

Versatile roles of trehalose in plant growth and development and responses to abiotic stress

Yuqing Han¹, Adan Liang¹, Dongdong Xu¹, Yujie Zhang¹, Jiali Shi³, Meng Li², Tao Liu^{1*} and Hongyan Qi^{1*}

¹ College of Horticulture, Shenyang Agricultural University, Key Laboratory of Protected Horticulture, Ministry of Education, Northern National & Local Joint Engineering Research Center of Horticultural Facilities Design and Application Technology (Liaoning), Shenyang 110866, China

² College of Horticulture, Henan Agricultural University, Zhengzhou, Henan 450002, China

³ Jiuquan Academy of Agricultural Sciences, Jiuquan, Gansu 735000, China

* Corresponding authors, E-mail: taoliu@syau.edu.cn; qihongyan@syau.edu.cn

Abstract

Trehalose is a natural nonreducing disaccharide that is found in most organisms, such as yeasts, bacteria, invertebrates and plants. Trehalose plays an important role in regulating plant growth and development and stress response. Thus, in this review, we discuss the physical and chemical properties of trehalose, its function in plant growth and development, and the regulatory mechanism of its response to abiotic stresses such as drought, salt, and extreme temperature. The purpose of this review is to provide a reference for further analysis on the mechanism of trehalose in regulating plant growth and development and stress resistance.

Citation: Han Y, Liang A, Xu D, Zhang Y, Shi J, et al. 2024. Versatile roles of trehalose in plant growth and development and responses to abiotic stress. *Vegetable Research* 4: e007 <https://doi.org/10.48130/vegres-0024-0007>

Introduction

In nature, plants are faced with many challenges posed by adverse environments, such as drought, extreme temperature and salinity. To cope with these disadvantages, plants adapt to abiotic stress by accumulating compatible solutes, such as soluble sugars and some free amino acids, which is often considered to be the basic strategy for their protection and survival under stress^[1]. Among these compatible substances, most sugars not only play roles in osmotic regulation but also play a signaling role, such as glucose^[2–4], sucrose^[4–6] and trehalose-6-phosphate^[7–9]. Sugars are the basis of energy storage and material transport in plants. Different types of sugars are formed by metabolism after photosynthesis and play key roles in many metabolic processes throughout the whole life cycle of plants. In the process of plant growth and development and environmental response, sugars mostly act as signal molecules to regulate a variety of physiological and biochemical processes^[10]. Trehalose is a kind of nonreducing disaccharide with special physical and chemical properties that has strong hydration ability under drying and freezing conditions and can replace the bound water on the surface of biomolecules to improve the stability of proteins and biofilms^[11,12]. Trehalose is widely found in a variety of organisms, including bacteria, yeasts, fungi and algae, as well as some insects, invertebrates and plants^[13]. Trehalose is easily induced by stress, stimulates plant resistance mechanisms^[14], and plays an important role in dealing with a variety of abiotic stresses, such as drought stress^[15,16], salt stress^[15,17] and extreme temperature stress^[18,19]. This review discusses the advances of trehalose in regulating plant growth and development and response to abiotic stress.

Physicochemical properties of trehalose

Trehalose is composed of two glucose subunits linked by a 1,1-glycosidic bond^[20]. Since both reducing subunits are involved in the formation of glycosidic bonds (Fig. 1), trehalose has certain molecular stability and unique properties compared with other disaccharides^[21]. Trehalose can resist acid hydrolysis and maintain its stability under acidic pH and high temperature conditions^[22]. Trehalose cannot form hydrogen bonds inside it, which creates its high degree of hydrophilicity^[23]. When the organism is dehydrated or frozen, trehalose can replace water molecules to form hydrogen bonds with surrounding macromolecules and membranes, thus playing a protective role. In the case of extreme dehydration, trehalose can be crystallized into glass, avoiding the denaturation of biomolecules and restoring its functional activity during rehydration^[24].

Biosynthesis and decomposition of trehalose in plants

Trehalose in plants was first found in *Selaginella tamariscina* and then identified in algae, liverworts and other lower plants, while trehalose in angiosperms was found later^[25]. The biosynthesis of trehalose in plants mainly consists of the monosaccharides uridine-glucose diphosphate (UDP-Glc) and glucose-6-phosphate (Glc-6-P) as precursors (Fig. 2), catalyzed by trehalose-6-phosphate synthase (TPS) to form trehalose-6-phosphate (Trehalose-6-P, T6P). Then, trehalose is formed under the catalysis of trehalose-6-phosphate phosphatase (TPP)^[22,25]. Approximately 20 years ago, two enzymes of the trehalose biosynthesis pathway, trehalose-6-phosphate synthase (TPS) and trehalose-6-phosphate phosphatase (TPP), were identified

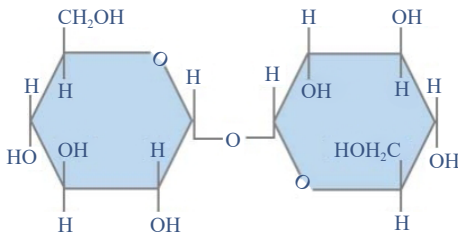


Fig. 1 Chemical structure of trehalose.

in *Arabidopsis thaliana*^[26,27]. In *Arabidopsis thaliana*, there are 11 genes encoding TPS or TPS-like protein (*AtTPS1-AtTPS11*) and 10 genes encoding TPP protein (*AtTPPA-AtTPPJ*)^[28,29]. A total of 14 *TPS* and 13 *TPP* genes were identified in rice, of which *OsTPP1* is regulated by abiotic stress^[30]. In cassava, there are 12 *TPS* and 10 *TPP* genes that encode proteins that play a key role in trehalose synthesis^[31]. Compared with the synthetic pathway, the decomposition of trehalose is simpler. Trehalose is directly hydrolyzed into two molecules of glucose under the action of trehalase (TRE), while glucose can be formed into

GLC-6-P under the action of hexokinase (HXK)^[10]. Trehalose in plants has only one biosynthesis and decomposition pathway, which plays an extensive role in plant growth and development and stress response^[29].

The function of trehalose in plant growth and development

Many studies have shown that trehalose and the key intermediate substances in its metabolism play crucial roles in regulating plant growth and development (Fig. 3). In plants, trehalose participates in various life and cell metabolism processes. For example, trehalose induced the expression of NADPH-dependent thioredoxin reductase C (NTRC) during the day and inhibited it at night, which affected the activity of ADP-glucose pyrophosphorylase (AGPase) and regulated the daily starch accumulation and metabolism of *Arabidopsis thaliana*^[32]. Trehalose and its biosynthetic gene can also participate in abscisic acid (ABA)-mediated root elongation, stomatal movement and seed germination. In rice, appropriate concentrations of trehalose and sucrose inhibited the synthesis of

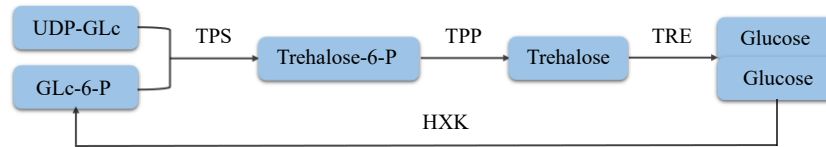


Fig. 2 Synthesis and degradation of trehalose.

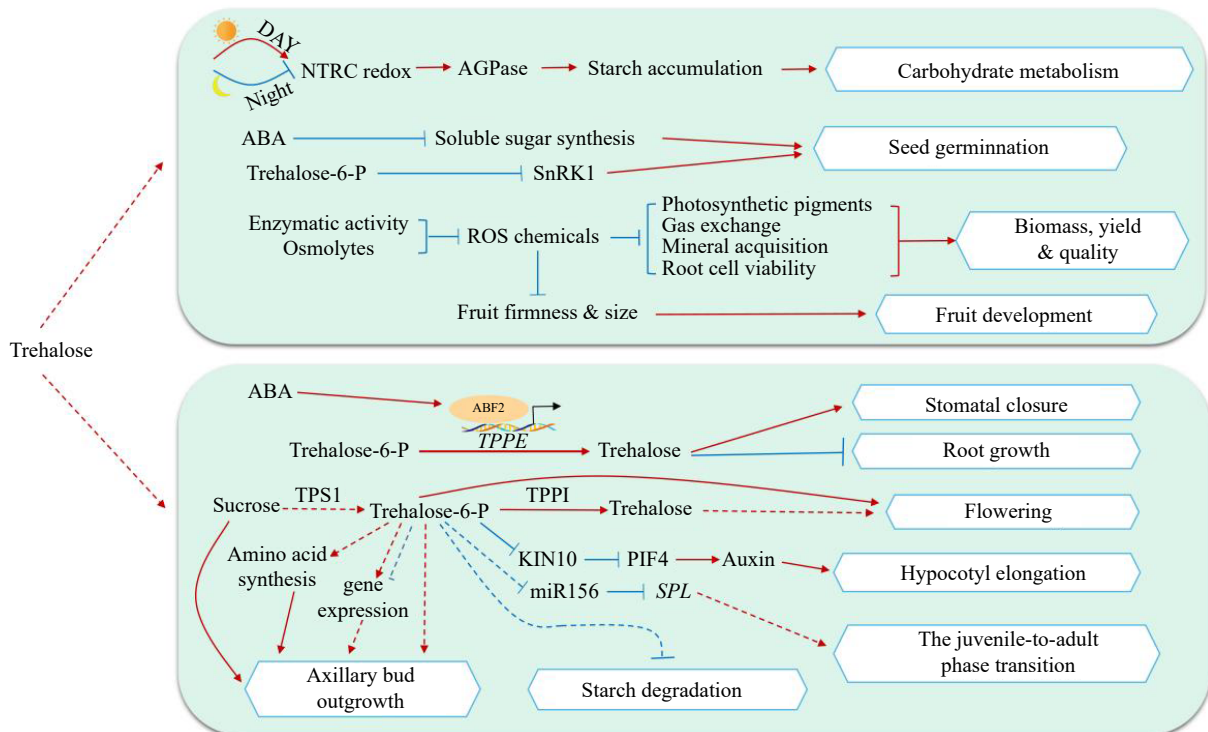


Fig. 3 Function of trehalose in plant growth and development. → Represent promoting; —| Means inhibiting; - - -> Represent promoting; - - -| Means inhibiting. However, they are uncertain and need to be verified. NTRC: NADPH-dependent thioredoxin reductase C; AGPase: ADP-glucose pyrophosphorylase; ABA: Abscisic acid; SnRK1: SNF1-related protein kinase 1; ROS: Reactive oxygen species; ABF2: ABA response element binding factor 2; TPPE: Trehalose-6-phosphate phosphatase E; TPPI: Trehalose-6-phosphate phosphatase I; TPS1: Trehalose-6-phosphate synthase 1; KIN10: A sugar signaling kinase; PIF4: Phytochrome interacting factor 4; miR156: microRNA 156; SPL: Squamosa promoter binding protein-like.

Versatile roles of trehalose in plants

soluble sugars and seed germination by promoting ABA synthesis^[33]. In another study with *Arabidopsis thaliana*, in the presence of ABA, ABF2 enhanced the expression of *AtTPPE* by directly binding to its promoter, increased trehalose content and triggered the accumulation of reactive oxygen species (ROS) in roots and stomata, thus enhancing the effect of ABA on inhibiting root growth and promoting stomatal closure^[34]. In addition, in legumes, *PvTPS9* can regulate the metabolism of trehalose in symbiotic root nodules and the whole plant, indicating the role of trehalose in affecting the growth of root nodules and plants^[35].

In recent years, key genes of trehalose biosynthesis have been reported to play key roles in plant flower development. Kataya et al. found that *AtTPPI* homozygous T-DNA insertion lines showed smaller leaves, shorter roots, delayed flowering and salt sensitivity^[36]. Meanwhile, deletion of *AtTPS1* led to late flowering, indicating that *AtTPS1* is necessary to regulate timely flowering^[37]. Zhao et al. transferred *JcTPPJ* into *Arabidopsis thaliana* and found that *35S:JcTPPJ* transgenic plants had higher sucrose content in inflorescences, showing late flowering and style atypia, indicating that *JcTPPJ* may regulate floral organ development by regulating sucrose status in plants^[38]. These three studies suggest the potential role of trehalose synthesis genes in regulating plant development. In the process of plant flowering, trehalose synthesis genes may also regulate the development of floral organs by affecting the content of other soluble sugars (such as sucrose). Additionally, in lotus, low light stress blocked photosynthesis, decreased the content of soluble sugar in the rhizome, decreased the expression of *NnTPS1*, activated the activity of NnSnRK1, induced programmed cell death and finally led to flower bud abortion^[39]. In Solanaceous crops, trehalose application affected style length *via* roots interaction with rhizosphere and promoted pistil to stamen ratio^[40]. There is little evidence that trehalose is directly involved in the regulation of plant flowering, but some reports have shown that trehalose synthesis genes are involved in flower organ development, which still needs further study.

Trehalose was also reported to play important roles in improving crop yield and quality. Islam & Mohammad reported that foliar spraying of 10 mM Tre significantly enhanced plant photosynthesis, mineral acquisition and root cell activity, ultimately improving the biomass, yield and quality of Indian mustard^[41]. In apple, potassium treatment significantly reduced the activity of trehalase (TRE) and increased trehalose content in fruit, which improved fruit firmness and quality^[42].

As the synthetic precursor of trehalose, the function of T6P in plant growth and development has been widely studied. T6P, as a signaling molecule, is related to plant growth and development and sucrose metabolism^[43]. In *Arabidopsis thaliana*, the change in sucrose directly affected the levels of T6P in plants, regulated the levels of PIF4 protein by regulating GRIK1-mediated KIN10 activation and finally affected auxin signaling to regulate hypocotyl elongation^[44]. Meanwhile, during the growth of *Arabidopsis thaliana*, sucrose activated the T6P pathway, which suppressed the level of mature miR156, leading to the upregulation of the *SPL* gene and regulating the plant development^[9]. In recent years, it has been reported that T6P plays a key role in the regulation of embryonic maturation^[7], axillary bud growth^[45,46] and leaf starch degradation^[47]. Whether the actions of T6P are related to trehalose, and

whether endogenous trehalose may act as a signal molecule related to T6P signal in regulating plant growth, development and stress response. These need to be further studied.

Trehalose function in plant abiotic stress

Trehalose protects biological cells and bioactive substances from destruction under adverse environmental conditions such as dehydration, drought, high temperature, freezing, high osmotic pressure and toxic reagents and plays an important role in plant resistance to abiotic stress^[48]. Overexpressing *OsTPS1* showed elevated trehalose and proline concentrations and upregulation of stress response-related genes, such as *WSI18*, *RAB16C*, *HSP70* and *ELIP*, resulting in enhanced resistance of rice seedlings to abiotic stresses^[15]. In sweet potato, *IbTPS1* is induced by drought, salt, heat and other environmental stresses. The tolerance of yeast to dehydration, salinity and oxidation was improved by expressing *IbTPS1* in yeast, indicating that *IbTPS1* is a candidate gene for improving the plant stress resistance^[49], which may be related to the increase of endogenous trehalose levels. Similarly, application of exogenous trehalose can also improve plant stress resistance^[50–53]. However, whether exogenous trehalose improves plant resistance by inducing endogenous trehalose or stimulating other signaling molecules in plants, thereby mobilizing the resistance system and improving resistance, still need further exploration.

Drought stress

Drought stress can destroy cellular ROS homeostasis, increase the accumulation of oxygen free radicals, inhibit plant growth, and damage the plant osmotic regulation system, biofilm system, respiration and photosynthesis metabolism^[54]. Han et al. found that cassava, a drought-tolerant crop, had a high expression of *MeTPS1* in tissues before and after stress, and the content of trehalose increased, which improved the tolerance to drought stress^[31], suggesting that trehalose plays a key role in plant resistance to drought stress.

Trehalose application was widely reported to improve the plants drought stress tolerance *via* reducing oxidative damage and restoring photosynthetic capacity (Table 1). For example, spraying exogenous trehalose increased the contents of antioxidants such as ascorbic acid (AsA) and reduced glutathione (GSH) in maize roots and leaves, increased the activities of antioxidant enzymes such as superoxide dismutase (SOD) and ascorbate peroxidase (APX), decreased the production rate of superoxide anion (O_2^-) and malondialdehyde (MDA), reduced the degree of membrane lipid peroxidation and resisted drought stress^[55], which similarly in wheat^[50], radish^[56], etc. These eventually alleviated the oxidative damage to chloroplasts and enhance plant photosynthetic capacity^[57,58].

In addition, exogenous trehalose could increase plant drought stress by inducing leaf anatomical changes, such as increasing leaf epidermis thickness, vascular bundle area, midvein thickness and number of vascular bundles^[59]. Also, trehalose can participate in seed germination under stress. Under drought conditions, 0.5 mmol·L⁻¹ trehalose increased starch degradation by upregulating the expression of the calcium-dependent *CBL1-OsSnRK3.1/3.23* gene and activating the *OsK1a-OsMYBS1/2-OsAmy3/8* pathway, and induced trehalose synthesis, thereby enhancing sugar metabolism, maintaining seed germination, significantly increasing drought tolerance during germination^[60].

Table 1. Roles of trehalose in regulating plant stress resistance.

Stress types	Regulatory mechanisms	Species	Treatments	References	
Drought stress	Enhance antioxidant capacity	<i>Zea mays</i>	0 and 30 mM Tre spraying plants	[16]	
		<i>Helianthus annuus</i>	0, 10, 20 and 30 mM Tre spraying plants	[101]	
		<i>Triticum aestivum</i>	Medium + 50 mM Tre	[50]	
		<i>Raphanus sativus</i>	0, 25 and 50 mM Tre soaking seeds and spraying plants	[56]	
		<i>Ocimum basilicum</i>	30 mM Tre and 1 mM SA alone or in combination with irrigating	[102]	
	Protect photosynthetic mechanism	<i>Chenopodium quinoa</i>	0, 5, 10, 15, 20 mmol-L ⁻¹ Tre spraying plants	[54]	
		<i>Zea mays</i>	10 mmol-L ⁻¹ Tre spraying plants	[55]	
		<i>Zea mays</i>	0 and 30 mM Tre spraying plants	[57]	
		<i>Zea mays</i>	1% Tre and different forms of zinc spraying plants	[58]	
		<i>Raphanus sativus</i>	25 mM Tre soaking seeds and spraying plants	[59]	
Salt stress	Cause anatomical changes of leaves	<i>Solanum lycopersicum</i>	1.5, 15 and 45 mM Tre spraying plants	[63]	
	Regulate endogenous ABA level and signal transduction				
		Enhance antioxidant capacity	<i>Oryza sativa</i>	25 mM Tre soaking seeds	[67]
			<i>Oryza sativa</i>	Nutrient solution + 10 mM Tre hydroponics	[68]
	<i>Zea mays</i>		Nutrient solution + 10 mM Tre hydroponics	[52]	
	Protect photosynthetic mechanism	<i>Arabidopsis thaliana</i>	Nutrient solution + 0.5, 1 and 5 mM Tre hydroponics	[51]	
		<i>Cucumis melo</i>	2%, 3%, 4%, 5% Tre spraying plants	[69]	
		<i>Citrullus lanatus</i>	Nutrient solution + 0, 5, 10, 20 and 30 mM Tre hydroponics	[70]	
		<i>Fragaria × ananassa</i>	Nutrient solution + 10, 30 mM Tre irrigating plants	[72]	
<i>Oryza sativa</i>		0, 10, 20 mM Tre spraying plants	[71]		
Heat stress	Co-regulation of stress response with other substances	<i>Oryza sativa</i>	Nutrient solution + 10 mmol-L ⁻¹ Tre hydroponics	[17]	
	Protect PSII and regulate plant photosynthesis	<i>Triticum aestivum</i>	Nutrient solution + 1.5 mM Tre hydroponics	[18,78,82]	
	Regulate plant redox dynamic balance and photosynthesis	<i>Paeonia lactiflora</i>	30 mmol-L ⁻¹ Tre spraying plants	[81]	
Cold stress	Enhance antioxidant capacity	<i>Capsicum annuum</i>	5%, 10%, 15% Tre soaking fruit	[88]	
		<i>Solanum lycopersicum</i>	10 mM Tre spraying plants	[86]	
	<i>Cucumis melo</i>	10 mM Tre spraying plants	[53,87]		
	Osmotic adjustment	<i>Zea mays</i>	3, 6, 9, 12, 15, 18 mmol-L ⁻¹ Tre irrigating plants	[85]	
		<i>Oryza sativa</i>	0, 0.5, 1 and 2 mM Tre/Spermidine soaking seeds	[19]	
Heavy metal stress	Regulate nitrogen assimilation and polyamine synthesis	<i>Oryza sativa</i>	Nutrient solution + 5 mM Tre irrigating plants	[90]	
		<i>Triticum aestivum</i>	0, 5, 10, 20, 40, 50 mmol-L ⁻¹ Tre soaking seeds and hydroponics	[89]	
	Enhance antioxidant capacity	<i>Triticum aestivum</i>	1, 10, 50 mmol-L ⁻¹ Tre spraying plants	[91]	
		<i>Oryza sativa</i>	Nutrient solution + 10 mM Tre hydroponics	[94]	
		<i>Oryza sativa</i>	0, 10, 20, 40, 60 mmol-L ⁻¹ Tre hydroponics	[95]	
Nitrogen deficiency	Activate nitrate and ammonia assimilation	<i>Triticum aestivum</i>	0, 25, 50 mM Tre spraying plants	[96]	
		<i>Nicotiana tabacum</i>	8 mM Tre spraying plants	[100]	
Acid rain stress	Enhance antioxidant capacity, maintain the stability of plasma membrane	<i>Hordeum vulgare</i>	0, 5, 10, 15 mM Tre soaking seeds	[98]	
Alkali stress	Enhance antioxidant capacity, osmotic adjustment	<i>Oryza sativa</i>	0, 5, 10, 15, 20 mmol-L ⁻¹ Tre spraying plants	[99]	

ABA is an important plant hormone that responds to various abiotic stresses, and its level increases rapidly under drought conditions^[61]. In recent years, the relationship between trehalose and ABA in the process of drought has been gradually clarified. In *Arabidopsis thaliana*, drought stress caused ABA signal response factors ABF1 and ABF4 to activate *AtTPPI* expression, changed the trehalose metabolism pathway, led to stomatal closure, improved water use efficiency, and made plants adapt to stress^[62]. Trehalose also can upregulate the expression of ABA signal-related genes *SIPYL1/3/4/5/6/7/9*, *SISnRK2.3/4*, *SIAREB1/2* and *SIDREB1*, activate the ABA signal

pathway and regulate stomatal closure and cell water loss under drought stress^[63]. Besides that, overexpression of trehalose synthetic gene *OsTPP3* also increased the ABA content and drought resistance of plants by increasing the expression of genes related to ABA biosynthesis^[64]. These suggest that trehalose could increase plant drought resistance by inducing ABA production.

Salt stress

Salt stress mainly includes osmotic stress and ion toxicity, redox disorder, which leads to nutrition deficits and disrupts

Versatile roles of trehalose in plants

the energy balance of plants^[65] and also affects water absorption and utilization, the anatomical structure of leaves and photosynthesis, eventually limited plant growth^[65,66]. Therefore, how to enhance plants salt stress tolerance is particularly important for plant normal growth and development.

Studies showed that feed with trehalose could trigger the expression of salt tolerance-related transcription factors genes, such as *bHLH*, *NAC*, *WRKY*, etc, and increase the level of endogenous trehalose in rice, thus enhancing the activity of antioxidant enzymes (Table 1)^[67–69], increasing K⁺ level and the ratio of K⁺/Na⁺ in leaves and stems, maintain ion dynamic balance and redox state^[51], regulating antioxidants and the glyoxylase system^[52], which synergistically improve the salt tolerance of plants^[70]. Foliar spraying trehalose can alleviate the adverse effects of salt stress on rice by improving growth traits, chlorophyll content, gas exchange characteristics, chlorophyll fluorescence and other parameters^[71]. In strawberries, external application of 30 mM trehalose significantly alleviated the inhibition of salt stress on strawberry growth. It could alleviate the inhibition of PSII function by increasing carotenoid content, thus reducing the injury caused by salt stress^[72]. Additionally, over-expressing of trehalose synthesis genes *AtTPPD* could improve plant salt stress resistance *via* regulating sugar metabolism^[73]. These suggest that trehalose could regulate sugar metabolism, which need further investigation. Over all, trehalose may improve plant salt stress tolerance and promote plant growth and development by regulating the balance of antioxidant system, ion exchange and other metabolism (such as sugar metabolism).

Additionally, trehalose can coordinate with other substances to regulate plant salt tolerance, such as osmoregulatory substances and growth-promoting bacteria. Nounjan & Theerakulpisut showed that external application of proline and trehalose could increase the activity of antioxidant enzymes in rice under salt stress or in the recovery stage, among which exogenous trehalose had the most obvious promoting effect on the activity of antioxidant enzymes in rice^[17]. In another study with tomato, 1-aminocyclopropane-1-carboxylate (ACC) deaminase and trehalose had synergistic protective effects on tomato plants under salt stress during interaction with the plant growth-promoting strain *Pseudomonas* sp. UW4^[74]. Strigolactones can upregulate the expression of the Tre biosynthetic genes *TPS1*, *TPS2*, *TPP1* and *TPP2*, enhance the activity of TPS and TPP, accelerate the conversion of glutamic acid to Tre, and inhibit the degradation of Tre by weakening the activity of trehalase, thereby improving tomato plant salt stress resistance^[75]. Under salt stress, the upregulation of *OsNCED3* leads to ABA accumulation, thereby activating the expression of *OsTPP3*, increasing the Tre content of rice seedlings, and enhancing their salt tolerance^[76].

Heat stress

In plants, high temperature often leads to the outbreak of ROS, which destroys proteins, DNA and lipids and leads to adverse changes in plant growth, development and physiological status^[77,78]. Osmotic protective agents, such as proline^[79,80] and trehalose (Table 1)^[81,82], play a positive role in alleviating heat stress. Trehalose can significantly promote the PSII complex to maintain a stable oxygen evolution rate and cell redox homeostasis, which eventually alleviated the damage caused by heat stress^[78,81,83]. Meanwhile, trehalose effectively

regulated the level of photosynthesis-related proteins, alleviated the chloroplasts structure damage and increased the proton gradient (ΔpH) and ATP synthase activity by promoting cyclic electron flow (CEF) to alleviate PSII photoinhibition caused by heat stress in wheat^[78,82]. Thus, trehalose can alleviate heat caused damage mainly by maintaining normal photosynthesis in plants. However, most studies are limited to physiological mechanisms, and the related molecular regulatory mechanisms need to be further studied.

Cold stress

Cold stress is one of the main abiotic stresses in a plants life, including chilling/cold injury (> 0 °C) and freezing injury (< 0 °C). Cold stress is the main environmental factor affecting plant growth and development, limiting geographical distribution and reducing crop yield. Trehalose can be widely detected in cold-tolerant crops, indicating that trehalose may be involved in the regulation of plant tolerance to cold stress (Table 1)^[84].

Cold stress can cause an imbalance of the antioxidant system and serious damage to the cell membrane structure of plants. Liu et al. reported that irrigation trehalose could increase the activities of antioxidant enzymes in maize roots, enhance the ability of cells to scavenge ROS, and maintain the stability of the cell membrane structure and function of maize seedling roots, thus alleviating the damage caused by cold stress^[85]. And exogenous trehalose may play roles in H₂O₂→NO→antioxidation→cold tolerance pathway^[86,87] and maintain the integrity of cell structure^[53]. In addition, trehalose could alleviate the chilling injury of pepper fruit at low temperature and maintain quality by reducing the damage to the cell membrane structure caused by ROS^[88].

Cold stress also leads to osmotic stress, and trehalose can improve plants resistance to stress by regulating some osmotic regulatory substances. For example, trehalose could increase the content of osmotic regulating substances (such as proline and soluble sugar) which may result in increasing plant cold tolerance^[19,89]. In addition, application of trehalose regulated the water absorption of arbuscular mycorrhizal fungi (AMF) and mycorrhizal symbiotic rice under cold stress by inducing the expression of *GintAQPF* and *OsPIPs*, thus creating more suitable growth conditions for rice^[90]. These results indicate that trehalose plays a regulatory role in osmotic stress induced by low temperatures.

Studies have also shown that trehalose can regulate plant resistance to cold stress in other ways. For example, trehalose could promote floret fertility to alleviate the decrease in the number of grains per spike caused by cold stress at the booting stage, mainly because trehalose regulates nitrogen assimilation, the GSH-AsA cycle and spermidine synthesis^[91]. Liu et al. transferred the *TaTPS11* gene of cold-tolerant wheat varieties into *Arabidopsis thaliana* and found that the overexpression line had higher carbohydrates, such as sucrose, fructose and starch, which improved the cold tolerance of *Arabidopsis thaliana*^[92]. In rice, 'SAPK10-ABF1-TPS2' participates in plant cold tolerance by regulating the homeostasis of trehalose^[93].

Other stresses

In addition to drought, salt and extreme temperature, plants often suffer from other stresses during their growth and development, such as heavy metals, acid rain, and nutrient deficiency. Studies have shown that trehalose can play a role when plants are poisoned by heavy metals (Table 1). When exposed

to Cu stress, trehalose application can increase the level of endogenous trehalose and regulate antioxidant and glyoxylase systems, thus improving rice tolerance to Cu stress^[94]. Additionally, exogenous trehalose could effectively alleviate the decrease in chlorophyll a and b contents in rice caused by Cd stress, reduce the excessive accumulation of ROS, and form Cd-Tre chelates to reduce Cd toxicity^[95], improve the growth, physiology, and defense systems, thereby increase the crop quality and yield^[96]. Elevating endogenous trehalose by transferring *AtTPS1* gene into tobacco showed the same roles in improving plant Cd and Cu stress tolerance^[97]. In barley, exogenous trehalose increased the activity of antioxidant enzymes and plasma membrane H⁺-ATPase and the content of chlorophyll to maintain the stable pH of the plasma membrane in roots and leaves, thus resisting acid rain stress^[98]. In rice, spraying an appropriate concentration of trehalose could effectively improve alkali tolerance and alleviate the damage of alkali stress to rice seedlings, which is mainly achieved by improving the ability of ROS scavenging and osmotic regulation^[99]. Foliar spraying trehalose could partly alleviate the symptoms of nitrogen deficiency in tobacco by upregulating nitrate and ammonia assimilation, increasing the activities of enzymes such as nitrate reductase (NR) and glycolate oxidase (GO), and changing the contents of NH₄⁺ and NO₃⁻^[100].

In general, when plants encounter abiotic stress, trehalose application can enhance the antioxidative ability to maintain ROS homeostasis, protect photosynthetic institutions, and regulate osmotic regulators in plants, such as endogenous

trehalose^[67] and proline^[19]. Moreover, the overexpression of two key enzyme (TPP and TPS)-related genes in the trehalose biosynthesis pathway can also alleviate the damage caused by abiotic stress by increasing the level of endogenous trehalose and regulating the changes in downstream stress-related genes^[15], plant hormones^[64] and sugar^[73].

Summary and prospects

Sugar plays a key role in many metabolic processes throughout plant life. Trehalose, as a multifunctional biomolecule, plays an important role in seed germination^[33], plant development and reproduction^[37] and yield and quality formation^[42]. Trehalose shows great potential in regulating plant growth and development and improving plant stress tolerance (Fig. 4). Although the preliminary physiological effects of trehalose on key plant physiological processes and stress have been elucidated, most of them are at the physiological level, and the related molecular mechanisms need to be further studied. For example, how trehalose is involved in the regulation of redox balance, photosynthesis and glucose metabolism and how to regulate downstream stress-related genes to improve plant resistance remain to be studied. Furthermore, the current reports on the interaction between trehalose and plant hormones are mainly focused on ABA. Whether there is a certain relationship between trehalose and other plant hormones (such as ethylene, auxin and gibberellin) in the regulation of plant growth, development and stress resistance. And whether

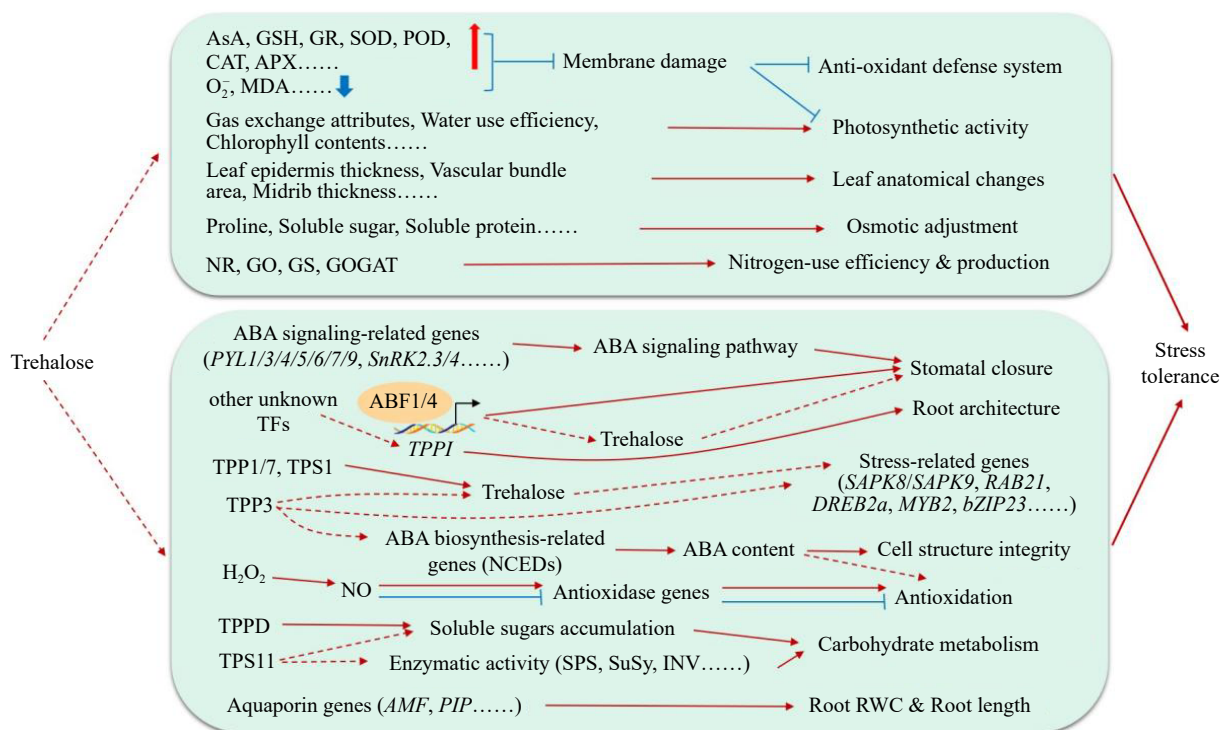


Fig. 4 Function of trehalose in the abiotic stress response of plants. —→ Represent promoting; —→ Means inhibiting; - - - -> Represent promoting; - - - -> Means inhibiting. However, they are uncertain and need to be verified. AsA: Ascorbic acid; GSH: Reduced glutathione ; GR: Glutathione reductase; SOD: Superoxide dismutase; POD: Peroxidase; CAT: Catalase; APX: Ascorbate peroxidase; MDA: Malondialdehyde; O₂⁻: Superoxide anion; NR: Nitrate reductase; GO: Glycolate oxidase; GS: Glutamine synthetase; GOCAT: Glutamine oxoglutarate aminotransferase; PYL: Pyrabactin resistance-like; SAPK: Stress activated protein kinase; RAB: Responsive to ABA; DREB: Dehydrationresponsive element-binding protein; MYB: MYB transcription factors; bZIP: bZIP transcription factors; H₂O₂: Hydrogen peroxide; NO: Nitric oxide; SPS: Sucrose phosphate synthase; SuSy: Sucrose synthase; INV: Invertase; AMF: Arbuscular mycorrhizal fungi; PIP: Plasma membrane intrinsic protein.

Versatile roles of trehalose in plants

endogenous trehalose also acts as a signal molecule and is related to the mechanism of action of exogenous trehalose needs further exploration.

Author contributions

The authors confirm contribution to the paper as follows: literature collection: Zhang Y, Xu D; tables and models design: Liang A; manuscript layout and format: Li M, Shi J; draft manuscript preparation: Han Y, Liu T; manuscript modification and review: Qi H. All authors reviewed the results and approved the final version of the manuscript.

Data availability

The data generated during and analyzed during the current study are available from the corresponding author upon reasonable request.

Acknowledgments

This work was supported by grants from the Basic Research Project of Liaoning Provincial Department of Education (JYTZD2023117), the China Agriculture Research System of MOF and MARA (CARS-25), the China Postdoctoral Science Foundation (2020M670793) and the and the Research Start Funding of Shenyang Agricultural University (880419015).

Conflict of interest

The authors declare that they have no conflict of interest.

Dates

Received 3 October 2023; Accepted 23 January 2024; Published online 27 February 2024

References

- Benina M, Obata T, Mehterov N, Ivanov I, Petrov V, et al. 2013. Comparative metabolic profiling of *Haberlea rhodopensis*, *Thellungiella halophylla*, and *Arabidopsis thaliana* exposed to low temperature. *Frontiers in Plant Science* 4:499
- Gupta A, Singh M, Laxmi A. 2015. Interaction between glucose and brassinosteroid during the regulation of lateral root development in *Arabidopsis*. *Plant Physiology* 168:307–20
- Miao L, Li Q, Sun T, Chai S, Wang C, et al. 2021. Sugars promote graft union development in the heterograft of cucumber onto pumpkin. *Horticulture Research* 8:146
- Meng L, Bao Q, Mu X, Tong C, Cao X, et al. 2021. Glucose- and sucrose-signaling modules regulate the *Arabidopsis* juvenile-to-adult phase transition. *Cell Reports* 36:109348
- Liu X, An X, Liu X, Hu D, Wang X, et al. 2017. MdSnRK1.1 interacts with MdJAZ18 to regulate sucrose-induced anthocyanin and proanthocyanidin accumulation in apple. *Journal of Experimental Botany* 68:2977–90
- Salam BB, Barbier F, Danieli R, Teper-Bamnlöcher P, Ziv C, et al. 2021. Sucrose promotes stem branching through cytokinin. *Plant Physiology* 185:1708–21
- Meitzel T, Radchuk R, McAdam EL, Thormählen I, Feil R, et al. 2021. Trehalose 6-phosphate promotes seed filling by activating auxin biosynthesis. *New Phytologist* 229:1553–65
- Paul MJ, Gonzalez-Urriarte A, Griffiths CA, Hassani-Pak K. 2018. The role of trehalose 6-phosphate in crop yield and resilience. *Plant Physiology* 177:12–23
- Ponnu J, Schlereth A, Zacharakis V, Dziado MA, Abel C, et al. 2020. The trehalose 6-phosphate pathway impacts vegetative phase change in *Arabidopsis thaliana*. *The Plant Journal* 104:768–80
- Chen T, Zhang Z, Li B, Qin G, Tian S. 2021. Molecular basis for optimizing sugar metabolism and transport during fruit development. *ABIOTECH* 2:330–40
- del C Luzardo M, Amalfá F, Nuñez AM, Díaz S, Biondi de Lopez AC, et al. 2000. Effect of trehalose and sucrose on the hydration and dipole potential of lipid bilayers. *Biophysical Journal* 78:2452–58
- Elbein AD, Pan Y, Pastuszak I, Carroll D. 2003. New insights on trehalose: a multifunctional molecule. *Glycobiology* 13:17R–27R
- Gong T, Li L, Zhao Z, Liu D. 2016. Advances in trehalose biosynthesis pathways and application of molecular biology technique. *Chinese Agricultural Science & Technology* 17:1790–95
- Fernandez O, Béthencourt L, Quero A, Sangwan RS, Clément C. 2010. Trehalose and plant stress responses: friend or foe? *Trends in Plant Science* 15:409–17
- Li H, Zang B, Deng X, Wang X. 2011. Overexpression of the trehalose-6-phosphate synthase gene *OsTPS1* enhances abiotic stress tolerance in rice. *Planta* 234:1007–18
- Ali Q, Ashraf M, Anwar F, Al-Qurainy F. 2012. Trehalose-induced changes in seed oil composition and antioxidant potential of maize grown under drought stress. *Journal of the American Oil Chemists' Society* 89:1485–93
- Nounjan N, Theerakulpisut P. 2012. Effects of exogenous proline and trehalose on physiological responses in rice seedlings during salt-stress and after recovery. *Plant, Soil & Environment* 58:309–15
- Luo Y, Liu H, Fan Y, Wang W, Zhao Y. 2018. Comparative chloroplast proteome analysis of exogenously supplied trehalose to wheat seedlings under heat stress. *Photosynthetica* 56:1123–33
- Fu Y, Zhang Z, Liu J, Chen M, Pan R, et al. 2020. Seed priming with spermidine and trehalose enhances chilling tolerance of rice via different mechanisms. *Journal of Plant Growth Regulation* 39:669–79
- Han J, He D, Zou C, Luo Y. 2021. Research progress on response mechanism of trehalose to abiotic stress in plants. *Biology Teaching* 46:2–4
- Jain NK, Roy I. 2010. Trehalose and protein stability. *Current Protocols in Protein Science* 59:4.9.1–4.9.12
- Kosar F, Akram NA, Sadiq M, Al-Qurainy F, Ashraf M. 2019. Trehalose: a key organic osmolyte effectively involved in plant abiotic stress tolerance. *Journal of Plant Growth Regulation* 38:606–18
- Paul S, Paul S. 2014. Trehalose induced modifications in the solvation pattern of *N*-methylacetamide. *The Journal of Physical Chemistry B* 118:1052–63
- Jia H, Li K, Luan Q, Wu J, Chen S. 2017. The application and protection mechanism of trehalose on biomaterials. *Fine and Specialty Chemicals* 25:51–53
- Wang W, Zhang Y, Yu M, Wang P. 2020. Research progress of trehalose in regulating plant response to abiotic stress. *Molecular Plant Breeding* 18:3433–40
- Blázquez MA, Santos E, Flores CL, Martínez-Zapater JM, Salinas J, et al. 1998. Isolation and molecular characterization of the *Arabidopsis* *TPS1* gene, encoding trehalose-6-phosphate synthase. *The Plant Journal* 13:685–89
- Vogel G, Aeschbacher RA, Müller J, Boller T, Wiemken A. 1998. Trehalose-6-phosphate phosphatases from *Arabidopsis thaliana*: identification by functional complementation of the yeast *tps2* mutant. *The Plant Journal* 13:673–83
- Vandesteene L, López-Galvis L, Vanneste K, Feil R, Maere S, et al. 2012. Expansive evolution of the trehalose-6-phosphate phosphatase gene family in *Arabidopsis*. *Plant Physiology* 160:884–96
- Lunn JE, Delorge I, Figueroa CM, Van Dijck P, Stitt M. 2014. Trehalose metabolism in plants. *The Plant Journal* 79:544–67
- Ge L, Chao D, Shi M, Zhu M, Gao J, et al. 2008. Overexpression of the trehalose-6-phosphate phosphatase gene *OsTPP1* confers stress tolerance in rice and results in the activation of stress responsive genes. *Planta* 228:191–201

31. Han B, Fu L, Zhang D, He X, Chen Q, et al. 2016. Interspecies and intraspecies analysis of trehalose contents and the biosynthesis pathway gene family reveals crucial roles of trehalose in osmotic-stress tolerance in cassava. *International Journal of Molecular Sciences* 17:1077
32. Noroozipoor A, Aghdasi M, Sadeghipour HR. 2020. Differential carbohydrate dynamics in *Arabidopsis* wild-type and *ntrc* mutant after trehalose feeding. *Acta Physiologiae Plantarum* 42:78
33. Zhao Y, Teng Z, Yu H, Wang Y, Ye N. 2023. Trehalose and sucrose inhibit rice seed germination by promoting the biosynthesis of abscisic acid. *Molecular Plant Breeding* 21:2693–702
34. Wang W, Chen Q, Xu S, Liu W, Zhu X, et al. 2020. Trehalose-6-phosphate phosphatase E modulates ABA-controlled root growth and stomatal movement in *Arabidopsis*. *Journal of Integrative Plant Biology* 62:1518–34
35. Barraza A, Contreras-Cubas C, Estrada-Navarrete G, Reyes JL, Juárez-Verdayes MA, et al. 2016. The class II trehalose 6-phosphate synthase gene *PvTPS9* modulates trehalose metabolism in *Phaseolus vulgaris* nodules. *Frontiers in Plant Science* 7:1589
36. Kataya ARA, Elshobaky A, Heidari B, Dugassa NF, Thelen JJ, et al. 2020. Multi-targeted trehalose-6-phosphate phosphatase I harbors a novel peroxisomal targeting signal 1 and is essential for flowering and development. *Planta* 251:98
37. Wahl V, Ponnuraj J, Schlereth A, Arrivault S, Langenecker T, et al. 2013. Regulation of flowering by trehalose-6-phosphate signaling in *Arabidopsis thaliana*. *Science* 339:704–07
38. Zhao M, Ni J, Chen M, Xu Z. 2019. Ectopic expression of *Jatropha curcas* *TREHALOSE-6-PHOSPHATE PHOSPHATASE J* causes late-flowering and heterostylous phenotypes in *Arabidopsis* but not in *Jatropha*. *International Journal of Molecular Sciences* 20:2165
39. Zhang X. 2019. *Mechanism analysis of lotus flower bud abortion under weak light mediated by trehalose-6-phosphate-synthase*. Dissertation. Nanjing Agricultural University, China
40. Mori IC, Matsuura T, Otao M, Ooi L, Nishimura Y, et al. 2023. Application of trehalose mitigates short-styled flowers in Solanaceous crops. *Journal of Agricultural & Food Chemistry* 71:5476–82
41. Islam S, Mohammad F. 2021. Modulation of growth, photosynthetic efficiency, leaf biochemistry, cell viability and yield of Indian mustard by the application of trehalose. *Scientia Horticulturae* 290:110527
42. Zhang W, Zhang N, Zhao J, Guo Y, Zhao Z, et al. 2017. Potassium fertilization improves apple fruit (*Malus domestica* Borkh. Cv. Fuji) development by regulating trehalose metabolism. *The Journal of Horticultural Science & Biotechnology* 92:539–49
43. Fichtner F, Lunn JE. 2021. The role of trehalose 6-phosphate (Tre6P) in plant metabolism and development. *Annual Review of Plant Biology* 72:737–60
44. Hwang G, Kim S, Cho JY, Paik I, Kim JI, et al. 2019. Trehalose-6-phosphate signaling regulates thermoresponsive hypocotyl growth in *Arabidopsis thaliana*. *EMBO Reports* 20:e47828
45. Fichtner F, Barbier FF, Feil R, Watanabe M, Annunziata MG, et al. 2017. Trehalose 6-phosphate is involved in triggering axillary bud outgrowth in garden pea (*Pisum sativum* L.). *The Plant Journal* 92:611–23
46. Fichtner F, Barbier FF, Annunziata MG, Feil R, Olas JJ, et al. 2021. Regulation of shoot branching in *Arabidopsis* by trehalose 6-phosphate. *New Phytologist* 229:2135–51
47. Martins MCM, Hejazi M, Fettek J, Steup M, Feil R, et al. 2013. Feedback inhibition of starch degradation in *Arabidopsis* leaves mediated by trehalose 6-phosphate. *Plant Physiology* 163:1142–63
48. Feng D. 1999. Brief introduction of biological function of trehalose. *Bulletin of Biology* 34:13–14
49. Wang W, Yu H, Kim HS, Yang Y, Qiu X, et al. 2019. Molecular characterization of a sweet potato stress tolerance-associated trehalose-6-phosphate synthase 1 gene (*IbTPS1*) in response to abiotic stress. *Plant Biotechnology Reports* 13:235–43
50. Ma C, Wang Z, Kong B, Lin T. 2013. Exogenous trehalose differentially modulate antioxidant defense system in wheat callus during water deficit and subsequent recovery. *Plant Growth Regulation* 70:275–85
51. Yang L, Zhao X, Zhu H, Paul M, Zu Y, et al. 2014. Exogenous trehalose largely alleviates ionic imbalance, ROS burst, and PCD occurrence induced by high salinity in *Arabidopsis* seedlings. *Frontiers in Plant Science* 5:570
52. Rohman MM, Islam MR, Monsur MB, Amiruzzaman M, Fujita M, et al. 2019. Trehalose protects maize plants from salt stress and phosphorus deficiency. *Plants* 8:568
53. Liu T, Han Y, Shi J, Liang A, Xu D, et al. 2022. Abscisic acid involved in trehalose improved melon photosynthesis via regulating oxidative stress tolerance and cell morphology structure under cold stress. *Environmental and Experimental Botany* 202:105042
54. Hao X, Wang X, Liu K. 2021. Effects of exogenous trehalose on the physiological characteristics of quinoa under drought stress. *Journal of Shandong Agricultural University (Natural Science Edition)* 52:739–45
55. Ye Y, Lu D, Wang F, Chen X, Qi M, et al. 2020. Effects of exogenous trehalose on physiological characteristics in waxy maize seedlings under drought stress. *Journal of Maize Sciences* 28:80–86
56. Shafiq S, Akram NA, Ashraf M. 2015. Does exogenously-applied trehalose alter oxidative defense system in the edible part of radish (*Raphanus sativus* L.) under water-deficit conditions? *Scientia Horticulturae* 185:68–75
57. Ali Q, Ashraf M. 2011. Induction of drought tolerance in maize (*Zea mays* L.) due to exogenous application of trehalose: growth, photosynthesis, water relations and oxidative defence mechanism. *Journal of Agronomy and Crop Science* 197:258–71
58. Klofac D, Antosovsky J, Skarpa P. 2023. Effect of zinc foliar fertilization alone and combined with trehalose on maize (*Zea mays* L.) growth under the drought. *Plants* 12:2539
59. Akram NA, Shafiq S, Ashraf M, Aisha R, Sajid MA. 2016. Drought-induced anatomical changes in radish (*Raphanus sativus* L.) leaves supplied with trehalose through different modes. *Arid Land Research and Management* 30:412–20
60. Li J, Xie Y, Li X, Wang J. 2021. Effects of trehalose on seed germination and drought tolerance of *C₄-PEPC* transgenic rice. *Journal of Nuclear Agricultural Sciences* 35:2879–92
61. Kuromori T, Seo M, Shinozaki K. 2018. ABA transport and plant water stress responses. *Trends in Plant Science* 23:513–22
62. Lin Q, Wang S, Dao Y, Wang J, Wang K. 2020. *Arabidopsis thaliana* trehalose-6-phosphate phosphatase *OsTPP3* increases drought tolerance by regulating stomatal apertures. *Journal of Experimental Botany* 71:4285–97
63. Yu W, Zhao R, Wang L, Zhang S, Li R, et al. 2019. ABA signaling rather than ABA metabolism is involved in trehalose-induced drought tolerance in tomato plants. *Planta* 250:643–55
64. Jiang D, Chen W, Gao J, Yang F, Zhuang C. 2019. Overexpression of the trehalose-6-phosphate phosphatase *OsTPP3* increases drought tolerance in rice. *Plant Biotechnology Reports* 13:285–92
65. Wang H. 2011. Exogenous trehalose improve *Arabidopsis thaliana* salt tolerance. Dissertation. Northeast Forestry University, China.
66. Zhao C, Zhang H, Song C, Zhu J, Shabala S. 2020. Mechanisms of plant responses and adaptation to soil salinity. *The Innovation* 1:100017
67. Abdallah MMS, Abdelgawad ZA, El-Bassiouny HMS. 2016. Alleviation of the adverse effects of salinity stress using trehalose in two rice varieties. *South African Journal of Botany* 103:275–82
68. Mostofa MG, Hossain MA, Fujita M. 2015. Trehalose pretreatment induces salt tolerance in rice (*Oryza sativa* L.) seedlings: oxidative damage and co-induction of antioxidant defense and glyoxalase systems. *Protoplasma* 252:461–75

Versatile roles of trehalose in plants

69. Xu T, Zhou C, Zhou C, Zhao S, Wu L, et al. 2014. Effects of trehalose on antioxidant system of melon seedlings under salt stress. *Northern Horticulture* 19:28–30
70. Yuan G, Sun D, An G, Li W, Si W, et al. 2022. Transcriptomic and metabolomic analysis of the effects of exogenous trehalose on salt tolerance in watermelon (*Citrullus lanatus*). *Cells* 11:2338
71. Shahbaz M, Abid A, Masood A, Waraich EA. 2017. Foliar-applied trehalose modulates growth, mineral nutrition, photosynthetic ability, and oxidative defense system of rice (*Oryza sativa* L.) under saline stress. *Journal of Plant Nutrition* 40:584–99
72. Samadi S, Habibi G, Vaziri A. 2019. Exogenous trehalose alleviates the inhibitory effects of salt stress in strawberry plants. *Acta Physiologiae Plantarum* 41:112
73. Krasensky J, Broyart C, Rabanal FA, Jonak C. 2014. The redox-sensitive chloroplast trehalose-6-phosphate phosphatase AtTPPD regulates salt stress tolerance. *Antioxidants & Redox Signaling* 21:1289–304
74. del Carmen Orozco-Mosqueda M, Duan J, DiBernardo M, Zetter E, Campos-García J, et al. 2019. The production of ACC deaminase and trehalose by the plant growth promoting bacterium *Pseudomonas* sp. UW4 synergistically protect tomato plants against salt stress. *Frontiers in Microbiology* 10:1392
75. Lu X, Liu X, Xu J, Liu Y, Chi Y, et al. 2023. Strigolactone-mediated trehalose enhances salt resistance in tomato seedlings. *Horticulturae* 9:770
76. Ye N, Wang Y, Yu H, Qin Z, Zhang J, et al. 2023. Abscisic acid enhances trehalose content via *OsTPP3* to improve salt tolerance in rice seedlings. *Plants* 12:2665
77. Wahid A, Gelani S, Ashraf M, Foolad MR. 2007. Heat tolerance in plants: an overview. *Environmental and Experimental Botany* 61:199–223
78. Luo Y, Wang W, Fan Y, Gao Y, Wang D. 2018. Exogenously-supplied trehalose provides better protection for D1 protein in winter wheat under heat stress. *Russian Journal of Plant Physiology* 65:115–22
79. Oukarroum A, Madidi SE, Strasser RJ. 2012. Exogenous glycine betaine and proline play a protective role in heat-stressed barley leaves (*Hordeum vulgare* L.): a chlorophyll *a* fluorescence study. *Plant Biosystems* 146:1037–43
80. Hussain R, Ayyub CM, Shaheen MR, Rashid S, Nafees M, et al. 2021. Regulation of osmotic balance and increased antioxidant activities under heat stress in *Abelmoschus esculentus* L. triggered by exogenous proline application. *Agronomy* 11:685
81. Zhao D, Li T, Hao Z, Cheng M, Tao J. 2019. Exogenous trehalose confers high temperature stress tolerance to herbaceous peony by enhancing antioxidant systems, activating photosynthesis, and protecting cell structure. *Cell Stress and Chaperones* 24:247–57
82. Luo Y, Xie Y, He D, Wang W, Yuan S. 2021. Exogenous trehalose protects photosystem II by promoting cyclic electron flow under heat and drought stresses in winter wheat. *Plant Biology* 23:770–76
83. Mamedov MD, Petrova IO, Yanykin DV, Zaspaa AA, Semenov AY. 2015. Effect of trehalose on oxygen evolution and electron transfer in photosystem 2 complexes. *Biochemistry Moscow* 80:61–66
84. Williams B, Njaci I, Moghaddam L, Long H, Dickman MB, et al. 2015. Trehalose accumulation triggers autophagy during plant desiccation. *PLoS Genetics* 11:e1005705
85. Liu X, Tong H, Tian L, Zuo S, Sun L, et al. 2018. Effects of exogenous trehalose on growth and physiological characteristics of maize seedling roots under chilling stress. *Chinese Journal of Agrometeorology* 39:538–47
86. Liu T, Ye X, Li M, Li J, Qi H, et al. 2020. H₂O₂ and NO are involved in trehalose-regulated oxidative stress tolerance in cold-stressed tomato plants. *Environmental and Experimental Botany* 171:103961
87. Liu T, Shi J, Li M, Ye X, Qi H. 2021. Trehalose triggers hydrogen peroxide and nitric oxide to participate in melon seedlings oxidative stress tolerance under cold stress. *Environmental and Experimental Botany* 184:104379
88. Ding F, Wang R. 2018. Amelioration of postharvest chilling stress by trehalose in pepper. *Scientia Horticulturae* 232:52–56
89. Xie D, Wang X, Fu L, Sun J, Li Z, et al. 2015. Effect of exogenous trehalose on germ length and seedling freeze resistance of winter wheat under cold stress. *Journal of Triticeae Crops* 35:215–23
90. Liu Z, Ma L, He X, Tian C. 2014. Water strategy of mycorrhizal rice at low temperature through the regulation of PIP aquaporins with the involvement of trehalose. *Applied Soil Ecology* 84:185–91
91. Liang Z, Luo J, Wei B, Liao Y, Liu Y. 2021. Trehalose can alleviate decreases in grain number per spike caused by low-temperature stress at the booting stage by promoting floret fertility in wheat. *Journal of Agronomy and Crop Science* 207:717–32
92. Liu X, Fu L, Qin P, Sun Y, Liu J, et al. 2019. Overexpression of the wheat trehalose 6-phosphate synthase 11 gene enhances cold tolerance in *Arabidopsis thaliana*. *Gene* 710:210–17
93. Shu Y, Zhang W, Tang L, Li Z, Liu X, et al. 2023. ABF1 positively regulates rice chilling tolerance via inducing trehalose biosynthesis. *International Journal of Molecular Sciences* 24:11082
94. Mostofa MG, Hossain MA, Fujita M, Tran LSP. 2015. Physiological and biochemical mechanisms associated with trehalose-induced copper-stress tolerance in rice. *Scientific Reports* 5:11433
95. Wang K, Li F, Gao M, Huang Y, Song Z. 2020. Mechanisms of trehalose-mediated mitigation of Cd toxicity in rice seedlings. *Journal of Cleaner Production* 267:121982
96. Alharby HF, Al-Zahrani HS, Hakeem KR, Ali S. 2023. Exogenous application of trehalose minimizes cadmium toxicity and alleviates oxidative stress in wheat under cadmium stress. *Turkish Journal of Agriculture and Forestry* 47:364–77
97. Martins LL, Mourato MP, Baptista S, Reis R, Carvalheiro F, et al. 2014. Response to oxidative stress induced by cadmium and copper in tobacco plants (*Nicotiana tabacum*) engineered with the trehalose-6-phosphate synthase gene (AtTPS1). *Acta Physiologiae Plantarum* 36:755–65
98. Ding F, Wang R, Wang T. 2018. Enhancement of germination, seedling growth, and oxidative metabolism of barley under simulated acid rain stress by exogenous trehalose. *Crop Science* 58:783–91
99. Zou D, Wang S, Sun J, Li J, Yin T, et al. 2020. Effect of exogenous trehalose on seedling growth and physiological characteristics of different rice varieties under alkali stress. *Journal of Northeast Agricultural University* 51:1–9
100. Lin Y, Zhang J, Gao W, Chen Y, Li H, et al. 2017. Exogenous trehalose improves growth under limiting nitrogen through upregulation of nitrogen metabolism. *BMC Plant Biology* 17:247
101. Kosar F, Akram NA, Ashraf M, Ahmad A, Alyemeni MN, et al. 2021. Impact of exogenously applied trehalose on leaf biochemistry, achene yield and oil composition of sunflower under drought stress. *Physiologia plantarum* 172:317–33
102. Zulfikar F, Chen J, Finnegan PM, Younis A, Nafees M, et al. 2021. Application of trehalose and salicylic acid mitigates drought stress in sweet basil and improves plant growth. *Plants* 10:1078



Copyright: © 2024 by the author(s). Published by Maximum Academic Press, Fayetteville, GA. This article is an open access article distributed under Creative Commons Attribution License (CC BY 4.0), visit <https://creativecommons.org/licenses/by/4.0/>.