed after the application of GA<sub>3</sub> (19.39  $\pm$  0.40 cm) and normal water (19.0  $\pm$  0.83 cm) (Table 1). In contrast, the control group exhibited a significantly shorter seedling length of 8.89  $\pm$  0.39 cm.

These findings indicate that  $H_2SO_4$  treatment not only positively influenced the seedling length of Spineless *Mucuna* but also outperformed the effects of both  $GA_3$  and normal water treatments. This suggests that  $H_2SO_4$  may play a crucial role in promoting seedling growth, potentially by enhancing nutrient uptake and improving the physiological processes essential for growth. The effectiveness of  $H_2SO_4$  in stimulating seedling development could be attributed to its ability to break seed dormancy and improve seed coat permeability, thereby facilitating better access to water and nutrients.

Conversely, the significantly reduced seedling length observed in the control group highlights the critical importance of external treatments in fostering the growth and development of *Mucuna pruriens* seedlings. The lack of any treatment in the control group likely resulted in suboptimal conditions for germination and early growth, underscoring the necessity of implementing effective pregermination strategies to enhance seedling vigor.

These results are consistent with the findings of Fiallos et al.<sup>[53]</sup>, who also reported the positive effects of various treatments on seedling growth in leguminous species. The implications of this study extend beyond the immediate effects on seedling length; they suggest that the application of treatments such as H<sub>2</sub>SO<sub>4</sub> and GA<sub>3</sub> can significantly improve the establishment and overall health of Spineless *Mucuna* seedlings.

# Survival (%) (SP %)

The survival rates of spineless *Mucuna* seedlings following treatment with  $H_2SO_4$  were significantly higher, reaching 56.98%  $\pm$  1.73%, in contrast to the remaining treatments. Subsequent in efficacy were hot water treatment (43.94%  $\pm$  1.16%) and normal water treatments (39.42%  $\pm$  1.04%) (Table 1). The control group exhibited a significantly lower survival rate of 20.56%  $\pm$  0.53% compared with the other treatments, with thiourea treatment following closely. The results highlight the remarkable impact of  $H_2SO_4$  treatment on the survival of Spineless *Mucuna* seeds, surpassing the effects of both hot and normal water treatments. This underscores the potential utility of  $H_2SO_4$  in enhancing seed viability and germination. The lower survival rate observed in the control group emphasizes the importance of specific treatments in fostering optimal conditions for seedling survival.

These findings underscore the significant positive impact of  $H_2SO_4$  treatment on the survival of Spineless *Mucuna* seedlings, highlighting its effectiveness in enhancing seed viability and promoting successful germination. The superior survival rate associated with  $H_2SO_4$  treatment can be attributed to its role in breaking seed dormancy and improving seed coat permeability, which facilitates better water absorption and nutrient uptake. This treatment likely creates more favorable conditions for seedling establishment, leading to higher survival rates.

The notably lower survival rate observed in the control group emphasizes the critical need for specific treatments to foster

 Table 1.
 Thermo-hydro-chemical seed treatments and germination dynamics of Mucuna pruriens L. DC. var. utilis.

Treatments	Germination (%)	Mortality (%)	SL (cm)	Survival (%)	DG	SDW (g)
T <sub>1</sub> : Control	22.00 + 0.96e	78.00 + 0.96a	8.89 + 0.39c	20.56 + 0.53f	14.26 + 0.38a	0.30 + 0.04f
T <sub>2</sub> : Hot water	44.25 + 1.35b	55.75 + 1.35d	17.33 + 0.76b	43.94 + 1.16b	11.23 + 0.54d	0.68 + 0.05d
T <sub>3</sub> : Normal water	42.86 + 1.87bc	57.14 + 1.87d	19.00 + 0.83a	39.42 + 1.04c	10.67 + 0.81f	0.74 + 0.03c
T <sub>4</sub> : H <sub>2</sub> SO <sub>4</sub>	57.14 + 2.49a	42.86 + 2.49e	19.67 + 0.86a	56.98 + 1.73a	8.69 + 0.26g	0.90 + 0.04a
T <sub>5</sub> : KNO <sub>3</sub> + HNO <sub>3</sub>	28.94 + 1.26d	71.06 + 1.26bc	16.81 + 0.73b	27.42 + 0.87d	13.33 + 0.34b	0.36 + 0.08
T <sub>6</sub> : GA <sub>3</sub>	27.65 + 0.58d	72.35 + 0.58b	19.39 + 0.40a	26.89 + 0.92d	9.67 + 0.21e	0.82 + 0.02b
T <sub>7</sub> : Thiourea	27.01 + 0.71d	72.99 + 0.71b	16.98 + 0.45b	26.05 + 0.54de	12.65 + 0.37c	0.50 + 0.04de

SL, Seedling length (cm); SP, Survival (%); DG, Days to germination; SDW (g), Seedling dry weight; Average data followed by similar letter in the same column means not significantly different based on 5% of DMRT (Duncan's Multiple Range Test).

optimal conditions for seedling survival. Without these interventions, seedlings may struggle to thrive because of factors such as physical dormancy and inadequate access to moisture and nutrients. This finding reinforces the idea that pre-germination treatments are essential for improving the establishment and growth of *Mucuna pruriens* seedlings.

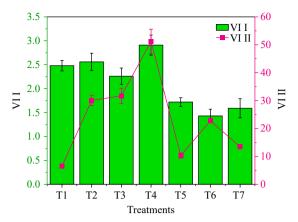
# Days for germination (DFG)

The germination period for spineless *Mucuna* seeds was significantly shorter under the treatment with  $H_2SO_4$ , requiring only 8.69  $\pm$  0.26 days, compared with the other treatments. Following closely in efficiency was the  $GA_3$  treatment. Conversely, the control group exhibited a prolonged germination period of 14.26  $\pm$  0.38 days and followed by  $KNO_3 + HNO_3$  treatment (13.33  $\pm$  0.34 d) (Table 1). The notable reduction in germination days observed with  $H_2SO_4$  treatment suggests its effectiveness in expediting the germination process of spineless *Mucuna* seeds. This finding indicates the potential utility of  $H_2SO_4$  in optimizing germination conditions.

The implications of these results are particularly relevant for the cultivation of *Mucuna pruriens*, as faster germination can lead to earlier establishment of the crop and potentially higher yields. Previous studies have shown that treatments such as acid scarification can effectively reduce germination times in various leguminous species by overcoming physical dormancy and enhancing seed coat permeability<sup>[41]</sup>. Moreover, the prolonged germination periods observed in the control group and in the KNO<sub>3</sub> + HNO<sub>3</sub> treatment underscore the challenges posed by untreated seeds, which may struggle to germinate due to inherent dormancy mechanisms.

#### Vigour Index (VI)

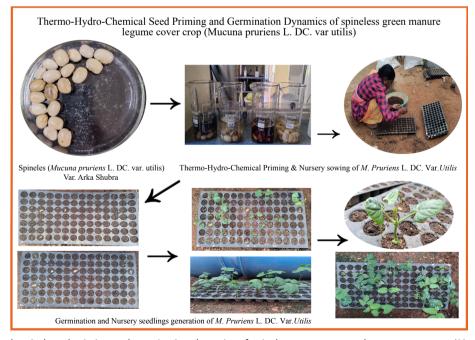
 $\rm H_2SO_4$  treatment resulted in significantly higher values for VI I and VI II, measuring 2.91  $\pm$  0.21 and 51.18  $\pm$  4.41, respectively, compared to all other treatments. The hot water treatment was closely followed, with values of 2.56  $\pm$  0.18 and 30.08  $\pm$  1.81 for VI I and VI II, respectively. Conversely, the control group exhibited significantly lower VI I (6.53  $\pm$  0.41) compared to the other treatments, with the KNO<sub>3</sub>  $\pm$  HNO<sub>3</sub> treatment recording the second-lowest values of 1.72  $\pm$  0.09 and 10.30  $\pm$  0.94 for VI I and VI II, respectively (Table 1 &



**Fig. 1** Vigour index with the different treatments (error bars with standard deviation) in *Mucuna pruriens* L. DC. var. utilis). [ $T_1$  (control),  $T_2$  (hot water 80 °C for 5 min),  $T_3$  (normal water for 24 h),  $T_4$  ( $H_2SO_4$  1% for 5 h),  $T_5$  (KNO $_3$  1% + HNO $_3$  1% for 24 h),  $T_6$  (GA $_3$  500 ppm for 24 h), and  $T_7$  (thiourea for 24 h); ASL = Average seedling length; DFG = Days for germination; GP/Germ% = Germination (%); Mort% = Mortality (%); SDW= Seedling Dry Weight; SP/Sur% = Survival (%); VI I = Vigour index 1; VI II = Vigour Index 2].

Fig. 1). The substantial increase in VI I and VI II observed after  $\rm H_2SO_4$  treatment highlights its effectiveness in promoting these indices, suggesting a positive influence on the physiological and biochemical aspects of treated specimens. The hot water treatment also demonstrated notable effects, albeit to a lesser extent. In contrast, the control group exhibited significantly lower values, highlighting the importance of specific treatments in achieving favorable physiological responses (Figs 2, 3 & 4).

While the hot water treatment was also demonstrated beneficial effects on vigor indices, its impact was less pronounced than that of  $H_2SO_4$ . This suggests that while hot water can be an effective treatment for promoting germination and early growth, it may not be as potent as  $H_2SO_4$  for optimizing physiological responses in Spineless *Mucuna* seedlings.



**Fig. 2** Thermo-hydro-chemical seed priming and germination dynamics of spineless green manure legume cover crop (*Mucuna pruriens* L. DC. var. utilis).



Fig. 3 Seedling height of *Mucuna pruriens* in (a) (T<sub>4</sub>: H<sub>2</sub>SO<sub>4</sub> 1% for 5 min), and (b) (T<sub>2</sub>: hot water 80 °C for 5 min) thermo-hydro-chemical seed priming treatments.



Fig. 4 Trichomeless varieties of (a) CIM Ajar, (b) CIM-Nirom, and (c) wild collection with trichomes of Mucuna pruriens.

The significantly lower values observed in the control group highlight the critical role of specific treatments in achieving favorable physiological outcomes. Without these interventions, seedlings may experience suboptimal growth conditions, leading to reduced vigor and overall health.

# Seedling dry weight (g) (SDW)

Applying  $\rm H_2SO_4$  resulted in a significantly higher seedling dry weight of  $0.90 \pm 0.04$  g compared to all other treatments, with GA<sub>3</sub> followed closely at  $0.82 \pm 0.02$  g. Conversely, the control group exhibited a significantly lower seedling dry weight of  $0.30 \pm 0.04$  g. The substantial increase in seedling dry weight under the  $\rm H_2SO_4$  treatment suggests its efficacy in promoting robust growth and biomass accumulation. The observed higher dry weight in the GA<sub>3</sub> treatment further underscores the positive impact of specific treatments on seedling development. In contrast, the control group's significantly lower seedling dry weight emphasizes the importance of external factors in influencing plant growth. The markedly lower seedling dry weight in the control group underscores the critical importance of external factors, such as pre-germination treatments, in influencing plant growth. Without these interventions, seedlings may struggle to establish themselves, leading to reduced biomass

and overall vigor. This finding aligns with the existing literature that emphasizes the necessity of employing effective treatments to optimize seedling growth and development<sup>[41,54]</sup>.

# Correlation matrix and principal component analysis (PCA)

The correlation matrix provides insights into the degree of association between different variables. From the results, a strong positive correlation (r = 0.996) was observed between germination % and survival %, indicating that higher germination rates are strongly associated with better seedling survival. Similarly, a strong positive correlation (r = 0.956) was found between germination % and vigor index II, suggesting that seeds with higher germination rates tended to exhibit better overall vigor (Fig. 5). On the other hand, a strong negative correlation was observed between mortality % and germination % (r = -0.99), as well as between days to germination and seedling dry weight (r = -0.991). This implies that seeds that germinate faster tend to produce seedlings with higher dry weight, indicating that early germination is advantageous for seedling development. Additionally, negative correlations were noted between days to germination and vigor index (r = -0.896) and between seedling length and days to germination (r = -0.812) (Fig. 5). These findings

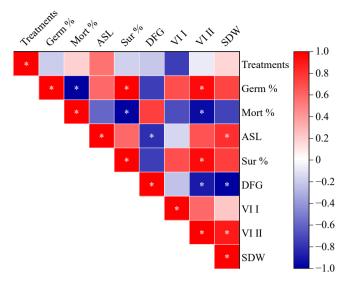
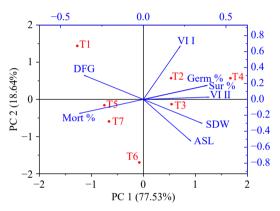


Fig. 5 Correlation matrix of germination traits in *Mucuna pruriens*.



**Fig. 6** Principal component analysis (PCA) for treatments with germination-related traits in *Mucuna pruriens* [ $T_1$  (control),  $T_2$  (hot water 80 °C for 5 min),  $T_3$  (normal water for 24 h),  $T_4$  ( $H_2SO_4$  1% for 5 h),  $T_5$  (KNO $_3$  1% + HNO $_3$  1% for 24 h),  $T_6$  (GA $_3$  500 ppm for 24 h), and  $T_7$  (thiourea for 24 h); ASL = Average seedling length; DFG = Days for germination; GP/Germ% = Germination (%); Mort% = Mortality (%); SDW= Seedling Dry Weight; SP/Sur% = Survival (%); VI I = Vigour index 1; VI II = Vigour Index 2].

**Table 2.** Correlation analysis of germination related traits of *Mucuna pruriens* L. DC. var. utilis.

Characters	G (%)	M (%)	SL (g)	SP (%)	DG	VII	VIII	SDW (g)
G (%)	1	-0.99	0.596	0.996	-0.747	0.697	0.956	0.739
M (%)		1	-0.600	-0.996	0.746	-0.698	-0.957	-0.738
SL (g)			1	0.591	-0.812	-0.155	0.670	0.802
SP (%)				1	-0.754	0.698	0.958	0.742
DG					1	-0.228	-0.896	-0.991
VII						1	0.593	0.231
VIII							1	0.887
SDW (g)								1

GP (%), Germination (%); Mort (%), Mortality (%); SP (%), Survival (%); DG, Days to germination; VI, Vigour Index; SL, Seedling length (cm); SDW (g), Seedling dry weight; Correlation is significant at 5%.

highlight that early and high germination rates are critical for overall seedling success. Reducing the time to germination may enhance multiple seedling characteristics, and selecting for high germination percentages is likely to improve survival rates. To assess the variability among treatment effects and germination characteristics, principal component analysis (PCA) was employed on eight parameters of M. pruriens. The resulting PCA plot (Fig. 6) revealed that the first two principal components (PC-1 and PC-2) collectively accounted for 96.17% of the total variation. PC-1 explained 77.53% of this variation, with positive contributions from VI I, VI II, Germination %, and Survival %, and negative contributions from ASL and SDW. PC-2 explained an additional 18.64% of the total variation, with a positive contribution from DFG and a negative contribution from mortality %. Notably, treatments  $T_2$  and  $T_4$  exhibited positive contributions, while the remaining treatments had negative contributions. Furthermore, the correlation matrix depicted significant correlations among various parameters. Germination % exhibited noteworthy correlations with VI 2 and survival (Table 2).

#### **Conclusions**

The findings of this study underscore the significant impact of various seed treatments on the germination and growth characteristics of spineless M. pruriens. The application of H<sub>2</sub>SO<sub>4</sub> emerged as the most effective treatment, leading to the highest germination percentage, reduced mortality rates, enhanced seedling length, and increased seedling dry weight. These results suggested that H2SO4 not only facilitates the breaking of seed dormancy but also promotes robust growth and biomass accumulation, thereby optimizing the conditions for seedling establishment. Hot water and normal water treatment also demonstrated positive effects, although to a lesser extent than H<sub>2</sub>SO<sub>4</sub>. The control group, which lacked any specific treatment, exhibited significantly lower germination rates and higher mortality, highlighting the critical role of targeted interventions in improving seedling survival and growth. Our results align with existing literature, reinforcing the notion that effective seed treatments can substantially enhance the physiological and biochemical responses of M. pruriens. Principal component analysis (PCA) further elucidated the relationships among various growth parameters, revealing that specific treatments positively influenced key indices such as vigor and survival rates. The significant correlations identified among germination percentage, seedling length, and survival rates emphasize the interconnectedness of these traits in determining overall seedling performance. The results of this study have implications for developing sustainable agriculture practices, particularly in regions where M. pruriens is a key crop.

# **Author contributions**

The authors confirm contribution to the paper as follows: study conception and design: Jnanesha AC; analysis and interpretation of results: Venu Gopal S, Kumar A, Sravya K; draft manuscript preparation: Ranjith Kumar S, Bharath Kumar S; manuscript review and revision: Lal RK. All authors have reviewed the results and approved the final version of the manuscript.

# **Data availability**

All data generated or analyzed during this study are included in this published article.

# **Acknowledgment**

The authors thank the Director, CSIR-Central Institute of Medicinal and Aromatic Plants, Lucknow (U.P.), India for encouragement and support to experiment. CIMAP Publication No.: CIMAP/PUB/2023/167.

#### Conflict of interest

The authors declare that they have no conflict of interest.

#### **Dates**

Received 19 August 2024; Revised 5 February 2025; Accepted 17 February 2025; Published online 2 April 2025

# References

- Fiebrig IN, Van de Vijver M, Van Kan CJ. 2023. Mucuna pruriens vs. Morbus Parkinson: making the case for medicinal supplements within medicinal agroecology. In Medicinal Agro ecology, 1<sup>st</sup> edition, ed. Fiebrig I. Boca Raton: CRC Press. pp. 45–87. doi: 10.1201/9781003146902-5
- Lepcha P, Shekhar M, Murugesan L, Jaheer M, Chopra R, et al. 2024.
   Association mapping of important agronomic traits in *Mucuna pruriens* (L.) DC. *Botanical Studies* 65:26
- Nooreen Z, Wal A, Shukla A, Yadav AK. 2023. Mucuna pruriens magical velvet bean the wonder plant - a review. Scope Journal 13(2):340–63
- Ortiz Ceballos Ál, Aguirre Rivera JR, Osorio Arce MM, Peña Valdivia C. 2012. Velvet bean (*Mucuna pruriens* var. utilis) as a cover crop and bioherbicide to preserve the environmental services of soil. In *Herbi*cides - Environmental Impact Studies and Management Approaches, 1st edition, ed. Alvarez-Fernandez R. UK: IntechOpen. pp. 168–84. doi: 10.5772/31833
- Eagleton GE, Tanzi AS, Mayes S, Massawe F, Ho WK, et al. 2023. Winged bean (Psophocarpus tetragonolobus (L.) DC.). In Neglected and Underutilized Crops, eds Farooq M, Siddique KHM. Academic Press: Elsevier. pp. 437–86. doi: 10.1016/B978-0-323-90537-4.00022-3
- Heuzé V, Tran G, Hassoun P, Renaudeau D, Bastianelli D. 2015. Velvet bean (Mucuna pruriens). Feedipedia, a programme by INRAE, CIRAD, AFZ and FAO. https://feedipedia.org/node/270
- Vishwakarma KS, Mohammed SI, Chaudhari AR, Salunkhe NS, Maheshwari VL. 2017. Micropropagation and Agrobacterium rhizogenes-mediated transformation studies in Mucunapruriens (L.). Indian Journal of Natural Products Research 8(2):172–78
- Ogbole OO, Akin-Ajani OD, Ajala TO, Ogunniyi QA, Fettke J, et al. 2023. Nutritional and pharmacological potentials of orphan legumes: Subfamily Faboideae. *Heliyon* 9(4):e15493
- 9. Singh B, Kumar D. 2019. Ethnobotanical aspects of Nagaur District, Rajasthan. Raleigh NC, United States: Lulu Publication. pp. 1–147
- Suryawanshi SS, Kamble PP, Bapat AV, Jadhav PJ. 2021. Parkinsonism and potential of Mucuna beans. In *Bioethics in Medicine and Society*. UK: Intech Open. doi: 10.5772/intechopen.92855
- Suryawanshi SS, Kamble PP, Bapat AV, Jadhav PJ. 2020. Bioactive components of magical velvet beans. In *Legume Crops - Prospects*, *Production and Uses*. UK: Intech Open. doi: 10.5772/intechopen.92124
- 12. Lahiri K, Mukhopadhyay MJ, Desjardins Y, Mukhopadhyay S. 2012. Rapid and stable in vitro regeneration of plants through callus morphogenesis in two varieties of *Mucunapruriens* L. an anti-Parkinson's drug-yielding plant. *The Nucleus* 55:37–43
- Lal RK. 2015. Genotype selection for agronomical trait seed yield in Kewanch (Mucuna pruriens (L.)). Industrial Crops and Products 65:62–70
- Patil SA, Apine OA, Surwase SN, Jadhav JP. 2013. Biological sources of L-DOPA: an alternative approach. Advances in Parkinson's Disease 2:81–87
- 15. Lampariello LR, Cortelazzo A, Guerranti R, Sticozzi C, Valacchi G. 2012. The magic velvet bean of *Mucuna pruriens*. *Journal of Traditional and Complementary Medicine* 2(4):331–39
- 16. Joshua BI, Elisha IL, Arin IE, Abubakar AS, Dennis K, et al. 2023. *Mucuna pruriens* (Karara) leaf extracts enhance certain haematological parameters in albino rats. *Acta Scientific Veterinary Sciences* 5(9):8–14
- Lal RK, Singh S, Chandra R, Sarkar S, Singh S, Dhawan OP, et al. 2015.
   Registration of new variety CIM-Nirom: A hairless pods, high seed, and L-Dopa yielding variety of Kewanch (Mucuna pruriens L.). Journal of Medicinal and Aromatic Plant Science 37(1–4):57–60
- Atakoun AM, Tovihoudji PG, Diogo RVC, Yemadje PL, Balarabe O, et al. 2023. Evaluation of cover crop contributions to conservation agriculture in northern Benin. Field Crops Research 303:109118

- 19. Rai SN, Chaturvedi VK, Singh P, Singh BK, Singh MP. 2020. *Mucuna pruriens* in Parkinson's and in some other diseases: recent advancement and future prospective. *3 Biotech* 10(12):522
- Singh SK, Dhawan SS. 2018. Analyzing trichomes and spatio-temporal expression of a cysteine protease gene Mucunain in *Mucuna pruriens* L. (DC). *Protoplasma* 255(2):575–84
- 21. Florence B, Washa WB, Nnungu S. 2024. Comparison of nutritional values of *Mucuna pruriens* L. (velvet bean) seeds with the most preferred legume pulses. *Food Production, Processing and Nutrition* 6:17
- Acevedo-Rodríguez P. 2005. Vines and climbing plants of Puerto Rico and the Virgin Islands. In Contributions from the United States National Herbarium. DC: Department of Botany, National Museum of Natural History. 483 pp
- Bindu KH, Sujatha S, Tanuja SP. 2023. Genotypic variability in yield, L-Dopa, and mineral nutrient profile in velvet bean (*Mucuna pruriens* L.) production systems. *Journal of Food Composition and Analysis* 118:105152
- Liranso TP, Wolkaro T, Gadissa S. 2023. Effect of vetch intercropping and harvesting stage on morphological characteristics, dry matter yield, and chemical composition of desho grass (Pennisetum glaucifolium). Thesis. Haramaya University, Ethiopia. http://ir.haramaya.edu.et//hru/handle/ 123456789/7005
- Ekanem AU, Edem GD, Okon KA, Emmanuel IE. 2023. Mucuna urens consumption and its implications on the ultra structure of the kidney and spleen: a laboratory investigation. Asian Journal of Immunology 6(1):224–33
- Ibrahim SE, Ochekwu EB, Ngobiri NC. 2023. Allelopathic effect of Chromolaena odorata foliage as soil amendment on the growth and chlorophyll content of Solanum lycopersicum, Mucuna pruriens, Abelmoschus esculentus and Citrullus lanatus. International Journal of Applied Science and Engineering Review 4(2):96–118
- Inamdar SA, Surwase SN, Jadhav SB, Bapat VA, Jadhav JP. 2013. Statistically optimized biotransformation protocol for continuous production of L-DOPA using Mucunamono sperma callus culture. Springer Plus 2:570
- Liu X, Han X, Peng Y, Tan C, Wang J, et al. 2022. Rapid production of L-DOPA by Vibrio natriegens, an emerging next-generation whole-cell catalysis chassis. Microbial Biotechnology 15(5):1610–22
- 29. Rai N, Dubey RK. 2023. Developments and prospects in imperative underexploited vegetable legumes breeding with special reference to biotic and abiotic stresses. *Proc of International Seminar on Exotic and Underutilized Horticultural Crops: Priorities & Emerging Trends. Bengaluru, 2013*, India: ICAR-IIHR. pp. 541–50
- 30. Mishra MK, Kumari N. 2023. *Plants for immunity and conservation strate-gies*. Singapore: Springer. xii, 383 pp. doi: 10.1007/978-981-99-2824-8
- Rakesh B, Bindu KH, Praveen N. 2021. Variations in the L-DOPA content, phytochemical constituents, and antioxidant activity of different germlines of *Mucuna pruriens*(L.) DC. *Asian Journal of Chemistry* 33(8):1881–89
- Rakesh B, Praveen N. 2022. Establishment of Mucuna pruriens (L.) DC. callus and optimization of cell suspension culture for the production of anti-Parkinson's drug: L-DOPA. Journal of Applied Biology & Biotechnology 10(5):125–35
- 33. Lohidasan S, Arulmozhi S. 2007. *Mucuna pruriens* Linn. a comprehensive review. *Pharmacognosy Review* 1(1):157–62
- Sathiyanarayanan L, Kumar V, Ramesh CK, Parmesha M, Khan MHM.
   Characterization of polyphenol oxidase in two in vitro regenerated cultivars of Mucuna: *Mucuna pruriens* L. and Mucuna prurita H. *Turkish Journal of Biology* 35(5):575–83
- 35. Elagbani NHM, Gangi AS, Sulieman AM, Babiker SAE. 2023. Forage and seed yield of velvet bean (*Mucuna pruriens* (L.)) as affected by sowing date and plant spacing under irrigated conditions, Gezira state, Sudan. World News Natural Sciences 46:1–13
- Katayama T, Kumagai H. 2010. L-DOPA, microbial production. In: Encyclopedia of Industrial Biotechnology: Bioprocess, Bioseparation, and Cell Technology. Hoboken, NJ: John Wiley & Sons, Inc. doi: 10.1002/9780470054581.eib258
- Koyanagi T, Katayama T, Suzuki H, Nakazawa H, Yokozeki K, et al. 2005.
   Effective production of 3,4-dihydroxyphenyl-L-alanine (L-DOPA) with Erwinia herbicola cells carrying a mutant transcriptional regulator TyrR. Journal of Biotechnology 115(3):303–6

- 38. Adjatin RCF, Koura BI, Adewumi M, Houinato M. 2023. Effects of supplementing processed velvet beans (*Mucuna pruriens* (L.) DC. var. *utilis*) on nutrient intakes, growth performance, and blood profile in goats. *Tropical Animal Health and Production* 55(5):311
- 39. Epriliati I. 2020. Minimum water consumption method screening of velvet bean (Mucuna sp.) processings to produce functional food ingredients. *Journal of Functional Foods and Nutraceuticals* 2(1):1–28
- 40. Mhlanga B, Thierfelder C. 2015. Cover crops in different production systems of southern Africa: their roles, benefits, and constraints. In *Cover Crops: Cultivation, Management and Benefits*, ed. Reuter J. US: Nova Science Publishers. pp. 1–22
- 41. Patel ST, Azeez MA, Krishnamurthy R. 2023. Genetic variability and divergence studies on seed traits and L-DOPA content of *Mucuna pruriens* (L.) DC. accessions. *Research Square* Preprint
- 42. Pugalenthi M, Vadivel V, Siddhuraju P. 2005. Alternative food/feed perspectives of an underutilized legume *Mucuna pruriens* var. utilis a review. *Plant Foods for Human Nutrition* 60:201–18
- 43. Raina AP, Tomar JB, Dutta M. 2012. Variability in *Mucuna prur*iens L. germplasm for L-Dopa, an anti-parkinsonian agent. *Genetic Resources and Crop Evolution* 59:1207–12
- 44. Panse VG, Sukhatme PV. 1957. Genetics of quantitative characters in relation to plant breeding. *Indian Journal of Genetics and Plant Breeding* 17(2):318–28
- 45. Jaiswal M, Deshmukh R, Patel A, Nagori K, Shukla A, et al. 2023. Parkinson's disease: neurodegeneration and the potential role of medicinal plants. *International Neurourology Journal* 27(4):567–79
- Sharma T, Ramamurthy A, Nathani S, Sharma G. 2017. A comparative pharmacognosy study of black and white seeds of Kapikacchu (*Mucuna* pruriens (L.) DC.). International Journal of Pharmaceutical Science Research 8(2):838–44
- 47. Satrio RD, Nikmah IA, Fendiyanto MH, Pratami MP, Dewi AP, et al. 2023. A complete chloroplast and mitochondrial genome for velvet bean (*Mucuna pruriens*, Fabaceae), with genome structure and intergenomic sequence transfer analyses. *Research Square* Preprint

- 48. Wanjekeche E, Wakasa V, Mureithi JG. 2003. Effect of germination, alkaline and acid soaking and boiling on the nutritional value of mature and immature *Mucuna (Mucuna pruriens)* beans. *Tropical and Subtropical Agroecosystems* 1:183–92
- Ravi C, Hadapad BS, Shetty GR, Shivaprasad M, Bindu H, et al. 2018. Evaluation of velvet bean (*Mucuna pruriens* L.) genotypes for growth, yield, L-DOPA content, and soil nitrogen fixation in rubber plantation under hill zone of Karnataka. *Journal of Pharmacognosy and Phytochemistry* 7(3S):26–29
- Ravindra PC. 2007. Effect of pre-treatment on seed germination and shade on seedling growth and yields of Mucuna pruriens (L.) DC. Thesis. Department of Forest Management and Utilization, College of Forestry, Vellanikkara. http://krishikosh.egranth.ac.in/handle/1/5810135640
- 51. Tanaka-Oda A, Kenzo T, Fukuda K. 2009. Optimal germination condition by sulfuric acid pretreatment to improve seed germination of *Sabina vulgaris* Ant. *Journal of Forest Research* 14(4):251–56
- Bhuse VH, Lad BL, Ghule ST. 2001. Effect of various seed treatments for enhancement of seed germination and seedling vigor in senna (Cassia angustifolia). Research on Crops 2(3):359–63
- Fiallos FG, Silva WM, Benavides OP. 2012. Germinação e qualidade sanitária de sementes de mucuna branca e preta utilizadas como adubo verde em Quevedo, Eguador. Scientia Agropecuaria 3(1):15–21
- 54. Barthès B, Azontonde A, Blanchart E, Girardin C, Villenave C, et al. 2004. Effect of a legume cover crop (*Mucuna pruriens* var. *utilis*) on soil carbon in an Ultisol under maize cultivation in southern Benin. *Soil Use and Management* 20(2):231–39

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