

# Genetic diversity and association analysis of the reblooming trait in bearded iris cultivars using SSR fluorescent markers

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## Abstract

Reblooming is a highly desirable trait in ornamental plants as it extends the flowering period and enhances horticultural value. This study investigated the genetic relationships of 17 bearded iris cultivars, comprising nine reblooming, and eight once-flowering cultivars. Transcriptome-wide screening identified 29,800 Simple Sequence Repeat (SSR) loci from 100,391 unigenes, with an occurrence frequency of 29.68%. Dinucleotide repeats were the most abundant (69.10%), predominantly AG/CT and AT/AT motifs, while trinucleotide repeats accounted for 30.90%, mainly AAG/CTT and AGG/CCT motifs. From 110 SSR primers designed based on these loci, eight highly polymorphic primers were selected, detecting 43 alleles with polymorphic information content (PIC) values ranging from 0.5072 to 0.6986 (average 0.6228). These markers effectively distinguished between reblooming and once-flowering cultivars, enabling the construction of molecular fingerprints and a cluster dendrogram for the 17 cultivars. Association analysis revealed that one SSR marker (P2S19) was significantly correlated with the reblooming trait. Together, these findings provides valuable insights for understanding reblooming regulation and facilitate the molecular breeding of reblooming iris cultivars.

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## Introduction

The ornamental appeal of flowering plants largely depends on their flowering characteristics, with multiple-season flowering significantly extending the visual display period. As a result, reblooming has become a major breeding goal for many ornamental plant species<sup>[1]</sup>. Reblooming, defined as the ability of plants to flower twice within a single growth cycle<sup>[2]</sup>, manifests differently across geographical regions. In temperate climates, reblooming typically occurs in spring and autumn, whereas in subtropical climates, it is more common in spring and summer. By diversifying flowering times, reblooming plants contribute to extended bloom periods, enriching the seasonal dynamics of gardens and landscapes. Consequently, plants exhibiting this trait have considerable ornamental and practical value for horticultural applications.

One notable group of ornamental plants that have attracted significant attention for reblooming traits is the bearded iris (*Iris germanica* L.), a perennial herbaceous plant of the family Iridaceae. Bearded iris is known for the distinctive beard-like appendages at the base of its outer tepals. These plants are renowned for their unique floral architecture, vibrant color variations, and primary flowering period from April to May, making them essential components in both garden settings and groundcover applications. In recent years, numerous reblooming iris cultivars, predominantly bearded iris, have been developed worldwide<sup>[3,4]</sup>. However, the breeding and identification of these cultivars face significant challenges. The unregulated introduction processes, and the poorly understood genetic mechanisms underlying reblooming traits make it difficult to identify reblooming cultivars outside their flowering periods.

Additionally, the breeding cycle for new iris cultivars typically requires a minimum of three years to reach flowering maturity, complicating early-stage evaluations of reblooming traits in hybrid progenies. These limitations have substantially hindered the broader application of reblooming *Iris* in landscape architecture and the development of novel cultivars.

To address these challenges, molecular marker technology has emerged as a powerful tool for the genetic analysis and identification of key traits in ornamental plants. Molecular markers are particularly valuable for their genetic stability, environmental independence, and high reproducibility, making them ideal for use in germplasm analysis. In *Iris* spp., a variety of molecular marker techniques, including EST-SSR, SRAP, ISSR, AFLP, SSR, and RAPD, have been employed for the analysis of genetic diversity and phylogenetic relationships. For example, the EST-SSR markers identified by Tang et al.<sup>[5]</sup> demonstrated high amplification efficiency across both wild *Iris* and horticulturally important cultivars. Furthermore, 400 ortholog-specific EST-SSR markers have provided many highly polymorphic molecular tools for comparative genetic mapping and genotyping studies in *Iris*<sup>[5]</sup>. Zhang et al.<sup>[6]</sup> integrated chloroplast and nuclear genomic markers to conduct a phylogeographic study of *Iris*. QTL mapping was utilized to elucidate the genetic architecture of reproductive isolation between *Iris fulva* and *Iris brevicaulis*<sup>[7]</sup>. Collectively, these advances highlight the critical role of molecular marker technologies not only in evolutionary studies—such as speciation and adaptive differentiation—but also in horticultural trait improvement, including cultivar identification and parental selection.

Among various molecular markers, Simple Sequence Repeats (SSR), also known as microsatellites, have gained particular attention. These tandemly repeated sequences, consisting of repeat units ranging from 1 to 6 bp, are widely distributed throughout eukaryotic genomes<sup>[8]</sup>. SSR markers offer numerous benefits, including high polymorphism, co-dominant inheritance patterns, result reliability, and minimal DNA requirements. Traditional SSR analysis relied on polyacrylamide gel electrophoresis, which was time-consuming, toxic, and limited in its ability to detect small allelic variations. In contrast, the use of fluorescently labeled primers combined with capillary electrophoresis enables more precise detection of these small variations, enhancing the efficiency and accuracy of molecular marker-based studies<sup>[9]</sup>.

The integration of association analysis with molecular marker technology has opened new avenues in plant breeding research. This approach, which leverages linkage disequilibrium (LD) in natural populations, facilitates the identification of gene loci associated with specific phenotypic traits<sup>[10–12]</sup>. By correlating highly polymorphic molecular markers with flowering traits, SSR markers linked to reblooming traits can be identified. Such markers provide invaluable tools for early-stage identification and breeding of reblooming iris cultivars, offering reliable molecular indicators for trait evaluation and scientific guidance for hybrid progeny selection and cultivar improvement. Tian et al.<sup>[13]</sup> conducted an association analysis of the molecular characteristics and floral traits of *I. germanica*, which revealed that 11 EST-SSR markers were associated with nine floral traits.

In this study, eight core SSR markers were employed to genotype 17 bearded iris cultivars to evaluate their genetic relationships, and identify molecular markers associated with reblooming traits. Firstly, the effectiveness of these markers in distinguishing reblooming and once-flowering cultivars was demonstrated. Furthermore, molecular fingerprints and a cluster tree were constructed for the 17 cultivars, providing a clear genetic differentiation among them.

Through association analysis between SSR markers and blooming traits, one SSR marker was identified as significantly associated with the reblooming trait. This discovery not only enhances our understanding of the genetic mechanisms underlying reblooming but also provides valuable molecular tools for early-stage identification and breeding of reblooming iris cultivars. The findings of this research offer theoretical insights into gene discovery related to flowering traits and molecular breeding strategies, contributing to the classification, identification, and utilization of reblooming *Iris* germplasm resources.

## Materials and methods

### Plant materials

The plant materials used in this study consisted of 17 bearded iris cultivars, as shown in Fig. 1, which were collected from a controlled environment in the horticultural experimental area in Beicang District, Ningbo City, China (29°53' N, 121°47' E). Each cultivar was assigned a unique cultivar code (CC), ranging from 1 to 17 (Table 1). Leaf samples were cleaned, sectioned, and placed in resealable bags. These samples were then transported on dry ice to the laboratory and stored at -80 °C for further use. The reblooming trait, defined as the ability to bloom twice within a single year, was scored as a binary trait: reblooming cultivars were assigned a value of 1, while once-flowering (blooming once a year) cultivars were assigned a value of 0. During the peak blooming period from April to August 2021, the maximum flower diameter and plant height of these 17 bearded iris cultivars were measured. The flower diameter was measured as the maximum horizontal width of a fully opened flower, and the plant height was determined as the vertical distance from the ground level to the highest naturally growing point of the plant. All data were measured from at least three biological replicates.



**Fig. 1** Bearded iris cultivars used in this study. Cultivar codes 1 to 17 correspond to the 17 bearded iris cultivars listed in Table 1. Bars = 1 cm.

**Table 1.** Information on bearded iris cultivars in this study.

Cultivar code	Cultivar name	Source location
1*	Antyrmn Gircys	Ningbo, Zhejiang
2*	Autumn Encorc	Ningbo, Zhejiang
3*	City Lights	Ningbo, Zhejiang
4*	Donhle Your	Ningbo, Zhejiang
5*	Earl of Essex	Ningbo, Zhejiang
6*	Gnn Rayz	Ningbo, Zhejiang
7*	I'll Be Back	Ningbo, Zhejiang
8*	Peach Jam	Ningbo, Zhejiang
9*	Victona Fall	Ningbo, Zhejiang
10	Black Beard	Ningbo, Zhejiang
11	Change of P	Ningbo, Zhejiang
12	Broad Way Star	Ningbo, Zhejiang
13	Berkeley Gold	Ningbo, Zhejiang
14	John Naa	Ningbo, Zhejiang
15	Sultan's Palace	Ningbo, Zhejiang
16	Fufted Cloud	Ningbo, Zhejiang
17	Tide'sim	Ningbo, Zhejiang

Cultivars marked with \* are reblooming.

## Genomic DNA extraction

Genomic DNA was extracted from the samples using the Genomic DNA Extraction Kit (Vazyme, Nanjing, China). The quality of the extracted DNA, including its concentration, purity, and integrity, was assessed using 1% agarose gel electrophoresis and a NanoDrop spectrophotometer (ND-1000, Isogen Life Science, Utrecht, Netherlands). The DNA samples were subsequently diluted to 20 ng/μL and stored at –20 °C for further analysis.

## Primer design

Based on the transcriptome data of bearded iris<sup>[14]</sup>, the Micro SAtellite identification tool (MISA)<sup>[15]</sup> was employed to identify SSRs. Both the 'misa.pl' and 'misa.ini' files provided by the MISA tool were utilized for SSR detection, using the default search parameters. Subsequently, Primer3<sup>[16]</sup> was utilized to design primers for SSR-containing contigs. The output from MISA served as the input for Primer3, with the process automated using Perl scripts. From the results obtained by Primer3, 110 SSR primer pairs were initially screened. The selection criteria included a PCR product size ranging from 150 to 500 bp, a melting temperature (Tm) around 60 °C, and a GC content of 40% to 60%. Among these, 55 pairs were dinucleotide repeats and 55 were trinucleotide repeats. All primers were synthesized by Sangon Biotech (Shanghai, China), and the primer sequences are listed in *Supplementary Table S1*.

## SSR marker screening and detection

A total of 110 primer pairs were initially screened for PCR amplification using four randomly selected iris cultivars, including two reblooming, and two once-flowering cultivars. Ultimately, 26 primer pairs showing clear and polymorphic bands across all four cultivars were selected for fluorescently labeling and subsequent analysis. Details of the 110 primer pairs, and the 26 selected ones are provided in *Supplementary Tables S1* and *S2*, respectively.

The PCR amplification system was as follows: every PCR reaction contained 10 μL Premix Taq™ (Code No. RR901Q, TaKaRa, Kyoto, Japan), 1 μL of DNA template (20 ng), 1 μL of forward primer (10 mM), 1 μL of reverse primer (10 mM), and 7 μL sterile distilled water, for a total volume of 20 μL. The RT-PCR cycling conditions were as follows: 3 min at 94 °C; 30 s at 93 °C; 32 cycles of 30 s at 58 °C and 72 °C for 1 min; and a final extension of 10 min at 72 °C. The amplification products were separated by 1% agarose gel electrophoresis for 12 min, then visualized and photographed under UV light.

Selected primers were labeled with a fluorescent marker (FAM) at the 5' end of the forward primer (sequence: TGTAAACGACGCCAGT), while the reverse primer sequence remained unchanged (*Supplementary Table S2*). Fluorescent-labeled primers were then used for PCR amplification of all samples, following the same reaction conditions and program.

## Fluorescent capillary electrophoresis

The PCR products (1 μL), GeneScan™ 500 LIZ™ marker (0.5 μL, Thermo Fisher Scientific, Waltham, MA, USA), and deionized formamide (8.5 μL) were mixed and loaded into a 96-well plate. The mixture was denatured at 95 °C for 5 min, cooled on ice for a minimum of 10 min, and centrifuged at 1,000 rpm for 10 s. The plate was then loaded into a capillary electrophoresis system (3730XL Genetic Analyzer, ABI, USA) for detection. The fluorescent capillary electrophoresis fragment analysis was conducted by Sangon Biotech (Shanghai) Co., Ltd (Shanghai, China).

## Statistical analysis

Flower diameter and plant height were measured based on three biological replicates. Data were analyzed with SPSS 23.0 (IBM Corp., Armonk, NY, USA). One-way ANOVA followed by Duncan's multiple range test was used to assess differences among samples, with a significance threshold level of  $p < 0.05$ .

Fluorescent capillary electrophoresis data were analyzed using GENEMAPPER v4.0<sup>[17]</sup>. For each SSR primer, the amplified fragment sizes across the 17 iris cultivars were recorded and subsequently arranged in decreasing size. The largest fragment was designated as allele A, followed by B, C, D, ... in alphabetical order. The resulting allelic data matrix was then imported into PopGen32<sup>[18]</sup> to calculate population genetic parameters, including the number of alleles (Na), observed heterozygosity (Ho), expected heterozygosity (He), Nei's genetic distance (Nei), and Shannon's information index (I). The polymorphic information content (PIC) of each SSR locus was calculated using PIC\_CALC<sup>[19]</sup>. To assess the genetic structure of the 17 bearded iris cultivars, STRUCTURE<sup>[20]</sup> was employed. K values were set from 2 to 10, with 10 independent iterations for each K. The optimal number of clusters was identified based on  $\Delta K$  values calculated using the Evanno method via the Structure Harvester tool (<https://github.com/dentearl/structureHarvester>). Genetic clustering analysis was performed using NTSYSpc v2.10e<sup>[21]</sup>, with dendrogram construction based on the unweighted pair group method with arithmetic mean (UPGMA). Associations between traits and SSR markers were evaluated using TASSEL 5.0<sup>[22]</sup>, which employed both general linear models (GLMs), and mixed linear models (MLMs).

## Results and discussion

### SSR abundance and distribution in the bearded iris transcriptome

A total of 29,800 SSR loci were identified from 100,391 unigenes in the bearded iris transcriptome, corresponding to an SSR occurrence frequency of 29.68%, and an average length of 15.05 bp (*Supplementary Table S3*). Fourteen types of repeat motifs were detected, with trinucleotide repeats representing the most diverse type (10 motif types). However, dinucleotide repeats were the most abundant, with 20,593 SSRs (69.10% of all SSRs), an occurrence frequency of 20.51%, and an average length of 14.10 bp. Trinucleotide repeats were less frequent, comprising 9,207 SSRs with an occurrence frequency of 9.17% and an average length of 16.00 bp.

Among the 20,593 dinucleotide repeats, the AG/CT motif was the most frequent (10,454, 10.41%), followed by AT/AT (6,690, 6.66%), while CG(CG repeats were relatively rare (534, 5.21%). Among the 9,207 trinucleotide repeats, AAG/CTT and AGG/CCT motifs were the

most common, with 1,933 (1.93%) and 1,761 (1.75%) occurrences, respectively, whereas ACT/AGT repeats were the least frequent (159, 0.16%) (Supplementary Fig. S1).

The lengths of SSRs in the bearded iris transcriptome ranged from 12 to 24 bp (Supplementary Table S4). Repeats shorter than 15 bp accounted for 62.89%, those between 15 and 20 bp for 1.95%, and those longer than 20 bp for 35.16%. In general, SSRs with more repeat units tended to be longer, whereas longer SSRs were less abundant.

### Identification and screening of core SSR primers

A total of 110 SSR primer pairs were initially selected for PCR amplification using four iris cultivars. The amplification products were evaluated the amplification stability and clarity through 1% agarose gel electrophoresis. Based on the quality of the amplification bands, 26 primer pairs exhibiting clear and stable bands were selected for further analysis. For instance, primer P3S41 produced distinct and stable bands during amplification (Supplementary Fig. S2). These 26 selected primers, labeled with fluorescent dyes, were further used to amplify genomic DNA from 17 bearded iris cultivars. The amplification products were subjected to fragment analysis using an ABI 3730XL Genetic Analyzer. Ultimately, eight core primer pairs were selected as optimal candidates for downstream analyses, with their sequences detailed in Table 2. These primers generated high-quality fluorescence capillary electropherograms, characterized by strong signal intensity and no interference from nonspecific peaks, demonstrating high amplification efficiency and specificity. Compared to conventional SSR techniques, this study employed fluorescent labeling combined with capillary electrophoresis, which enabled rapid, stable, and low-toxicity identification of iris cultivars. This approach offers significant advantages over traditional methods, including improved accuracy, efficiency, and reliability in cultivar identification.

Currently, molecular markers have been successfully developed and applied in genetic studies of various *Iris* species, including *I. loczyi*<sup>[6]</sup>, *I. laevigata*<sup>[23]</sup>, *I. ensata*<sup>[24]</sup>, *I. brevicaulis*<sup>[5]</sup>, *I. fulva*<sup>[5]</sup>, and *I. germanica*<sup>[13]</sup>. Within the genus *Iris*, particularly in *I. germanica*, prolonged artificial selection and cultivation have led to a continuous increase in the number of cultivars, accompanied by a rising degree of similarity among them. This has rendered traditional morphological classification increasingly inadequate for accurately distinguishing subspecies and specific cultivars. In this context, molecular marker technologies have emerged as powerful tools for the exploration of *I. germanica* resources, cultivar identification,

and breeding improvement. Studies have shown that *Iris* species and cultivars generally exhibit high levels of genetic diversity. Such genetic diversity is the product of long-term evolutionary processes and underpins the species' ability to adapt to diverse ecological environments and expand its geographic range<sup>[25]</sup>. This characteristic aligns well with the broad ecological adaptability observed across *Iris* species. However, since only 17 cultivars were tested, the representativeness of the selected core SSR primers across the wider spectrum of bearded iris germplasm remains uncertain. Additional validation with larger and more diverse cultivar sets will be required to ensure their universal applicability.

### Genetic diversity analysis using core SSR primers

The genetic diversity of 17 bearded iris cultivars was assessed using eight core SSR primers, resulting in the identification of 43 alleles. The number of alleles detected per primer ranged from 3 to 9, with an average of 5 (Table 3). The *Ho* ranged from 0.2280 to 0.8077, with a mean of 0.5877, while the *He* ranged from 0.1500 to 0.8095, with a mean of 0.4759. *Nei* ranged from 1.0943 to 1.9982, with a mean of 1.3203. The *I* varied between 1.0232 and 1.9982, averaging 1.3203. The *P/C* ranged from 0.5072 to 0.6986, all exceeding 0.5, with an average of 0.6228. Overall, these results demonstrated that the eight SSR core primers exhibited high polymorphism and genetic diversity, making them suitable for analyzing the genetic diversity of *Iris* germplasm. These findings are consistent with those reported by Zhang et al.<sup>[26]</sup> in their study on *I. germanica* cultivars.

The high genetic diversity among these iris cultivars may be attributed to their broad introduction, complex cultivation history, and extensive artificial hybridization. The rich genetic variation accumulated during the breeding process provides a robust foundation for the formation of diversity<sup>[27]</sup>. Consistent with previous studies on other horticultural plants, the SSR primers selected in this study proved to be highly effective and reliable in revealing the genetic diversity of *Iris* germplasm<sup>[28,29]</sup>. Furthermore, the results expand the potential application of SSR molecular markers in horticultural plants, offering a solid scientific foundation for the conservation, utilization, and breeding of *Iris* germplasm. Although high levels of polymorphism and genetic diversity were detected, the relatively restricted sample size may have led to an underestimation of rare alleles or subtle population structures within the broader iris germplasm. Future work incorporating more cultivars may reveal additional layers of genetic variation.

**Table 2.** Information on eight core SSR marker primers.

Primer name	Repeat unit	Repeat numbers	PCR amplicon lengths (bp)	Primer ID	Primer Sequence (5'-3') F/R	Fluorescent label
P2S19	AC	2	173	P2S19-F	TTGTAGCTCTGCAGAACGCC	5-FAM
				P2S19-R	CTGAATGATGCTCCGAACG	
P2S45	AT	2	201	P2S45-F	CGAGCTTGTCTCTCTTC	5-FAM
				P2S45-R	GGTACACAAACGAACAAGACA	
P2S48	AT	2	266	P2S48-F	AGCCAGGGCGTTCAACATA	5-FAM
				P2S48-R	TCGTGCTTCAGACAGACAAGCT	
P3S12	AAT	3	102	P3S12-F	TGACGTGGACAATGCATCA	5-FAM
				P3S12-R	AACAGCCTCGAAAGCTTAT	
P3S17	AAT	3	262	P3S17-F	TCTGCTTCAGGCTCATGG	5-FAM
				P3S17-R	AGGATTTATCAGGCCGCG	
P3S28	AGA	3	208	P3S28-F	AGGAGCCAACCCGAATGAAG	5-FAM
				P3S28-R	TTGAGGCCAGGCACATGAT	
P3S30	AGA	3	162	P3S30-F	TAGAACCAACCCCTGTCACCT	5-FAM
				P3S30-R	CCGAGGCCCTCTCAACGAAT	
P3S41	AGT	3	259	P3S41-F	TGCTGTTACGCCCTCACCTT	5-FAM
				P3S41-R	AATTCCCTCTCAGGCCGGCC	

**Table 3.** Genetic diversity analysis of bearded iris cultivars based on eight core SSR markers.

Marker	<i>Na</i>	<i>Ho</i>	<i>He</i>	<i>Nei</i>	<i>I</i>	<i>PIC</i>
P2S19	5	0.5294	0.6934	0.6730	1.2830	0.6210
P2S45	9	0.8095	0.8606	0.8401	1.9982	0.8226
P2S48	5	0.3810	0.7619	0.7438	1.4294	0.6986
P3S12	6	0.7619	0.5784	0.5646	1.1331	0.5235
P3S17	3	0.1500	0.6808	0.6638	1.0943	0.5897
P3S28	6	0.1579	0.7496	0.7299	1.4915	0.6916
P3S30	4	0.4615	0.6062	0.5828	1.0232	0.5072
P3S41	5	0.5556	0.6079	0.5910	1.1095	0.5279
Mean	5	0.4759	0.6924	0.6737	1.3203	0.6228

Note: *Na*, number of alleles; *Ho*, observed heterozygosity; *He*, expected heterozygosity; *Nei*, Nei's genetic distance; *I*, Shannon's information index; *PIC*, polymorphic information content.

### Genetic relationship and clustering analysis of 17 bearded iris cultivars

Using the eight core SSR primers, 39 polymorphic bands were amplified by capillary electrophoresis across 17 bearded iris cultivars. Each band was assigned a sequential number from 1 to 39. For each cultivar, the presence or absence of bands was coded as 1 or 0,

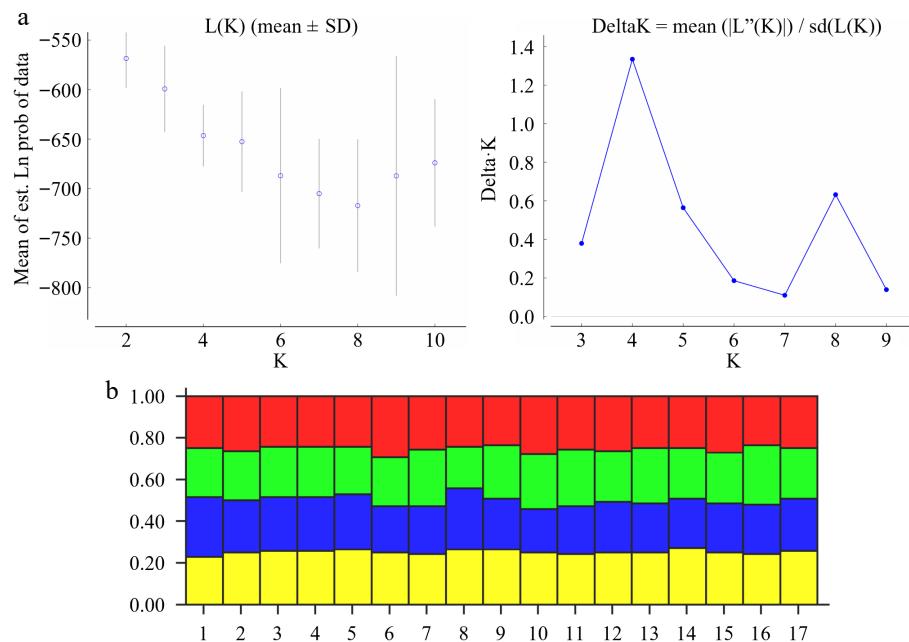
respectively, resulting in a binary (0,1) matrix. This matrix represented a 39-bit molecular identification (ID) (**Table 4**).

The genetic structure of the 17 bearded iris cultivars was analyzed using STRUCTURE software based on SSR markers. The optimal number of genetic clusters was determined to be  $K = 4$ , as this value corresponded to the highest  $\Delta K$ , indicating the most likely population structure (**Fig. 2a**). At  $K = 4$ , most cultivars exhibited admixed genetic backgrounds without clear differentiation, although slight variations were observed in some individuals (e.g., CCs 7 and 8) (**Fig. 2b**). These results highlight the overall genetic composition and admixture patterns among the cultivars.

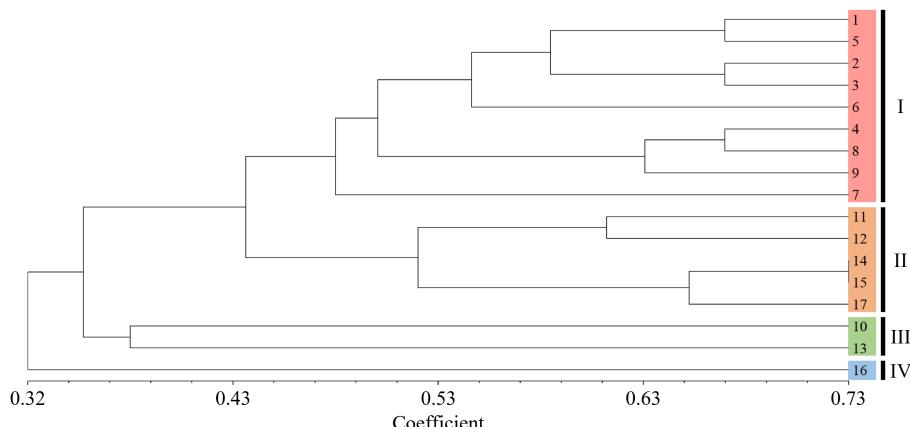
To validate the clustering pattern and explore genetic relationship, UPGMA-based hierarchical clustering was performed. At a relative genetic distance of 0.43, the 17 bearded iris cultivars were divided into four distinct groups (**Fig. 3**). Group I comprised nine cultivars (CCs 1–9), all of which exhibited reblooming traits; Five cultivars (CCs 11, 12, 14, 15, and 17) clustered into Group II, while two cultivars (CCs 10 and 13) clustered into Group III. Group IV was represented solely by 'Fufted Cloud' (CC 16), which formed an independent cluster. Notably, Groups II, III, and IV consisted of once-flowering cultivars. These clustering results suggest a significant association between blooming periodicity traits and genetic

**Table 4.** Binary (0,1) matrix representing molecular IDs of 17 bearded iris cultivars.

Band	Cultivar																
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
Band 1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	1
Band 2	1	1	1	0	0	1	1	1	1	1	1	0	1	0	0	0	0
Band 3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Band 4	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
Band 5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
Band 6	0	0	1	0	0	1	0	0	0	0	1	0	0	0	1	0	0
Band 7	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
Band 8	1	0	0	0	0	0	0	0	0	1	0	1	0	0	0	1	0
Band 9	0	0	1	1	1	0	0	1	1	0	0	0	0	1	1	0	1
Band 10	0	1	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0
Band 11	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
Band 12	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Band 13	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
Band 14	1	0	0	0	1	1	0	0	0	0	0	0	0	0	1	1	0
Band 15	0	1	1	0	0	0	0	0	0	1	0	0	0	0	0	0	0
Band 16	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0
Band 17	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
Band 18	0	1	1	0	1	0	0	0	1	0	1	1	0	1	1	0	1
Band 19	1	1	1	0	1	0	0	0	0	0	0	0	0	1	0	1	1
Band 20	0	0	1	1	0	0	0	0	0	0	1	0	0	1	1	0	0
Band 21	1	0	0	1	1	0	1	1	1	1	1	0	0	0	0	1	0
Band 22	1	0	1	0	0	0	1	0	1	0	1	1	0	1	1	1	1
Band 23	1	1	0	0	1	1	1	1	0	0	1	1	1	1	1	0	0
Band 24	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Band 25	1	1	0	0	1	1	1	0	0	0	0	0	0	1	0	1	0
Band 26	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Band 27	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Band 28	0	0	1	0	0	0	0	0	0	1	0	0	1	0	1	0	1
Band 29	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Band 30	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
Band 31	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
Band 32	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
Band 33	0	1	1	1	1	1	0	1	1	0	1	0	0	0	1	0	0
Band 34	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0
Band 35	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
Band 36	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
Band 37	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0
Band 38	0	0	0	0	0	0	0	1	0	1	1	1	0	1	1	0	1
Band 39	0	0	0	0	0	1	1	0	1	0	0	0	1	0	0	0	0



**Fig. 2** Population structure of 17 bearded iris cultivars. (a) Estimation of sub-population number. The left figure presents  $\ln$  (data probability) vs  $K$  (2–10), and the right figure illustrates sub-population number against  $\Delta K$ . (b) Population structure analysis of 17 bearded iris cultivars. Cultivar codes 1 to 17 correspond to the 17 bearded iris cultivars listed in Table 1.



**Fig. 3** Cluster analysis of 17 bearded iris cultivars. Cultivar codes 1 to 17 correspond to the 17 bearded iris cultivars listed in Table 1. The clustering was performed based on the genetic relationships revealed by SSR fluorescence molecular markers. Genetic clustering analysis was performed using NTSYSpc v2.10e, with dendrogram construction based on the unweighted pair group method with arithmetic mean (UPGMA).

relationships among the iris cultivars. The eight core SSR primers effectively distinguished reblooming iris cultivars from once-flowering cultivars, supporting the feasibility of molecular markers for analyzing reblooming traits.

The clustering analysis also revealed significant genetic diversity among once-flowering cultivars. For instance, 'Fufted Cloud' formed an independent group, suggesting its unique genetic background or specific breeding history compared to other once-flowering cultivars. This uniqueness may be attributed to its distinct genetic composition, potentially involving specific gene combinations that differentiate it from other once-flowering cultivars. Such genetic distinctiveness could be linked to its morphological or physiological traits, such as flower size, color, or environmental adaptability. This finding aligns with previous studies highlighting the genetic complexity and diversity within iris cultivars<sup>[30,31]</sup>.

It is noteworthy that the 17 bearded iris cultivars analyzed in this study included nine well-established reblooming varieties, such as 'Peach Jam' (No. 8), which was previously documented by Tian

et al.<sup>[13]</sup>. To minimize the potential impact of the environment on phenotype, all plant materials were cultivated under controlled conditions. Furthermore, DNA-based molecular markers are largely unaffected by environmental influences and are characterized by high polymorphism, thereby offering a robust approach for reducing environmental bias<sup>[32]</sup>. The use of core SSR markers combined with cluster analysis effectively differentiated reblooming and once-flowering cultivars and revealed distinct clustering patterns reflecting their underlying genetic relationships. It should also be noted that the clustering patterns observed here were based on a limited number of cultivars. With broader sampling, the boundaries between groups may shift, and additional subgroups might be detected, providing a more refined picture of the genetic structure.

#### Association analysis of SSR markers with the reblooming trait in Iris

Association analyses between SSR molecular markers and the reblooming trait, flower diameter, and plant height of 17 bearded iris cultivars were conducted using the GLM and MLM models. The

phenotypic traits of the bearded iris cultivars during the blooming period are summarized in **Table 5**. Specifically, the cultivars numbered 1–9 are reblooming types, whereas cultivars 10–17 are once-flowering types. Among these 17 cultivars, flower diameter ranges from 8.6 to 12.5 cm, and plant height ranges from 42.3 to 99 cm. Quantile-quantile (QQ) plots were generated to visualize the results of the association analyses.

The GLM analysis revealed a significant association between the SSR marker P2S19 and the reblooming trait ( $p < 0.05$ ), whereas no significant association was detected using the MLM model (**Fig. 4a** and **b**). This discrepancy may arise from differences in the characteristics of the two models. The GLM model exhibits higher sensitivity for small sample populations, facilitating the detection of potential associations; however, it is also prone to false positives. In contrast, the MLM model accounts for population structure and kinship effects, thereby reducing the likelihood of false positives, but it is more suitable for large sample sizes and may have reduced sensitivity in small populations<sup>[33]</sup>. The significant association of P2S19 suggests that it may reside in a key genetic region regulating the reblooming trait. This finding warrants further verification through the construction of high-density genetic maps or genome-wide association studies (GWAS). No SSR markers were identified as being

significantly associated with flower diameter or plant height, which may reflect the complex genetic architecture underlying these traits.

Despite the limited number of SSR markers significantly associated with the reblooming trait identified in this study, the credibility of the present findings remains robust. The genetic mechanism underlying the reblooming phenotype is likely highly complex, potentially involving the coordinated regulation of multiple genes and the contribution of rare or low-frequency alleles. Tian et al.<sup>[13]</sup> conducted a genetic diversity analysis of 25 *I. germanica* cultivars using EST-SSR markers, and identified 22 primer pairs with high polymorphism, of which 50% were significantly associated with nine phenotypic traits. However, although their study documented the blooming types of the cultivars, it did not include an association analysis specifically targeting the reblooming trait—possibly due to the absence of detectable correlations.

In the present study, 110 SSR primer pairs were screened and eight core markers with high levels of polymorphism (average PIC = 0.6228) selected, which successfully distinguished all 17 tested cultivars, including a clear separation between reblooming and once-flowering types. Notably, one of these markers showed a significant association with the reblooming trait. Compared to previous studies, the present research provides a novel perspective by focusing on molecular marker identification related to the reblooming trait in *Iris*, laying a preliminary foundation for understanding its genetic basis. The findings offer valuable theoretical support for marker-assisted selection and genetic improvement of *Iris*, and present a promising strategy for the molecular breeding of other reblooming ornamental plants. The ability of SSR markers to distinguish reblooming from once-flowering cultivars, as demonstrated by clustering analysis, further highlights their potential to facilitate breeding programs aimed at enhancing desirable traits, such as prolonged flowering duration. Given the restricted sample size, the statistical power of association analysis was limited, which may partly explain why only one marker (P2S19) showed a significant correlation with the reblooming trait. Future research should incorporate larger and more geographically diverse cultivar sets, employ a greater density of molecular markers (e.g., SNP-based genotyping or additional SSRs), and integrate detailed phenotypic data. Such approaches will enhance the robustness of diversity estimates, provide a more comprehensive understanding of the genetic architecture of Iris germplasm, and improve the power of trait–marker association analyses.

## Conclusions

This study systematically evaluated the genetic diversity and established genetic relationships among 17 bearded iris cultivars

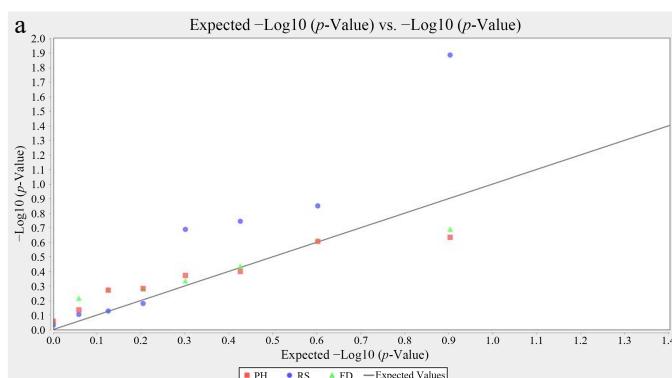
**Table 5.** Phenotypic traits of bearded iris cultivars during blooming.

Cultivar code	Reblooming status <sup>a</sup>	Flower diameter (cm) <sup>b</sup>	Plant height (cm) <sup>c</sup>
1	1	10.3 ± 0.3cde	61.8 ± 0.8cd
2	1	10.2 ± 0.6cdef	70.2 ± 1.6bc
3	1	10.3 ± 0.6cde	70.0 ± 16.6bc
4	1	9.5 ± 0.1ef	42.3 ± 4.5e
5	1	10.5 ± 0.4cde	62.0 ± 6.2bcd
6	1	12.5 ± 0.7ab	77.7 ± 4.2b
7	1	10.8 ± 0.5bcd	53.3 ± 4.8de
8	1	11.0 ± 0.4bcd	92.3 ± 7.4a
9	1	13.8 ± 0.9a	99.0 ± 9.2a
10	0	10.0 ± 1.1def	66.3 ± 8.5bcd
11	0	11.3 ± 0.6bcd	70.7 ± 1.7bc
12	0	8.6 ± 0.3f	66.7 ± 1.8bcd
13	0	11.5 ± 0.1bcd	62.0 ± 7.4bcd
14	0	9.8 ± 1.7def	72.3 ± 1.2bc
15	0	11.3 ± 0.2bcd	65.3 ± 1.7bcd
16	0	11.7 ± 0.2bc	60.3 ± 2.5cd
17	0	11.5 ± 0.4bcd	60.0 ± 6.4cd

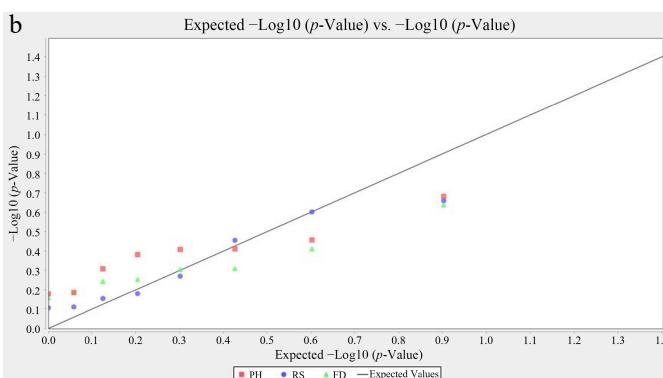
<sup>a</sup> 1 indicates a reblooming cultivar, while 0 indicates a once-flowering cultivar.

<sup>b</sup> The average maximum flower diameter of three biological replicates.

<sup>c</sup> The average maximum plant height of three biological replicates. Different letters indicate significant differences at  $p < 0.05$ , following statistical analysis by ANOVA.



**Fig. 4** Quantile-quantile plot of the association analysis for the reblooming trait in bearded iris cultivars. (a) The generalized linear regression model (GLM). (b) The mixed linear model (MLM). RS, Reblooming Status; FD, Flower Diameter; PH, Plant Height.



using SSR marker analysis. A total of 29,800 SSR loci were identified from transcriptome data, providing abundant resources for molecular marker development. Among these, dinucleotide repeats (particularly AG/CT and AT/AT) were the most prevalent, while trinucleotide motifs such as AAG/CTT and AGG/CCT were common but less abundant. Through the screening of 110 SSR primers derived from these loci, eight core primers were identified as highly polymorphic and effective in differentiating reblooming from once-flowering cultivars. Notably, the SSR marker P2S19 was significantly associated with the reblooming trait ( $p < 0.05$ ), suggesting its potential linkage to genomic regions regulating the reblooming trait. The integration of transcriptome-derived SSR identification, fluorescent capillary electrophoresis, and association analysis not only enhances our understanding of *Iris* genetics but also demonstrates the practical utility of SSR markers in germplasm identification, genetic diversity assessment, and molecular breeding. These findings provide a solid genomic and technical foundation for the improvement and conservation of bearded iris cultivars.

## Author contributions

The authors confirm contributions to the paper as follows: study conception and design: Li D, Xia Y; data collection, analysis and interpretation of results: Shao L, Xu T, Wang X, Zhang R, Cui Y, Ren Z; draft manuscript preparation: Shao L; review of the manuscript: Li D, Xia Y, Zhou H, Zhang J. All authors reviewed the results and approved the final version of the manuscript.

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## Conflict of interest

The authors declare that they have no conflict of interest.

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## References

1. Garbutt S. 2003. Remontant Iris. *The Garden* 128(9):676
2. Tang HM, Ma WZ. 2021. Discussion on sustainable flower border construction in South China. *Southern Horticulture* 32(6):73–76 (in Chinese)
3. Warburton B, Hamblen M (Eds.). 1978. *The World of Irises*. Utah, US: The Publishers Press, The American Iris Society. pp. 138–44
4. Austin C, Austin J. 2005. *Irises: A Gardener's Encyclopedia*. America: Timber Press. pp. 22–195
5. Tang S, Okashah RA, Cordonnier-Pratt MM, Pratt LH, Johnson VE, et al. 2009. EST and EST-SSR marker resources for *Iris*. *BMC Plant Biology* 9:72
6. Zhang G, Han Y, Wang H, Wang Z, Xiao H, et al. 2021. Phylogeography of *Iris loczyi* (Iridaceae) in Qinghai-Tibet Plateau revealed by chloroplast DNA and microsatellite markers. *AoB Plants* 13:plab070
7. Ballerini ES, Brothers AN, Tang S, Knapp SJ, Bouck A, et al. 2012. QTL mapping reveals the genetic architecture of loci affecting pre- and postzygotic isolating barriers in Louisiana Iris. *BMC Plant Biology* 12:91
8. Schuelke M. 2000. An economic method for the fluorescent labeling of PCR fragments. *Nature Biotechnology* 18:233–34
9. Cheng BY, Xia JH, Gong JY, Yang SH. 2011. Application of capillary electrophoresis detection with fluorescent SSR markers in rice DNA fingerprint identification. *Chinese Journal of Rice Science* 25(6):672–76 (in Chinese)
10. Li Q, Lu X, Wang C, Shen L, Dai L, et al. 2022. Genome-wide association study and transcriptome analysis reveal new QTL and candidate genes for nitrogen - deficiency tolerance in rice. *The Crop Journal* 10:942–51
11. Dou J, Wang Y, Yang H, Niu H, Liu D, et al. 2022. Development of branchless watermelon near isogenic lines by marker assisted selection. *Horticultural Plant Journal* 8:627–36
12. Laosatit K, Amkul K, Chankaew S, Somta P. 2022. Molecular genetic diversity of winged bean gene pool in Thailand assessed by SSR markers. *Horticultural Plant Journal* 8:81–88
13. Tian F, Tian Y, Yu F, Qian J, Wang F, et al. 2025. Association analysis of the molecular characteristics and floral traits of *Iris × germanica*. *Czech Journal of Genetics and Plant Breeding* 61:55–66
14. Fan Z, Gao Y, Ren Y, Guan C, Liu R, et al. 2020. To bloom once or more times: the reblooming mechanisms of *Iris germanica* revealed by transcriptome profiling. *BMC Genomics* 21:553
15. Thiel T, Michalek W, Varshney R, Graner A. 2003. Exploiting EST databases for the development and characterization of gene-derived SSR-markers in barley (*Hordeum vulgare* L.). *Theoretical and Applied Genetics* 106:411–22
16. Rozen S, Skaletsky H. 2000. Primer3 on the WWW for general users and for biologist programmers. *Methods in Molecular Biology* 132:365–86
17. Chatterji S, Pachter L. 2006. Reference based annotation with GeneMapper. *Genome Biology* 7:R29
18. Yeh FC, Yang RC, Boyle T. 1997. *POPGENE, the User Friendly Shareware for Population Genetic Analysis*. Molecular Biology and Biotechnology Centre, University of Alberta, Alberta
19. Nagy S, Poczai P, Cernák I, Gorji AM, Hegedűs G, et al. 2012. PICcalc: an online program to calculate polymorphic information content for molecular genetic studies. *Biochemical Genetics* 50:670–72
20. Pritchard JK, Stephens M, Donnelly P. 2000. Inference of population structure using multilocus genotype data. *Genetics* 155:945–59
21. Rohlf FJ. 1987. NTSYS-pc: microcomputer programs for numerical taxonomy and multivariate analysis. *The American Statistician* 41:330
22. Bradbury PJ, Zhang Z, Kroon DE, Casstevens TM, Ramdoss Y, et al. 2007. TASSEL: software for association mapping of complex traits in diverse samples. *Bioinformatics* 23:2633–35
23. Sun MZ, Li MR, Shi FX, Li L, Liu Y, et al. 2012. Genomic and EST-derived microsatellite markers for *Iris laevigata* (Iridaceae) and other congeneric species. *American Journal of Botany* 99:e286–e288
24. Xiao YE, Hu YH, Liu M, Chen XY. 2012. Isolation and characterization of polymorphic microsatellites in *Iris ensata* (Iridaceae). *American Journal of Botany* 99:e498–e500
25. Tong J, Zhou Y, Mao J, Dong YF, Xu DY, et al. 2015. Genetic diversity analysis of Iris cultivars using ISSR markers. *Northern Horticulture* 10:104–7 (in Chinese)
26. Zhang YX, Miao M, Yuan HY, Yang YH, Liu QQ, et al. 2017. ISSR analysis of 23 German iris varieties (lines). *Jiangsu Agricultural Sciences* 45(2):47–50 (in Chinese)
27. Wang R. 2010. The Study of genetic diversity of populations of *Iris halophila* Pall. in Xinjiang with SSR and SRAP markers. Thesis. Xinjiang Agricultural University, China. pp. 2–3 (in Chinese)
28. Chang HB, Wang C, He MJ, Cao XM, Yu FF, et al. 2025. Genetic diversity analysis of 69 maize germplasm resources based on SSR markers. *Crops* 2:47–53 (in Chinese)
29. Qiu H, Wang M, Xiao X, Li K, Ye J, et al. 2025. Genetic diversity analysis of cassava germplasm resources based on phenotypic traits and SSR markers. *Journal of Southern Agriculture* 8:2490–500 (in Chinese)

30. Zalmat AS, Sotola VA, Nice CC, Martin NH. 2021. Genetic structure in Louisiana Iris species reveals patterns of recent and historical admixture. *American Journal of Botany* 108:2257–68

31. Roguz K, Gallagher MK, Senden E, Bar-Lev Y, Lebel M, et al. 2020. All the colors of the rainbow: diversification of flower color and intraspecific color variation in the genus *Iris*. *Frontiers in Plant Science* 11:569811

32. Agarwal M, Shrivastava N, Padh H. 2008. Advances in molecular marker techniques and their applications in plant sciences. *Plant Cell Reports* 27:617–31

33. Huang M, Li X, Wang K, Jiang Q, Li W, et al. 2024. Analysis of population genetics and effective components associations with SSR markers in *Callicarpa nudiflora*. *Chinese Traditional and Herbal Drugs* 55(23):8162–70 (in Chinese)



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